

**ANTIBACTERIAL ACTIVITY OF TWIGS OF THUJA OCCIDENTALIS****Pooja Tiwari\* and Deepak Rathore**

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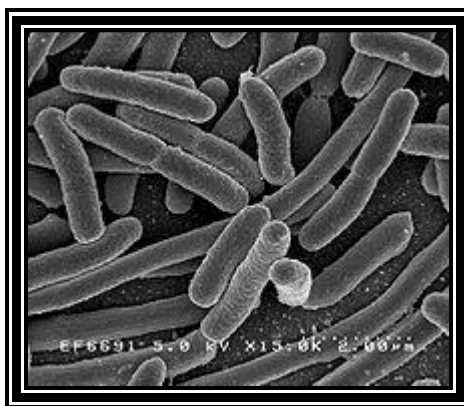
**ABSTRACT**

**Objective:** To evaluate the antibacterial activity of twigs of thuja occidentalis. **Methods:** Twigs extract of thuja occidentalis was prepared by the process of Soxhlet apparatus. **Results:** The plant Thuja occidentalis is an indigenous herb which was chosen for the present investigation study. The plant belongs to the family cupressaceae. The scanty availability of information on this plant facilitates the study on it. This attempt was made to study the pharmacognostical, phytochemical and antibacterial activities of plant. The study was divided into three major parts viz. Pharmacognostical studies, Phytochemical screening, Antibacterial activities. **Conclusion:** It was concluded that the Thuja occidentalis is antibacterial drug and shows good antibacterial activity against 'Gram Positive' bacteria and 'Gram Negative' bacteria.

**KEYWORDS:** antibacterial, Thuja occidentalis, extraction.

**INTRODUCTION**

Thuja occidentalis to family cupressaceae medicinally important plant, commonly grown in some parts of our country and used in the treatment of various disease and disorders of human ailments by tribal and rural people of our country. So, far no any systematic work was carried out to investigate the anti-bacterial activity of twigs of the selected plant therefore, the plant was selected for present investigation and Bacteria are a single-celled organism which can only be seen through microscope.<sup>[1,2]</sup> Bacteria come in different shapes and the size of bacteria is measured in micro-meter (which is a millionth part of a meter). Bacteria are found everywhere and in all type of environments. Bacteria are grouped as 'Gram Positive' bacteria and 'Gram Negative' bacteria, which is based on the results of Gram Staining.<sup>[3,4]</sup>



**Structure of Bacteria figure No.1**

## **MATERIAL AND METHOD**

### **Material**

The twig of the selected plant was collected in the months of august from the botanical gardens of Ujjain district of Madhya Pradesh and all the chemicals and reagents including drug were received from UIPS College Ujjain.

### **Methods**

**Selection of plant:** *Thuja occidentalis* to family cupressaceae medicinally important plant, commonly grown in some parts of our country and used in the treatment of various disease and disorders of human ailments by tribal and rural people of our country. So, far no any systematic work was carried out to investigate the anti-bacterial activity of twigs of the selected plant therefore, the plant was selected for present investigation.

**Macroscopic characters:** Various morphological studies of the twig were studied and result was reported in table

- 1) Color
- 2) Odour
- 3) Taste
- 4) Shape
- 5) Size

### **Physico-chemical evaluation**

The dried twigs of *thuja occidentalis* were subjected to standard procedure for the determination of various physicochemical parameters.

### Determination of ash values

The determination of ash values is meant for detecting low-grade products, exhausted drugs and sandy or earthy matter. It can also be utilized as a mean of detecting the chemical constituents by making use of water-soluble ash and acid insoluble ash.

### Extraction of Plant Material

The extraction of plant material (twigs) was carried by soxhlet apparatus. 25 gm of dried twig powder was taken and dissolved in 100 ml of water and ethanol and petroleum ether. Remained for 24 hours and filtered, concentrated, dried and then filtered.

### Preliminary Phytochemical screening

The aqueous extract obtained after extraction of twig was subjected to various Phytochemical screening as per the standard procedure to reveal various active phytoconstituents.

#### 1. Tests for fixed oils and fats

- **Spot test:** A small quantity of extract solution was separately pressed between two filter papers. Appearance of oil stain on the paper indicates the presence of fixed oil.

#### 2. Tests for steroids and triterpenoids

- **Libermann-burchard test:** The extract solution was treated with few drops of acetic anhydride, boiled and cooled. Then conc. Sulphuric acid was added from the side of test tube.
- **Salkowski test:** The extract solution was treated with few drop of conc. sulphuric acid,

**3. Test for proteins and free amino acids:** A small quantity of the extract solution was dissolved in few ml of water and treated with following reagents.

- **Million's reagent:** Small quantity of extract solution was taken, added few drops of millions reagent.
- **Ninhydrin reagent:** Small quantity of extract solution was taken, added few drops of ninhydrin reagent (0.1% solution in butanol).
- **Biuret's test:** Small quantity of extract solution was taken, added 5% of sodium hydroxide and 1% of copper.

**4. Test for tannins.**

- **Ferric chloride solution:** Treated the extract solution with ferric chloride solution.
- **Lead acetate solution:** Treated the extract solution with 10% lead acetate solution white precipitate was obtained.

**5. Test for Flavonoides.**

- **Alkaline reagent test**

To the extract solution added few drops of magnesium hydroxide solution.

- **Shinoda test**

To the extract solution added few magnesium turnings and concentrated hydrochloride drop wise pink.

**6. Test for mucilage's and gums:** Small quantities of extract solution was added separately to 25 ml of absolute alcohol with constant stirring and filtered

**7. Test for waxes:** To the extract solution added alcoholic alkali solution, wax got saponified.

**8. Test for Alkaloids**

- **Dragendorff's reagent:-**To the extract solution added few drops of Dragendorff's reagent (potassium bismuth iodide solution).
- **Mayer's reagent:** - To the extract solution added few drops of Mayer's reagent (potassium mercuric iodide solution)
- **Wagner's reagent:** - To the extract solution added few drops of Wagner's reagent (iodine- potassium iodide solution)
- **Hager's reagent:** - To the extract solution added few drops of Hager's reagent (saturated solution of picric acid)

**9. Test for Carbohydrates**

- **Molisch's test:-**To the extract solution added few drops of  $\alpha$ -naphthol, and then added few drops of sulphuric acid through the side of test tube.

**10. Test for Glycosides:** To the extract solution added the solution of  $\alpha$ -naphthol and sulphuric acid.

**11. Test for Starch:** To the aqueous extract added weak aqueous iodide.

**1) Procedure for Antibacterial Activity: Collection of bacteria:** - Two bacterial strains i.e. *Escherichia-coli* (gram –ve) and *Pseudomonas* (gram –ve) was collected from Ujjain institute of pharmaceutical and sciences.

**2) Drug Entrapped disc were Prepared:** - Whatman filter paper was pieces into small disc one quarter inch diameter. All dilutions were applied to autoclaved filter paper disc using micro pipette with sterile pipette tip.

**3) Culture media plate was prepared**

**Table 1: Nutrient agar media for bacteria.**

S. No.	Ingredients	Quantity Prescribed	Quantity Taken
1	Beef Extract	10 gm	2.5 gm
2	Peptone	10 gm	2.5 gm
3	Sodium chloride	5.0 gm	1.25 gm
4	Agar	20 gm	5 gm
5	Distilled Water	1000 ml	250 ml

• **Procedure for bacteria media**

1. 5 gm of agar was dissolved in 250 ml of water.
2. Then beef extract, peptone and sodium chloride was added in above agar solution with continuous stirring.
3. The media was heated to dissolve the agar to form the clear liquid.
4. Then pH was maintained at 7.2-7.4 by pH meter.
5. Sterilized by autoclave at 115°C at 15 lb pressure for 15 minutes.

**4) Culture for bacteria:** - The identified organism was applied to media for sensitivity test by streaking of medium with help of swab.

**5) Application of disc:** - All the disc of sample, standard, control with different dilution was placed on the inoculation plate with the help of flame sterilized forceps. After application of disc lead of petri-plate was closed, petri-plate was inoculated at 37°C for 24 hours for bacteria.

**6) Procedure for dilution**

- 1) 100 mg drug was dissolved in 100 ml of distilled water to prepared stock solution (1000µg /ml).

- 2) From the above solution 10 ml was taken in volumetric flask and diluted upto 100 ml to prepared sub-stock solution (100µg/ml).
- 3) From the above solution the adequate concentration 20, 40, 60 and 80 µg/ml was prepared and in the same way standard drug concentration 60µg/ml was prepared.

7) **Zone Of Inhibition:** - After incubation the plate were inspected to identify zone of inhibition. The diameter of zone of inhibition of each compound and the diameter of disc was recorded.

The result of zone of inhibition was reported in table 10. The structure of Initial Anti bacterial plate was reported in fig.26 and structure of anti fungal growth was reported in fig.27.

Zone of inhibition= Diameter of sample/ slanted/ control - diameter of disc

Zone of inhibition= Diameter of sample/ slanted/ control – 0.5 mm

## RESULTS AND DISCUSSION

The plant *Thuja occidentalis* is an indigenous herb which was chosen for the present investigation study. The plant belongs to the family *Cupressaceae*. The scanty availability of information on this plant facilitates the study on it. This attempt was made to study the pharmacognostical, phytochemical and antibacterial activities of plant. The study was divided into three major parts viz.

- Pharmacognostical studies
- Phytochemical screening
- Antibacterial activities

### Pharmacognostic studies

**Macroscopy:** The morphological features of the leaves of the selected plant *Thuja occidentalis* was presented below

**Table 2: Macroscopic Characters.**

S. No.	Character	Appearance
1	Color	Green
2	Odour	Strongly camphoraceous
3	Taste	Strongly camphoraceous
4	Shape	Lanceolate
5	Size	3mm long
7	Apex	Acute
8	Margin of Lamina	Entire Margin
9	Venation	Parallel Venation

10	Mid Rib	Present
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**Physico-chemical evaluation:** - The physicochemical analysis of leaves powder of *Thuja occidentalis* was carried out. In this study ash values (total ash, acid insoluble ash and water soluble ash) were determined, Swelling index, LOD, Foaming index, foreign organic matter (F.O.M.) was determined and the results are given below:

**Table 3: Physico-chemical evaluation.**

S. No.	Physicochemical Parameters	Result (w/v)
1	Ash Values	5.6%
1.1	Total Ash Value	4.8%
1.2	Acid Insoluble Ash Value	2.52%
1.3	Water Soluble ash Value	0.5%
2	Moisture Content (Loss On Drying)	0.9%

**Table 4: Foreign organic matter.**

S. No.	Drug	Weight of drug before	Weight of drug after	% of FOM
1	Twigs	100gm	96.2gm	4.2%

**Table.5. Swelling Index.**

S. No.	Drug	Drug	Inference
1	Before (cm)	After(cm)	
2	50cm	58.1cm	8.1

### Phytochemical Screening

The various extract of the plant of *Thuja occidentalis* were subjected to phytochemical screening which reveal the presence of various pharmacological active components. Phytochemical Screening of sample *Thuja occidentalis* was done as follows.

**Table 6: Preliminary Phytochemical Screening.**

S. No.	Tests	Aqueous extract
1	Fixed oil and fats Spot test	-
2	Tannins Ferric chloride test Alkaline reagent	+ -
3	Proteins Millions Reagent Ninhydrin Reagent Biuret Test	+ + + -
4	Flavonoides Alkaline reagent test Shinoda test	+ +
5	Steroids and triterpenoids Lieberman burchard test Salkowski test	- +
6	Mucilage and gum Reaction with 90% alcohol	-

7	Waxes Reaction with alcoholic KOH	-
8	Alkaloids Dragendorff's reagent	+
	Mayer's reagent	+
	Wagner's reagent	+
	Hager's reagent	+
9	Carbohydrates Molish test	-
10	Glycosides Borntrager's test	+
11	Starch (Amylum)	-

**Antibacterial Activity:** The antibacterial activity of *Thuja occidentalis* was determined.

**Table 7: Zone Of Inhibition Petry Plate.**

Strain	Escherichia-coli				
	20	40	60	80	100
Concentration ( $\mu\text{g/ml}$ )					
Sample <i>Thuja occidentalis</i>	-	4	5	7	9
<b>Standard</b>					
Penicillin(60 $\mu\text{g/ml}$ )	28				
<b>Control (distilled water)</b>	Nil				



**Figure 2: Zone Of Inhibition.**

## CONCLUSION

The antibacterial activity of *Thuja occidentalis* was determined on various bacteria.

## REFERENCES

1. Fredrickson JK, Zachara JM, Balkwill DL et al. "Geomicrobiology of high-level nuclear waste-contaminated vadose sediments at the Hanford site, Washington state". *Applied and Environmental Microbiology*, 2004; 70(7): 4230–41.
2. Whitman WB, Coleman DC, Wiebe WJ. "Prokaryotes: the unseen majority". *Proceedings of the National Academy of Sciences of the United States of America*, 1998; 95(12): 6578–83.



3. C. Michael Hogan. Bacteria. Encyclopedia of Earth. eds. Sidney Draggan and C.J. Cleveland, National Council for Science and the Environment, Washington DC, 2010.
4. Rappé MS, Giovannoni SJ "The uncultured microbial majority". Annual Review of Microbiology, 2003; 57: 369–94.
5. Sears CL. "A dynamic partnership: celebrating our gut flora". Anaerobe, 2005; 11(5): 247–51.
6. "2002 WHO mortality data". Retrieved, 2007-01-20.
7. Ishige T, Honda K, Shimizu S "Whole organism biocatalysis". Current Opinion in Chemical Biology, 2005; 9(2): 174–80.
8. Woese CR, Kandler O, Wheelis ML "Towards a natural system of organisms: proposal for the domains Archaea, Bacteria, and Eucarya". Proceedings of the National Academy of Sciences of the United States of America, 1990; 87(12): 4576-9. Bibcode1990PNAS...87.4576W. doi:10.1073/pnas.87.12.4576.PMC 54159. PMID 2112744.
9. βακτήριον, Henry George Liddell, Robert Scott, A Greek-English Lexicon, on Perseus.
10. βακτηρία, Henry George Liddell, Robert Scott, A Greek-English Lexicon, on Perseus  
bacterium, on Oxford Dictionaries.
11. Porter JR "Antony van Leeuwenhoek: tercentenary of his discovery of bacteria". Bacteriological Reviews, 1976; 40(2): 260–9.
12. Van Leeuwenhoek A. "An abstract of a letter from Mr. Anthony Leevvenhoek at Delft, dated Sep. 17, 1683, Containing Some Microscopical Observations, about Animals in the Scurf of the Teeth, the Substance Call'd Worms in the Nose, the Cuticula Consisting of Scales". Philosophical Transactions (1683–1775)14 (155–166), 1684; 568–574.
13. Van Leeuwenhoek A. "Part of a Letter from Mr Antony van Leeuwenhoek, concerning the Worms in Sheeps Livers, Gnats, and Animalcula in the Excrements of Frogs". Philosophical Transactions (1683–1775) 22 (260–276), 1700; 509–518.
14. Van Leeuwenhoek A. "Part of a Letter from Mr Antony van Leeuwenhoek, F. R. S. concerning Green Weeds Growing in Water, and Some Animalcula Found about Them". Philosophical Transactions (1683–1775) Ehrenberg's Symbolae Physioe. Animalia evertebrata. Decas prima. Berlin, 1702; 1828.
15. Breed, R.; Conn, H. "The Status of the Generic Term Bacterium Ehrenberg 1828". Journal of bacteriology, 1936; 31(5): 517–518.
16. Ehrenberg (C.G.): Dritter Beitrag zur Erkenntniss grosser Organisation in der Richtung des kleinsten Raumes. Physikalische Abhandlungen der Koeniglichen Akademie der Wissenschaften zu Berlin aus den Jahren 1833-1835, 1835; 143-336.

17. "Pasteur's Papers on the Germ Theory". LSU Law Center's Medical and Public Health Law Site, Historic Public Health Articles. Retrieved, 2006-11-23.
18. "The Nobel Prize in Physiology or Medicine 1905". Nobelprize.org. Retrieved, 2006-11-22.
19. O'Brien S, Goedert J "HIV causes AIDS: Koch's postulates fulfilled". *Curr Opin Immunol*, 1996; 8(5): 613–8.
20. Thurston A. "Of blood, inflammation and gunshot wounds: the history of the control of sepsis". *Aust N Z J Surg*, 2000; 70(12): 855–61.
21. Schwartz R. "Paul Ehrlich's magic bullets". *N Engl J Med*, 2004; 350(11): 1079–80.
22. "Biography of Paul Ehrlich". Nobelprize.org. Retrieved, 2006-11-26.
23. Woese C, Fox G "Phylogenetic structure of the prokaryotic domain: the primary kingdoms". *Proc Natl Acad Sci USA*, 1977; 74(11): 5088–90.
24. Woese CR, Kandler O, Wheelis ML "Towards a natural system of organisms: proposal for the domains Archaea, Bacteria, and Eucarya". *Proceedings of the National Academy of Sciences of the United States of America*, 1990; 87(12): 4576.
25. Schopf J "Disparate rates, differing fates: tempo and mode of evolution changed from the Precambrian to the Phanerozoic". *Proc Natl Acad Sci USA*, 1994; 91(15): 6735.
26. DeLong E, Pace N "Environmental diversity of bacteria and archaea". *Syst Biol*, 2001; 50(4): 470–8.
27. Brown JR, Doolittle WF "Archaea and the prokaryote-to-eukaryote transition". *Microbiology and Molecular Biology Reviews*, 1997; 61(4): 456–502.
28. Poole A, Penny D "Evaluating hypotheses for the origin of eukaryotes". *Bioessays*, 2007; 29(1): 74–84.
29. Dyall S, Brown M, Johnson P "Ancient invasions: from endosymbionts to organelles". *Science*, 2004; 304(5668): 253–7.
30. Lang B, Gray M, Burger G "Mitochondrial genome evolution and the origin of eukaryotes". *Annu Rev Genet*, 1999; 33: 351–97.
31. McFadden G "Endosymbiosis and evolution of the plant cell". *Curr Opin Plant Biol*, 1999; 2(6): 513–9.
32. Schulz H, Jorgensen B. "Big bacteria". *Annu Rev Microbiol*, 2001; 55: 105–37.
33. Williams, Caroline "Who are you calling simple?". *New Scientist*, 2011; 211(2821): 38–41.
34. Robertson J, Gomersall M, Gill P. "Mycoplasma hominis: growth, reproduction, and isolation of small viable cells". *J Bacteriol*, 1975; 124(2).

35. Velimirov, B. "Nanobacteria, Ultramicrobacteria and Starvation Forms: A Search for the Smallest Metabolizing Bacterium". *Microbes and Environments*, 2001; 16(2): 67–77.
36. Dusenbery, David B. *Living at Micro Scale*, Harvard University Press, Cambridge, Mass. ISBN 978-0-674-03116-6, 2009; 20–25.
37. Fritz I, Strömpl C, Abraham W "Phylogenetic relationships of the genera *Stella*, *Labrys* and *Angulomicrobium* within the 'Alphaproteobacteria' and description of *Angulomicrobium amanitifforme* sp. nov". *Int J Syst Evol Microbiol*, 2004; 54(Pt 3): 651–7.
38. Wanger, G; Onstott, TC; Southam, G. "Stars of the terrestrial deep subsurface: A novel 'star-shaped' bacterial morphotype from a South African platinum mine". *Geobiology*, 2008; 6(3): 325–30.
39. Cabeen M, Jacobs-Wagner C "Bacterial cell shape". *Nat Rev Microbiol*, 2005; 3(8): 601–10.
40. Young K. "The selective value of bacterial shape". *Microbiol Mol Biol Rev*, 2006; 70(3): 660–703.
41. Douwes K, Schmalzbauer E, Linde H, Reisberger E, Fleischer K, Lehn N, Landthaler M, Vogt T "Branched filaments no fungus, ovoid bodies no bacteria: Two unusual cases of mycetoma". *J Am Acad Dermatol*, 2003; 49(2 Suppl Case Reports): S170–3.
42. Donlan R "Biofilms: microbial life on surfaces". *Emerg Infect Dis*, 2002; 8(9): 881–90. PMC 2732559. PMID 12194761.
43. Branda S, Vik S, Friedman L, Kolter R. "Biofilms: the matrix revisited". *Trends Microbiol*, 2005; 13(1): 20–6.
44. Davey M, O'toole G "Microbial biofilms: from ecology to molecular genetics". *Microbiol Mol Biol Rev*, 2000; 64(4): 847–67.
45. Donlan RM, Costerton JW "Biofilms: survival mechanisms of clinically relevant microorganisms". *Clin Microbiol Rev*, 2002; 15(2): 167–93.
46. Shimkets L "Intercellular signaling during fruiting-body development of *Myxococcus xanthus*". *Annu Rev Microbiol*, 1999; 53: 525–49.
47. Kaiser D "Signaling in myxobacteria". *Annu Rev Microbiol*, 2004; 58: 75–98.
48. Berg JM, Tymoczko JL Stryer L. *Molecular Cell Biology* (5th ed.). WH Freeman. ISBN 0-7167-4955-6, 2002.
49. Gitai Z. "The new bacterial cell biology: moving parts and subcellular architecture". *Cell*, 2005; 120(5): 577–86.

50. Shih YL, Rothfield L "The bacterial cytoskeleton". *Microbiology and Molecular Biology Reviews*, 2006; 70(3): 729–54.
51. Norris V, den Blaauwen T, Cabin-Flaman A et al. "Functional taxonomy of bacterial hyperstructures". *Microbiology and Molecular Biology Reviews*, 2007; 71(1): 230–53.
52. Kerfeld CA, Sawaya MR, Tanaka S et al. "Protein structures forming the shell of primitive bacterial organelles". *Science*, 2005; 309(5736): 936–8.