BIOANALYTICAL METHOD DEVELOPMENT AND VALIDATION OF AMLODIPINE BESYLATE BY USING RP-HPLC ON HUMAN PLASMA

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ABSTRACT

A simple, sensitive, rapid and precise bioanalytical RP-HPLC method was developed for estimation of Amlodipine besylate in human plasma. The work carried out on Shimadzu LC-2010 CHT HPLC system equipped with Waters C18 (150×4.6 mm,5μ) column with mobile phase containing 10 mMpotassium dihydrogen phosphate buffer, pH 6.4: Acetonitrile: Methanol in the ratio 20:70:10 v/v/v (1ml/min.)and detectionwavelength 238 nm. The retention time of Amlodipine besylate was found to be 4.1 min. The developed bioanalytical method was found to be linear in concentration range of 24-48 ng/ml (R^2= 0.9913). The precision study revealed that the percentage cumulative variation was within acceptable limit and accuracystudy showed the value of mean percent recovery between 100.41-103.80 %. The Amlodipine besylate was stable in human plasma at different storage conditions. The validation parameters of the method met the acceptance criteria. Sufficient stability of both LQC and HQC was shown to allow for completion of sample analysis in clinical trials. From the results, we can conclude that developed bioanalytical method can be used for routine analysis of Amlodipine besylate.

KEYWORDS: Bioanalytical method, Amlodipine besylate, Human plasma, RP-HPLC method, Validation.

1. INTRODUCTION

Amlodipine besylate is chemically 3-Ethyl 5-methyl (4RS)-2-[(2-aminoethoxy) methyl]-4-(2-Chlorophenyl)-6-methyl-1, 4-dihydropyridine-3, 5-dicarboxylate benzene sulphonate(Fig.1).[1] It is long-acting calcium channel blocker of dihydropyridine class used in the treatment of hypertension and angina pectoris.[2] Amlodipine relax the smooth muscle in the arterial wall, decreases total peripheral resistance thereby reduces blood pressure and increases blood flow to the heart muscle.[1,2] Amlodipine besylate is official in USP and IP.[3] Literature survey revealed that a wide variety of analytical techniques have been reported for the estimation of Amlodipine besylate either in combinations or individually which includes, UV Spectroscopy[1,4], RP-HPLC[5,6,7] and few bioanalytical techniques: LC-MS[8,9], LC/MS/MS.[10] Therefore, our aim to developa simple, precise and accurate bioanalytical method for estimation of amlodipine besylate by using RP-HPLC in human plasma and validate as per USFDA guidelines.[11]
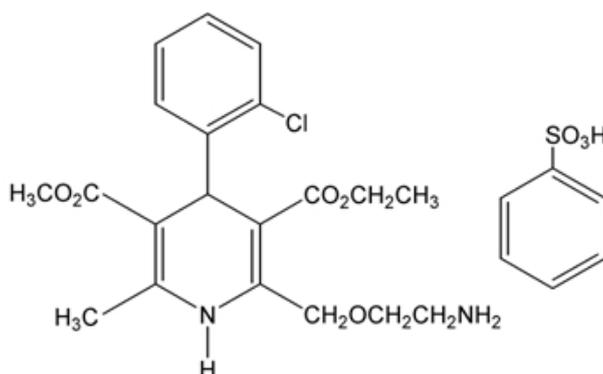

Fig. 1: Structure of Amlodipine besylate.

2. EXPERIMENTAL

2.1. Chemicals and reagents

Amlodipine besylate were obtained from Lupin Ltd.Aurangabad, Maharashtra. Water is purified by water purification system for HPLCpure lab UHQ, ELGA, Mumbai. HPLC grade methanol and acetonitrilewere obtained from Merck Ltd, Mumbai, India. Human plasma was procured from SKN hospital, Pune. Analytical grade of chemicals were obtained from Loba chemie Pvt. Ltd Mumbai, India.

2.2. Instrumentation

Chromatographic separation of drug was performed on Shimadzu LC-2010 CHT equipped with LC solution software with UV detector and weighing was done on electronic balance

Shimadzu ATY 224. Separation was carried out using "Waters" C18 column. Sample was prepared using REMI C24BL cooling centrifugation and REMI CM 101 cyclo mixer.

2.3. Standard stock solution

Amlodipine besylate API powder (100 mg) was accurately weighed and transferred to 100 ml volumetric flask. The drug was dissolved and diluted up to the mark with methanol. This solution was further diluted to get 100μg/ml Amlodipine besylate.

2.4. Preparation of mobile phase

Mobile phase was prepared by mixing 10mM Potassium dihydrogen phosphate buffer (pH 6.4), Acetonitrile and Methanol in ratio 20:70:10 v/v/v and filtered through 0.45μ membrane filter.

2.5. Preparation of buffer- (10mM Potassium dihydrogen phosphate) pH6.4

Disodium hydrogen phosphate dehydrate 1.78gwas weighed accurately, dissolved in 1000ml of distilled water and pH was adjusted to 6.4 by using O-Phosphoric acid.

2.6. Preparation of sample[12]

Protein precipitation technique

Frozen human plasma was thawed to ambient temperature and aliquots of 500 μlplasma were taken in centrifugation tubes of 10 ml capacity with the help of micropipette and 300 μl of stock solution was added and the plasma proteins were precipitated by using methanol. The tube was vortexed for 30 sec.Then the solution was centrifuged at10000 RPM for 12 min. below the temperature 10℃. The supernatant liquid wastaken and evaporated in water bath.

Then, the residue was dissolved in methanol and transfer to HPLC vials.

2.7. Method validation

The method performance was evaluated for accuracy, precision, linearity and stability during various stress conditions which include freeze thaw stability, stock solution and short term stability.

2.8. Linearity and Range

Working solution of various concentrations was injected under the operating chromatographic condition and peak area of each drug were calculated at 238 nm. The calibration curves were constructed using simple linear regression between peak area and

concentrations (Fig.4). The range of solution has been decided according to correlation coefficient of regression equation.

2.9. Accuracy

The accuracy of the method was performed by calculating % recovery for the different concentration levels of drug. The samples of three concentration levels prepared as LQC, MQC and HQC by standard addition method.

2.10. Precision

The precision of this method was evaluated by the % CV at different concentration levels corresponding to LQC, MQC and HQC. Intraday and interday precision was evaluated in 3 replicate batches of different concentrations (24, 36 and 48 ng/ml).

2.11. Stability studies

The stability of Amlodipine besylate in solution and plasma sample was evaluated using two concentration level (LQC and HQC i.e. 24and48ng/ml respectively). Thestability of Amlodipine besylate was also evaluated in deep freezing at -20 ℃ for 12 hrs. The plasma samples were kept at freezer and after stressed to three freeze thawing cycles (for 24 hrs. per cycle). All samples described above were compared to freshly prepared Amlodipine besylate sample at the same concentration level.

3. RESULTS AND DISCUSSION

3.1. Optimization of Chromatographic conditions

The chromatographic conditions were optimized in order to provide good system suitability parameters. The mobile phase was selected on the basis of its polarity and different trials were taken. Acetonitrile was selected as an organic modifier, because lower column efficiency was observed using methanol. Finally,a mobile phase consisting Buffer (pH 6.4): Acetonitrile: Methanol (20:70:10, v/v/v)at a flow rate of1ml/min was selected. The retention time of Amlodipine besylate was found to be 4.1 min. The chromatogram of Amlodipine besylate obtained by optimized conditions is shown in fig.3. The optimized chromatographic conditions and system suitability parameters are listed intable 1.

Table 1: Optimized chromatographic conditions and system suitability parameters.

Sr. No.	Condition/Parameter	Details
1	Column	Waters C18 5µm (4.6 x 250mm)
2	Mobile phase	Buffer: ACN: Methanol (20:70:10 v/v/v)
3	Flow rate	1.0 ml/min.
4	Column temperature	28 ℃
5	Volume of injection	20 µl
6	Detection wavelength	238 nm
7	Theoretical plate	2300
8	Retention time	4.1 min.
9	Tailing factor	1.29

Typical chromatograms of drug free human plasma and spiked drug-plasma of Amlodipine besylateare shown in fig.2 and 3, respectively. The retention time of amlodipine besylate was found to be 4.1 min indicating this method is faster than other methods. The typical column efficiency expressed as the number of theoretical plates was found to be 2300 for Amlodipine besylate.
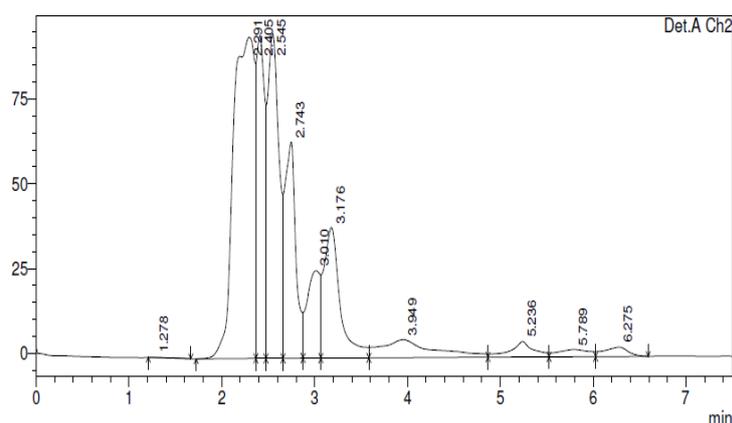
Fig. 2: Chromatogram of drug free human plasma.
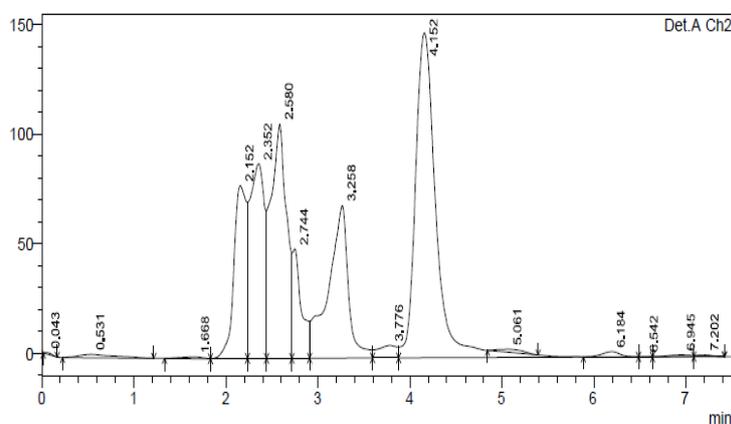
Fig. 3: Chromatogram of drug-human plasma.

3.2. Linearity and Range

The calibration curve was found to be linear in the range 24-48 ng/ml ($R^2 = 0.9913$) and equation is y = mx+c, where y represents the area of Amlodipine besylate and x represents concentration of Amlodipine besylate in ng/ml (fig. 4).
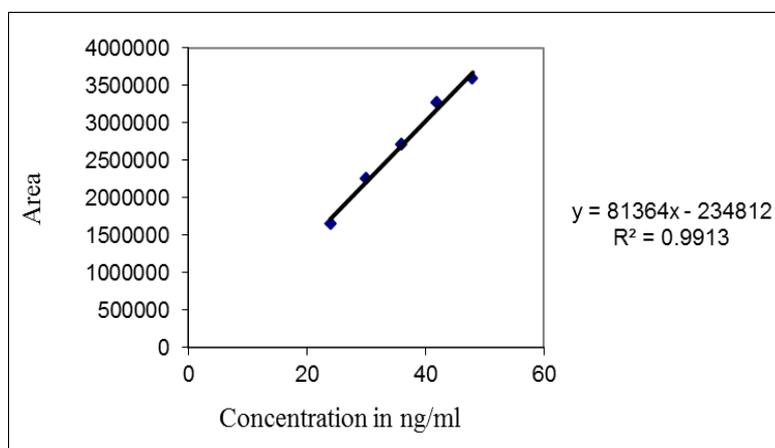

y = 81364x - 234812
R² = 0.9913

Fig. 4: Calibration curve of Amlodipine besylate

3.3. Accuracy

The mean % recovery of calculated concentrations for all quality control samples at LQC, MQC and HQC concentration levels are ranged from 100.41-103.80 %, which are within the acceptance criteria 85-115 % (Table 2).

Table 2: Results of Accuracy study.

	LQC	MQC	HQC
Mean (ng/ml)	22.512	32.476	51.063
S.D.	1.3021	0.0849	0.7791
% CV	5.7828	0.2614	1.5234
% Mean	93.79	90.19	108.18

3.4. Precision

The % CV of calculated concentrations for all quality control samples at LQC, MQC and HQC concentration levels are ranged from 1.478-0.6472 % for intraday and 3.586-1.6589 for interday precision, which is within acceptance criteria 15.00 % (Table 3).

Table 3: Results of intraday and interday precision.

Parameter		LQC	MQC	HQC
Intraday	S.D.	0.2471	0.2099	0.7127
	% CV	1.1338	0.6472	1.4786
Interday	S.D.	0.8533	1.2114	0.69143
	% CV	1.6589	3.5869	2.70504

(N=5)

3.5. Stability Studies

The results of all stability studies are within acceptance criteria (Table 4). The results of freeze thaw stability suggested that Amlodipine besylate was stable in human plasma for at least 24 hrs. The results of short term stability studies indicated that the quality control samples were stable for 12 hours at -20 ˚C. Similarly, the results of stock solution studies confirmed stability of stock solution.

Table 4: Results of stability studies.

Stability	Parameter	Concentrations	
		LQC	MQC
Freeze and thaw	Mean	22.18	47.96
	S.D.	0.2879	0.6460
	% CV	1.2981	1.3470
	% Mean stability	92.41	99.91
Short term stability	Mean	23.24	48.13
	S.D.	2.7942	0.8791
	% CV	12.02	1.82
	% Mean stability	96.83	100.27
Stock solution stability	Mean	21.48	45.01
	S.D.	0.7794	0.0832
	% CV	3.6288	0.1849
	% Mean stability	89.5	93.77

4. CONCLUSION

The work describe in this paper deals with analysis of Amlodipine besylate using RP-HPLC method in human plasma. The precision and accuracy of the method met the acceptance criteria laid down in guideline for industry, Bioanalytical method validation, USFDA. Sufficient stability of both LQC and HQC was shown to allow for completion of sample analysis in clinical trials. From the results, we can conclude that developed method id simple, accurate, rapid and precise. Thus, it can be used for routine analysis of Amlodipine besylate in human plasma.

5. ACKNOWLEDGEMENT

6. REFERENCES

1. Jampana P., Varma B., Babu G.,Surekha P., Praveen T., Rao P., Visible spectroscopic method for estimation of Amlodipine besylate in tablets, International Journal of Pharmaceutical, Chemical and Biological sciences, 2014; 4(1): 173-177.

2. Rang H., Dale M., Ritter J., Pharmacology. 5th ed., Elsevier publication, 2003; 282-283.

3. Indian pharmacopoeia, Government of India., New Delhi: Ministry of health and family welfare, 7th ed. 2014; 2: 1045-4018.

4. Topal B., Bozal B., Demircigil B., Uslu B., Ozekan S., Electroanalytical studies and simultaneous determination of Amlodipine Besylate and Atorvastatine calcium in binary mixtures using first derivative of the ratio-voltammetric methods, Wiley InterscienceElectroanalysis, 2009; 21: 2427-2439.

5. Sudhakar M., Rao V., Devika S., Ramesh R., A Validated RP-HPLC method for simultaneous estimation of NebivololHydrochloride and S-Amlodipine besylate in tablet dosage forms, International Journal of Chemical and Pharmaceutical Sciences, 2010; 1(2): 28-33.

6. Usha N., Satya B., Ankani N., Method development and validation for the simultaneous estimation of Nebivolol Hydrochloride and Amlodipine besylate in tablet dosage forms by RP-HPLC., International Journal of Pharmaceutical Sciences and Drug Research, 2015; 7(1): 110-115.

7. Ahmed M., Manohara N., Ravi C., RP-HPLC method development and validation for simultaneous estimation of Atorvastatin calcium and Amlodipine besylate, International Journal of ChemTech Research, 2012; 4(1): 337-345.

8. Streel B., Laine C., Zimmer C. Sibenaler R., CEccato A., Enantiomeric determination of Amlodipine in human plasma by liquid chromatography coupled to tandem mass spectrometry, Journal of Biochemical and Biophysical methods, 2002; 54: 357–368.

9. Sarkar A., Das D., Selvan P., Gowda K., Mandal U.,Bose A., Agarwal S., Bhaumik U., Pal T., Simultaneous determination of Metoprolol succinate and Amlodipine besylate in human plasma by liquid chromatography–tandem mass spectrometry method and its application in bioequivalence study, Journal of Chromatography B, 2008; 873: 77–85.

10. Narapusetti A., Bethanabhatla S., Sockalingam A., Repaka N., Saritha V., Simultaneous determination of Rosuvastatin and Amlodipine in human plasma using tandem mass spectrometry: Application to disposition kinetics, Journal of Advanced Research, 2015; 6; 931-940.

11. FDA Guidance for industry, Bioanalytical method validation, US Department of health and human services, Food and Drug Administration, 2013; 1: 1-23.

12. Pushpalatha E., Sailaja B., Biaanalytical method development and validation by HPLC: A review, Journal of medical and pharmaceutical innovation, 2014; 1(6): 1-9.

13. Murugan S., Pravallika N., Sisrisha P., Chandrakala K., A review on Bioanalytical method development and Validation by using LC-MS/MS,Journal of Chemical and Pharmaceutical Sciences, 2013; 6(1): 41-45.

14. Varghese A., Pandita N., Gaud R., A novel, sensitive, Bioanalytical method for estimation of Amlodipine besylate in rat plasma using Flourescence detection by RP-HPLC, International Journal of Pharmaceutical and Sciences and Research, 2014; 5(7): 2813-2820.