



## IN VIVO ANTIANXIETY ACTIVITY OF CALENDULA OFFICINALIS AERIAL PARTS IN MICE

Anita Rani<sup>1\*</sup> and Chander Mohan<sup>2</sup>

<sup>1\*</sup>Govt Pharmacy College, Nargota, Himachal Pradesh, India.

<sup>2</sup>Rayat-Bahra Institute of Pharmacy, Hoshiarpur, Panjab, India.

Article Received on  
03 April 2018,

Revised on 24 April 2018,  
Accepted on 14 May 2018,

DOI: 10.20959/wjpps20186-11697

### \*Corresponding Author

Anita Rani

Govt Pharmacy College,  
Nargota, Himachal Pradesh,  
India.

### ABSTRACT

Traditionally, all parts of *Calendula officinalis* have been used for treating various diseases including anxiety disorders. Thus, we focused our study to investigate the antianxiety potential of *C. officinalis* aerial parts extracts and fractions using elevated plus maze (EPM) model. Extracts of *C. officinalis* aerial parts were prepared using solvents of increasing polarity (petroleum ether (60-80°C), chloroform, methanol and water) and were subjected to antianxiety activity on mice at the doses of 100, 200, 400 mg/kg, po. 200 mg/kg of *C. officinalis* methanol extract produced antianxiety effect comparable to reference

compound, diazepam (2 mg/kg, po). Bioactive methanol extract of aerial parts *C. officinalis* was fractionated using ethyl acetate and found that it exhibits antianxiety activity and statistically equivalent to reference drug. Further, fraction was subjected to column chromatography in order to isolate sub-fractions responsible for antianxiety effect. Sub-fractions obtained were evaluated for the antianxiety activity in mice, among all only sub-fraction 4 (F<sub>4</sub>) was found active, and compound isolated (CO<sub>1</sub>) from the sub-fraction F<sub>4,3</sub> exhibited significant antianxiety activity at the dose of 6 mg/kg, po. Altogether, we can conclude that the results of *C. officinalis* aerial parts validate its traditional claim in the treatment of anxiety disorders.

**KEYWORDS:** *Calendula officinalis*, anxiety, methanol extract, chromatography, traditional, mice.

## INTRODUCTION

*Calendula* (Family:Asteraceae) is a genus of perennial or annual herb worldwide including 25 species which are mainly distributed throughout southwestern Asia, western Europe, Macaronesia and the Mediterranean region. These are often known as marigolds.<sup>[1-3]</sup> *Calendula* was a natural drug therefore used traditionally in the treatment as an anti-spasmodic, anti-worm and diuretic.<sup>[4]</sup> Leaves and flowers of *C. officinalis* were considered as a natural herb and used as.

Diaphoretic, stimulant, emmenagogue, antispasmodic and also in the treatment of measles and smallpox and metrorrhagia.<sup>[5,6]</sup> whereas in India ointment of florets used in treating herpes, wounds, frostbite, ulcers, skin damage and scars.<sup>[7-9]</sup>

Traditional use of *C. officinalis* in treating inflammations, gastrointestinal ulcers and dysmenorrhea was revealed by a survey of ethnopharmacologic records.<sup>[10]</sup> Dried flower heads of *Calendula* have been used as antipyretic, antitumor, for cicatrizing wounds and also as a diuretic and diaphoretic.<sup>[11]</sup> *C. officinalis* also used topically used as a antifungal and antiseptic for treating wounds, sprain, conjunctivitis and marks.<sup>[12]</sup> Infusion of *Calendula* is used in the treatment of chronic infections and act as a cleansing and detoxifying herb.<sup>[13]</sup> In homoeopathic medicine, its mother tincture is used for the mental tension and insomnia treatment.<sup>[14]</sup>

In Ayurvedic and Unani System of Medicine, leaves and flowers of *C. officinalis* are considered for the treatment of antimicrobial, anti-inflammatory, antipyretic and antiepileptic.<sup>[15]</sup> In traditional and homoeopathic medicine, whole plant used in the treatment of menstrual irregularities, duodenal ulcers, hemorrhoids and varicose veins.<sup>[16]</sup> In the middle ages, flowers of *C. officinalis* have been used for snake bites and liver obstruction. In 18<sup>th</sup> century; this plant was considered the best remedy for red eyes, jaundice and headache.

*C. officinalis* also included as part in the treatment of dry skin, bee stings and foot ulcers.<sup>[17]</sup> Essential oil extracted from the *C. officinalis* is used for soothing central nervous system and also act as a wound healer.<sup>[18]</sup>

After chemical research, various biologically active chemical constituents reported in *C. officinalis* include triterpenoids glycosides, flavonoids, sesquiterpenes glycoside, phenolic

acids, carotenoids, hydroxycoumarins, Volatile oils and fatty acids. The objective of the present study was to evaluate potential antianxiety activity of the *C. officinalis* aerial parts.

## MATERIAL AND METHODS

### Plant material and reagents

Aerial parts of *C. officinalis* were procured from the Himalaya Herbs Store, Saharanpur, UP, India. Identified by Head, Raw Materials, Herbarium & Museum at National Institute of Science Communication and Information Resources (NISCAIR), New Delhi vide letter dated 19.10.2014 bearing number NISCAIR/RHMD/Consult/-2010-11/1431/29. Petroleum ether (60–80°C), chloroform, methanol and ethyl acetate used for extraction of plant material. Anisaldehyde (0.5%) was used thin layer chromatography (TLC) for preparing visualizing reagent. Diazepam used as reference drug for the antianxiety activity.

### Preparation of extracts

*C. officinalis* aerial parts (100 g) were dried, powdered and successively soxhlet extracted with petroleum ether, chloroform, methanol and water in the order of increasing polarity and the extracts were preserved in vacuum desiccators containing anhydrous silica gel blue.

### Animals

The experimental animals (Swiss albino mice: males; n=6), weighing about 20-25g were bred at the Central Animal House, L R Institute of Pharmacy, Solan. All mice were maintained under controlled environmental conditions (25±2°C) with 12 h light- 12 h dark cycle, given standard feed, water *ad libitum* and were fasted for 4 h before use. Animal Ethical Committee, L R Institute of Pharmacy, Solan has approved the use of animal for carrying out the biological studies. All the experiments were carried out as per CPCSEA guidelines.

### Acute toxicity studies

These were conducted following OECD 423 guidelines (OECD, 2001). Single oral dose (500, 1000 or 2000 mg/kg of various prepared extracts of *C. officinalis* aerial parts were administered to mice in different groups. The mice were kept under observation immediately to see the sign of toxicity during the first 0.5, 1, 2, 4, 8 and 12 h, and at every 24 h for 14 days. Various behavioral parameters, lethargy, amount of water, feed taken, tremors and death were observed. Results reported that none of the extracts exhibited signs of toxicity upto the dose of 2000 mg/kg, po.

### Preparation of doses

Test doses (100, 200 or 400 mg/kg, po) of various extracts of *C. officinalis* aerial parts were prepared using simple syrup (IP)+Tween 80 in the ratio 95:5 (vehicle) and diazepam (2 mg/kg, po) as a reference drug was administered to mice 45 min prior to EPM study using tuberculin syringe fitted with oral canula in the volume ranging between 0.20-0.30 ml.

### Experimental model to test anxiolytic activity

**Antianxiety study of plant extracts was evaluated by using well established animal model i.e. EPM.** It is 25 cm elevated from the floor, consisting of two open arms (16×5 cm) and two closed arms (16×5×12 cm) and radiant open roof which form plus sign to observed anxiolytic behaviour of animal.<sup>[19]</sup> Individual animal was placed at the center of model with its head towards one of the open arms. During the 5-min experiment, behaviour of the mouse was recorded as: (a) the number of entries the open arms and (b) average time spent by the mouse in the open arms (average time = total time spent in open arms/number of entries in open arms). Test dose was administered orally using tuberculin syringe fitted with an oral canula and such adjusted that every animal gets its turn after 45 min of the dose administration. And we have to ensure that no external stimuli other than the height of plus-maze could invoke anxiety in the animals.

### TLC finger print profile of methanol extract

Dried powder of aerial parts of *C. officinalis* (2 g) was packed in filter paper sachet, placed inside 100 ml round bottom flask, macerated for 30 min with methanol (50 ml) followed by reflux (30 min) on a boiling water bath. After cooling, the extract was decanted off. Solvent was recovered under reduced pressure using rotary evaporator. The dried residue was reconstituted in 5 ml volumetric flask using methanol. For optimizing the mobile phase, aliquots 2 µl of extract solution was applied on TLC glass plate and developed in different solvent systems. Thin layer chromatograms were visualized initially under UV light at 366 nm followed by spraying with 0.5% anisaldehyde followed by heating at 110°C for 10 min. TLC finger print profile of methanol extract was obtained on precoated aluminium-based TLC plates, developed in toluene:chloroform:methanol (7.2:1), using CAMAG auto sampler and scanner under controlled conditions.

### Preparation and fractionation of methanol extract

Methanol extract of dried *C. officinalis* aerial parts (500 g) was prepared. Dried methanol extract (100 g), suspended in water, was placed in three-necked round bottom flask. It was, then, partitioned with ethyl acetate (100 ml) by heating (50°C) for 30 min using rotamantle. The procedure was repeated five times. All the ethyl acetate fractions were pooled. Solvent from the pooled fraction and the remaining aqueous fraction was recovered under reduced pressure using rotary vacuum evaporator to get ethyl acetate soluble fraction (EASF) and ethyl acetate insoluble fraction (EAIF), respectively.

### Column chromatography of EASF and sub-fractions

EASF (20 g) was subjected to column chromatography using silica gel. Elution was done with petroleum ether, chloroform and methanol, in appropriate proportions. A total of 150 fractions, each of 500 ml were collected and pooled on the basis of TLC profiles to get four fractions – F<sub>1</sub>-F<sub>4</sub>.

The F<sub>4</sub> (13 g) was subjected to column chromatography. The column was packed with silica gel in petroleum ether. Elution was done with petroleum ether, chloroform and methanol, in appropriate proportions. A total of 162 fractions, each of 250 ml, were collected and pooled on the basis of TLC profiles, to get three sub fractions F<sub>4.1</sub>- F<sub>4.3</sub>. A crystalline compound – CO<sub>1</sub> (2.67 g), was separated from concentrated solutions of the sub-fraction F<sub>4.3</sub>.

### Statistics

The results have been expressed as Mean ± Standard Error Mean (SEM). The test doses were compared among themselves, and also with standard and control by one way analysis of variance (ANOVA) followed by Student Neumann Keuls test.<sup>[20]</sup> Control group was also compared with the standard group.

## RESULTS

### Yield of extracts

After successive soxhlet extraction of *C. officinalis* aerial parts using solvents in increasing polarity viz. petroleum ether, chloroform, methanol and water extracts and yield was found to be 07.26%, 05.42%, 20.35% and 07.91% (w/w), respectively.

### Acute toxicity studies

Among all the extracts of *C. officinalis* aerial parts, none of them was observed toxic up to a dose of 2 000 mg/kg.

### Antianxiety activity of extracts

After oral administration of the extracts of *C. officinalis* aerial parts at the oral doses of 100, 200 or 400 mg/kg, diazepam (2 mg/kg) and the control (vehicle), the mean time spent by the mice in open arms shown in **Table 1**. Among the tested extracts, dose of 100 mg/kg, p.o. of methanol extract was observed to have maximum anxiolytic activity.

TLC finger print profile of methanol extract was obtained on precoated aluminium-based TLC plates, developed in toluene: chloroform: methanol (7.2:1), using CAMAG auto sampler and scanner under controlled conditions (**Table 2**).

Ethyl acetate fraction was obtained by further fractionation of bioactive methanol extract. Yields of EASF and ESIF were found to be 21.6 and 76.2 % (w/w), respectively and evaluated for antianxiety activity using EPM in mice. The mean number of entries and time spent in open arms of EPM were shown in **Table 3** at the doses 20 and 75 mg/kg, po, of *C. officinalis* aerial parts, diazepam (2 mg/kg, po) and the control (vehicle) and found that only EASF at the dose of 20 mg/kg significantly exhibited antianxiety with respect to control and the activity was equivalent to the reference drug.

EASF fraction was again fractionated by column chromatography; fraction yielded four sub-fractions (F<sub>1</sub>–F<sub>4</sub>), out of which only F<sub>4</sub> at a dose of 15 mg/kg exhibited significant antianxiety activity (Table 4). Four subfractions (F<sub>4.1</sub>–F<sub>4.3</sub> and CO<sub>1</sub>) were obtained from column chromatography of F<sub>4</sub> fraction and screened for antianxiety activity. Among them, isolated compound (CO<sub>1</sub>) at a dose of 6 mg/kg, po, show significant increase in the number of entries ( $7.5 \pm 0.10$ ) and the time spent ( $12.5 \pm 0.24$  s) in the open arms (Table 5).

**Table 1: Antianxiety activity of different extracts of *C. officinalis* aerial plants using EPM.**

| Treatment               | Dose (mg/kg, po) | Number of entries in open arms (Mean <sup>n</sup> ±SEM) | Average time spent in open arms (sec) (Mean <sup>n</sup> ±SEM) |
|-------------------------|------------------|---|--|
| Vehicle                 | 0.25 ml          | $3.2 \pm 0.13^b$  | $3.7 \pm 0.23^b$   |
| Diazepam                | 2                | $7.5 \pm 0.12^a$  | $15.2 \pm 0.32^a$  |
| Petroleum ether extract | 100              | $2.7 \pm 0.45^b$  | $6.4 \pm 0.34^b$   |
|                         | 200              | $3.1 \pm 0.77^b$  | $2 \pm 0.43^b$   |
|                         | 400              | $3.5 \pm 0.55^b$  | $2.2 \pm 0.24^b$   |
| Chloroform extract      | 100              | $2.5 \pm 0.55^b$  | $3.4 \pm 0.81^b$   |
|                         | 200              | $2.4 \pm 0.55^b$  | $2.1 \pm 0.36^b$   |

|                         |            |                                    |                                     |
|-------------------------|------------|------------------------------------|-------------------------------------|
|                         | 400        | $2.9 \pm 0.85^b$                   | $2.6 \pm 0.45^b$                    |
| <b>Methanol extract</b> | <b>100</b> | <b><math>5.9 \pm 0.89^b</math></b> | <b><math>11.9 \pm 0.31^b</math></b> |
|                         | 200        | $4.7 \pm 0.85^b$                   | $10.1 \pm 1.25^b$                   |
|                         | 400        | $3.8 \pm 0.84^b$                   | $8.7 \pm 0.73^b$                    |
| Water extract           | 100        | $2.3 \pm 0.45^b$                   | $3.3 \pm 0.45^b$                    |
|                         | 200        | $2.8 \pm 0.45^b$                   | $2.5 \pm 0.51^b$                    |
|                         | 400        | $3.2 \pm 0.77^b$                   | $3.3 \pm 0.53^b$                    |

n=6; The data is expressed as mean $\pm$ SEM; p<0.001, <sup>a</sup>: vs. control, <sup>b</sup>: vs. diazepam; one way ANOVA followed by Student Newmann Keul's test.

**Table 2: Mobile phases employed for TLC of methanol extract of *C. officinalis* aerial plants.**

| Sr. no. | Mobile phase                              | Proportions |
|---------|---|-------------|
| 1       | Chloroform:Methanol                       | 8:2         |
| 2       | Chloroform:Methanol                       | 9:1         |
| 3       | Toluene:Ethyl acetate                     | 9:1         |
| 4       | Toluene:Ethyl acetate:Glacial acetic acid | 7:2:1       |
| 5       | Toluene: Glacial acetic acid              | 9:1         |
| 6       | Propanol:Water                            | 9:1         |
| 7       | Propanol:Water                            | 8:2         |
| 8       | Toluene:Chloroform:Methanol               | 6:2:2       |
| 9       | Toluene:Chloroform:Methanol               | 6:3:1       |
| 10*     | Toluene:Chloroform:Methanol               | 7:2:1       |

\*Optimum resolution

**Table 3: Antianxiety activity of EASF and EAIF of methanol extract using EPM.**

| Treatment   | Dose (mg/kg, po) | Number of entries in open arms (Mean <sup>n</sup> $\pm$ SEM) | Average time spent in open arms (sec) (Mean <sup>n</sup> $\pm$ SEM) |
|-------------|------------------|--|---|
| Vehicle     | 0.25 ml          | $3.2 \pm 0.13^b$   | $3.7 \pm 0.23^b$  |
| Diazepam    | 2                | $7.5 \pm 0.12^a$   | $15.2 \pm 0.32^a$   |
| <b>EASF</b> | <b>20</b>        | <b><math>5.4 \pm 0.15^b</math></b>                           | <b><math>9.6 \pm 0.32^{a,b}</math></b>                              |
| EAIF        | 75               | $2.4 \pm 0.12^b$   | $4.8 \pm 0.21^b$  |

n=6; The data is expressed as mean $\pm$ SEM; p<0.001, <sup>a</sup>: vs. control, <sup>b</sup>: vs. diazepam, One way ANOVA followed by Student Newmann Keul's test. EASF: ethyl acetate soluble fraction, EAIF: ethyl acetate insoluble fraction.

**Table 4: Antianxiety activity of F<sub>1</sub>-F<sub>4</sub> fractions of EASF.**

| Treatment      | Dose (mg/kg, po) | Number of entries in open arms (Mean <sup>n</sup> $\pm$ SEM) | Average time spent in open arms (sec) (Mean <sup>n</sup> $\pm$ SEM) |
|----------------|------------------|--|---|
| Vehicle        | 0.25 ml          | $3.2 \pm 0.13^b$   | $3.7 \pm 0.23^b$  |
| Diazepam       | 2                | $7.5 \pm 0.12^a$   | $15.2 \pm 0.32^a$   |
| F <sub>1</sub> | 0.5              | $2.2 \pm 0.14^b$   | $3.2 \pm 0.31^b$  |



|                |     |                           |                            |
|----------------|-----|---------------------------|----------------------------|
| F <sub>2</sub> | 1.5 | 2.0 ± 0.16 <sup>b</sup>   | 2.8 ± 0.25 <sup>b</sup>    |
| F <sub>3</sub> | 3   | 2.7 ± 0.19 <sup>b</sup>   | 3.1 ± 0.33 <sup>b</sup>    |
| F <sub>4</sub> | 15  | 5.5 ± 0.13 <sup>a,b</sup> | 10.2 ± 0.21 <sup>a,b</sup> |

n=6; The data is expressed as mean±SEM, p<0.001, <sup>a</sup>: vs. control, <sup>b</sup>: vs. diazepam; One way ANOVA followed by Student Newmann Keul's test. F<sub>1</sub>-F<sub>4</sub>: fractions of EASF.

**Table 5: Antianxiety activity of various sub-fractions using EPM.**

| Treatment          | Dose (mg/kg, po) | Number of entries in open arms (Mean <sup>n</sup> ±SEM) | Average time spent in open arms (sec) (Mean <sup>n</sup> ±SEM) |
|--------------------|------------------|---|--|
| Vehicle            | 0.25 ml          | 3.2 ± 0.13 <sup>b</sup>                                 | 3.7 ± 0.23 <sup>b</sup>  |
| Diazepam           | 2                | 7.5 ± 0.12 <sup>a</sup>                                 | 15.2 ± 0.32 <sup>a</sup>                                       |
| F <sub>4.1</sub>   | 2                | 2.6 ± 0.20 <sup>b</sup>                                 | 2.3 ± 0.33 <sup>b</sup>  |
|                    | 4                | 2.5 ± 0.18 <sup>b</sup>                                 | 2.0 ± 0.36 <sup>b</sup>  |
| F <sub>4.2</sub>   | 5                | 2.4 ± 0.14 <sup>b</sup>                                 | 1.8 ± 0.30 <sup>b</sup>  |
|                    | 10               | 2.6 ± 0.19 <sup>b</sup>                                 | 2.6 ± 0.27 <sup>b</sup>  |
| F <sub>4.3.1</sub> | 4                | 3.0 ± 0.12 <sup>b</sup>                                 | 2.3 ± 0.31 <sup>b</sup>  |
|                    | 8                | 2.6 ± 0.19 <sup>b</sup>                                 | 2.6 ± 0.27 <sup>b</sup>  |
| CO <sub>1</sub>    | 3                | 4.0 ± 0.12 <sup>b</sup>                                 | 7.8 ± 0.31 <sup>b</sup>  |
|                    | 6                | 7.5 ± 0.10 <sup>b</sup>                                 | 12.5 ± 0.24 <sup>b</sup>                                       |

n=6; The data is expressed as mean±SEM, p<0.001, <sup>a</sup>: vs. control, <sup>b</sup>: vs. diazepam; One way ANOVA followed by Student Newmann Keul's test. F<sub>4.1</sub>, F<sub>4.2</sub>, F<sub>4.3</sub>, F<sub>4.3.1</sub> and CO<sub>1</sub> sub-fractions obtained after column chromatography of F<sub>4</sub>.

## DISCUSSION

Affective disorders, especially anxiety and depression, are among the most common disorders.<sup>[21]</sup> Both are associated with diminished health status and substantially lower health-related quality of life that persists over time.<sup>[22-23]</sup> To date, benzodiazepines (BZDs) are the most widely for psychiatric disorders.<sup>[24-25]</sup> but the adverse effects caused by BZDs are a matter of concern. Thus, drugs obtained from natural sources are emerging as adjuvant therapies in the treating the various diseases<sup>[26]</sup> and psychiatric disorders,<sup>[27]</sup> as these are relatively free from adverse effects.

Despite the widespread use of *C. officinalis* aerial parts for treating mental disorders, no systematic studies have been carried out for evaluating its antianxiety activity. The present investigation demonstrates that among the various extracts of *C. officinalis* aerial parts, methanol extract at the dose of 100 mg/kg, po exhibited significant anxiolytic effect with respect to vehicle and equivalent to the reference drug. Bioactivity-guided isolation of the methanol extract yielded anxiolytic fraction F<sub>4</sub> at the dose of 15 mg/kg, po which was at par



with that of diazepam. The column chromatography of bioactive fraction of F<sub>4</sub> yields three sub-fractions F<sub>4.1</sub>-F<sub>4.3</sub> and from F<sub>4.3</sub> sub-fraction yields CO<sub>1</sub> as a crystalline compound. This crystalline compound was separately suspended in a suitable vehicle, and evaluated for the antianxiety activity compared with that observed in the control group as well as with the group treated with the reference anxiolytic drug diazepam. Maximum anxiolytic activity was found to be in CO<sub>1</sub> at the dose of 6 mg/kg, po which was at par with that of diazepam as is evident from statistical equivalence between the results of this dose and that manifest by diazepam.

### CONFLICTS OF INTEREST

All authors have none to declare

### ACKNOWLEDGEMENTS

Authors duly acknowledge the L.L.R. Educational Trust, Solan, Himachal Pradesh, India, which runs the L R Institute of Pharmacy, Solan and IKG PTU, Jalandhar for providing assistance to Anita Rani for this research work.

### REFERENCE

1. Arora D, Rani, A, Sharma, A. A review on phytochemistry and ethnopharmacological aspects of genus *Calendula*. *Pharmacognosy Reviews*, 2013; 7(14): 179-187.
2. Naguib NY, Khalil MY, El Sherbeny SE. A comparative study on the productivity and chemical constituents of various sources and species of *Calendula* plants as affected by two foliar fertilizers. *Journal of Applied Sciences Research*, 2005; 1(2): 176-189.
3. Baciu AD, Mihalte L, Sestras AF, Sestras RE. Variability of decorative traits, response to the *Aphis fabae* attack and RAPD diversity in different genotypes of *Calendula*. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 2010; 38(3): 265.
4. Salehi- Sormaghi MH. *Medicinal plants and Herbal medicine*. Tehran, Iran: The culture institute of nutrition and health press; Chap 1 (In Persian), 2009; 1.
5. Kirtikar KR, Basu BD. *Indian Medicinal Plants Vol-2* Bishen Singh Mahendra Pal Singh; Dehradun, 1918; 4.
6. Khare CP. *Encyclopedia of Indian medicinal plants*. NewYork: Springes-Verlag Berlin Heidelberg, 2004.
7. Kemper KJ. *Calendula (Calendula officinalis)*. Longwood Herbal Task Force, 1999: 1.
8. Mehta D, Rastogi P, Kumar A, Chaudhary AK. Review on Pharmacological Update: *Calendula officinalis* Linn. *Planta Activa*, 2012; 4: 195-202.

9. Cordova CA, Siqueira IR, Netto CA, *et al.* Protective properties of butanolic extract of the *Calendula officinalis* L.(marigold) against lipid peroxidation of rat liver microsomes and action as free radical scavenger. *Redox report*, 2002; 7(2): 95-102.
10. Yoshikawa M, Murakami T, Kishi A, Kageura T, Matsuda H. Medicinal flowers. III. Marigold.(1): hypoglycemic, gastric emptying inhibitory, and gastroprotective principles and new oleanane-type triterpene oligoglycosides, calendasaponins A, B, C, and D, from Egyptian *Calendula officinalis*. *Chemical and Pharmaceutical Bulletin*, 2001; 49(7): 863-870.
11. Ukiya M, Akihisa T, Yasukawa K, Tokuda H, Suzuki T, Kimura Y. Anti-inflammatory, anti-tumor-promoting, and cytotoxic activities of constituents of marigold (*Calendula officinalis*) flowers. *Journal of Natural Products*, 2006; 69(12): 1692-1696.
12. Rehecho S, Uriarte-Pueyo I, Calvo J, Vivas LA, Calvo MI. Ethnopharmacological survey of medicinal plants in Nor-Yauyos, a part of the Landscape Reserve Nor-Yauyos-Cochas, Peru. *Journal of Ethnopharmacology*, 2011; 133(1): 75-85.
13. Blumenthal M, Goldberg A, Brinckmann J. Herbal Medicine: Expanded Commission E Monographs. Austin, TX, Boston: American Botanical Council. *Integrative Medicine Communications*, 2001; 376-378.
14. Boerick W. *Pocket Manual of Homoeopathic Materia Medica*. In :. B. Jain Publishers, 1998; 156–183.
15. Kasiram K, Sakharkar PR, Patil AT. (). Antifungal activity of *Calendula officinalis*. *Indian Journal of Pharmaceutical Sciences*, 2000; 62(6): 464-466.
16. Cetkovic GS, Đilas SM, Canadanovic-Brunet JM, Tumbas VT. Thin-layer chromatography analysis and scavenging activity of marigold (*Calendula officinalis* L) extracts. *Acta periodica technologica*, 2003; (34): 93-102.
17. Miliauskas G, Venskutonis PR, Van Beek TA. Screening of radical scavenging activity of some medicinal and aromatic plant extracts. *Food chemistry*, 2004; 85(2): 231-237.
18. Wynn SG, Fougere B. *Veterinary Herbal Medicine E-Book*. Elsevier Health Sciences, 2006.
19. Kulkarni SK. *Handbook of Experimental Pharmacology*, 3rd, Vallabh Prakashan, Pitampura, New Delhi, 2003; 135.
20. Scheffer WC. *Statistics for the Biological Sciences*. Addison-Wesley Publishing Company, Inc., Philippines, 1980; 121-41.
21. Farnsworth NR. Biological and phytochemical screening of plants. *Journal of Pharmarmaceutical Sciences*, 1966; 55(3): 225–276.

22. DiMatteo MR, Lepper HS, Croghan TW. Depression is a risk factor for noncompliance with medical treatment: meta-analysis of the effects of anxiety and depression on patient adherence. *Arch Intern Med*, 2000; 160(14): 2101–2107.
23. Sherbourne CD, Wells KB, Meredith LS, Jackson CA, Camp P. Comorbid anxiety disorder and the functioning and well-being of chronically ill patients of general medical providers. *Arch Gen Psychiatry*, 1996; 53(10): 889–895.
24. Baum C, Kennedy DL, Forbes MB, Jones JK. Drug use in the United States in 1981. *JAMA*, 1984; 251(10): 1293–1297.
25. Swartz M, Landerman R, George LK, Melville ML, Blazer D, Smith K. Benzodiazepine anti-anxiety agents: prevalence and correlates of use in a southern community. *American Journal of Public Health*, 1991; 81(5): 592–596.
26. Kumar S, Malhotra R, Kumar D. Euphorbia hirta: its chemistry, traditional and medicinal uses, and pharmacological activities. *Pharmacogn Rev*, 2010; 4(7): 58–61.
27. Anuradha H, Srikumar BN, Shankaranarayana Rao BS, Lakshmana M. Euphorbia hirta reverses chronic stress-induced anxiety and mediates its action through the GABA<sub>A</sub> receptor benzodiazepine receptor-Cl<sup>-</sup> channel complex. *Journal of Neural Transmission (Vienna)*, 2008; 115(1): 35–42.