



FLAVONOID-RICH EXTRACT OF *SOLANUM MACROCARPON* LEAVES SHOWS TISSUE PROTECTIVE AND AMELIORATE CERTAIN INFLAMMATORY AND OXIDATIVE INDICES ASSOCIATED WITH D-GALACTOSE EXPOSED RATS

Seyi Elijah Elasoru^{1,2,*}, Olaniyi Temitope Adedosu¹, John Olabode Fatoki¹ and Busuyi David Kehinde¹

¹Department of Biochemistry, Faculty of Basic Medical Sciences, Ladoke Akintola University of Technology P. M. B. 4000, Ogbomoso, Oyo State. Nigeria.

²Laboratório de Membranas Excitáveis e Biologia Cardiovascular (LAMEX), Departamento de Bioquímica e Imunologia, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Belo Horizonte, Brazil.

Article Received on
16 Dec. 2017,
Revised on 05 Jan. 2018,
Accepted on 26 Jan. 2018
DOI: 10.20959/wjpps20182-10987

***Corresponding Author**

Seyi Elijah Elasoru

Department of
Biochemistry, Faculty of
Basic Medical Sciences,
Ladoke Akintola University
of Technology P. M. B.
4000, Ogbomoso, Oyo
State. Nigeria.

ABSTRACT

The present study assessed the protective potentials of Flavonoid-Rich Extract of *Solanum macrocarpon* Leaves (FRESML) against D-galactose-induced oxidative stress in rats. Animals were randomly selected into four groups of seven per group and treated for 28 days. Group A (control) were given distilled water, Group B (extract only) were administered 100mg/kg/day of FRESML, Group C (D-galactose only) were administered 500mg/kg/day of D-galactose and Group D (D-galactose + extract) received combined administration of 500mg/kg/day of D-galactose and 100mg/kg/day of FRESML. Total Protein (TP), Urea and Creatinine concentrations, Alkaline Phosphatase (ALP), Alanine Aminotransferase (ALT), Aspartate Aminotransferase (AST), Gamma Glutamyl Transaminase (GGT), Superoxide Dismutase (SOD) and Catalase activities, Reduced

Glutathione (GSH), Malondialdehyde (MDA), Packed Cell Volume (PCV), White Blood Cell (WBC) and Fibrinogen levels were assessed using standard assay methods. D-galactose administration in rats triggered significant ($p < 0.05$) increases in serum levels of ALT, AST, GGT, ALP, Urea and Creatinine and significant ($p < 0.05$) decreases in serum, liver and

kidney TP concentrations. Significant ($p < 0.05$) increases in percentage PCV, WBC counts and Fibrinogen concentration were obtained. In addition, significant ($p < 0.05$) increases in liver and kidney MDA concentrations and depletion in liver and kidney SOD, Catalase and GSH levels were also obtained. Combined administration of D-galactose and FRESML significantly ($p < 0.05$) modulated the levels of ALT, AST, GGT, ALP, Urea and Creatinine and significantly ($p < 0.05$) increased serum, liver and kidney TP to nearly control levels. The extract significantly ($p < 0.05$) reversed D-galactose-induced increases in percentage PCV, WBC counts, Fibrinogen and MDA concentration and increases liver and kidney SOD, Catalase and GSH to nearly control level. Group treated with the extract alone showed no adverse effects on the parameters. Results are possible indication of FRESML ability to protect against cellular oxidative stress in tissues, boost antioxidant status and ameliorate inflammation associated with oxidative stress.

KEYWORDS: Antioxidant, D-galactose, Oxidative Stress, Protective, *Solanum macrocarpon*.

INTRODUCTION

All living organisms are exposed to reactive oxygen species (ROS), reactive nitrogen species (RNS) and other highly reactive mediators such as Advanced Glycation End Products (AGEs) throughout their life cycle.^[1] Though the body is equipped with the antioxidant system to protect it from excessive oxidant insults,^[2] oxidative stress occur when there is imbalance between the systemic manifestation of these reactive species and the biological system's ability to detoxify the reactive intermediates.^[3] Oxidative stress is among the largest known risk factors for human diseases.^[4] Oxidative stress plays a role in heart diseases, neurodegenerative diseases, cancer and in the aging process.^[5,6] D-galactose administration is associated with multiple physiological and biochemical changes, the chief among which is formation of Advanced Glycation End Products.^[7-9] Advanced Glycation End Products are a heterogeneous complex group of compounds that are formed when reducing sugar react non-enzymatically with amine groups of amino acids in proteins, lipids or DNA.^[10,11] They are highly reactive mediator that causes mitochondrial DNA (mtDNA) common deletion (CD), alters the structure and function of molecules and increases oxidative stress in biological system.^[1,12]

Prasad *et al.*^[2] reviewed that any intervention that inhibits formation of AGEs is capable of putting generation of other oxidants (ROS and RNS) under check and consequently reduce

oxidative stress. However many researchers have suggested that the best way to neutralize free-radical mediated oxidative stress is to ingest diet rich in phytochemicals with antioxidant properties or to take dietary supplements of antioxidants.^[13-16] These are widely found in nature, especially in plant products and are an extremely diversified group of chemicals.^[17-21] One such naturally occurring antioxidant is flavonoids.^[22-24] Flavonoids reportedly activate key enzymes in mitochondrial respiration and therefore protect cells by acting as antioxidants to break the vicious cycle of oxidative stress and tissue damage.^[25] Flavonoids have been confirmed to be abundant in *Solanum macrocarpon* leaves,^[13,26] an indigenous vegetable domesticated in Africa.^[27-29] The uses of *Solanum macrocarpon* leaves in indigenous medicine ranges from weight reduction to treatment of several ailments including asthma, allergic rhinitis, nasal catarrh, skin infections, rheumatic disease and swollen joint pains, gastro-esophageal reflux disease, constipation and dyspepsia.^[30,31]

Many flavonoid-rich extracts has been reported to revealed renal and hepato-protective effects in experimental animals.^[32,33] However, few reports were available on the protective potentials of flavonoid-rich extract of *Solanum macrocarpon* leaves against experimentally induced oxidative stress in rats. Therefore, the present study was carried out to assess the protective potentials of Flavonoid-Rich Extract of *Solanum macrocarpon* Leaves (FRESML) against D-galactose-induced oxidative stress in rats.

MATERIALS AND METHODS

Materials

Basic laboratory materials such as UV-Visible Spectrophotometer, Centrifuge Machine, Thermostatic Water Bath, Mortar and Pestle, Stop Watch, weighing balance, pH meter, micropipette and glasswares were used for this work. Reagents includes: methanol, D-galactose, Tris HCl, KCl, Distilled water, Corn Oil, Magnesium ribbon, Adrenaline, H₂O₂, Potassium dichromate, Glacial acetic acid, Thiobarbituric acid, Trichloroacetic acid and Ellman's reagent. All other reagents were of the best pure analytical grade available and were obtained from Sigma Aldrich Chemical Company, St. Louis, Missouri, USA.

Sample Collection and Preparation

Solanum macrocarpon plants were collected from the vegetable garden in Ote Village, via Ilorin, Kwara State, Nigeria and were identified and authenticated with voucher herbarium number LHO 481 issued by Prof. Ogunkunle, a taxonomist at the Department of Pure and Applied Biology, Ladoke Akintola University of Technology, Ogbomoso. Fresh leaves of the

plant were air-dried at room temperature and further powdered. About 625 grams of the powdered leaves was extracted in 6.25 Litres of methanol and kept in the dark for 72 hours. The solvent was concentrated at temperature below 40⁰C and the resulting methanol extract was subjected to liquid-liquid chromatographic separation technique using n-hexane, chloroform and ethyl acetate as the solvent to obtain their corresponding fractions.^[34] The three fractions were screened for total flavonoids, and the ethyl acetate fraction was found to be the most potent, thereafter it was used for the biological study.

Experimental Animals

Twenty eight male albino rats of Wistar strain, weighing averagely 120g were used for this study. Animals were randomly selected into four groups of seven per group and treated for 28 days. Group A (control) were given distilled water, Group B (extract only) were administered 100mg/kg/day of FRESML, Group C (D-galactose only) were administered 500mg/kg/day of D-galactose and Group D (D-galactose + extract) received combined administration of 500mg/kg/day of D-galactose and 100mg/kg/day of FRESML. The animals were sacrificed 24 hours after the last administration using a mild anaesthetizing solvent. Established international guiding principles for using animals in biomedical research was adopted for the experiments.

Phytochemical Evaluation

Total Flavonoids content was determined by magnesium hydrochloride reduction test and sodium hydroxide test described by Trease and Evans.^[35]

Biochemical Evaluation

Tissues and serum Total Protein concentration was determined by the Biuret method.^[36] ALT and AST activities were determined by the method of Reitman and Frankel.^[37] GGT activity was determined by the method of Szasz.^[38] ALP activity was determined by optimized standard method according to the recommendation of the Deutsche Gesellschaft fur Klinische Chemie.^[39] Urea concentration was determined by Urease-Berthelot method. Creatinine concentration was determined by modified method of Jaffe.^[40,41] SOD activity was determined by the method of Misra and Fridovich.^[42] Catalase activity was determined by the method of Sinha.^[43] GSH concentration was determined by the modified method of Ellman.^[44,45] MDA concentration was determined by the method reported by Varshney and Kale.^[46] PCV was estimated using micro haematocrit.^[47] WBC was counted using improved Neubaur haemocytometer. Fibrinogen concentration was measured by the Clauss method.^[48]

Statistical Analysis

Results are presented as Mean \pm SEM. Paired Student's t-test was used to compare variations amongst groups. The minimum level of significance was considered at $p < 0.05$. Statistical analysis was carried out using a software program (GraphPad Prism Ver. 5; GraphPad Software, San Diego, CA).

RESULTS AND DISCUSSION

Oxidative stress is associated with decline in synthesis rate of muscle proteins, such as myosin heavy chain^[49] and mitochondrial protein.^[50] The statistically significant decreased in serum (Fig. 1), liver (Fig. 2) and kidney (Fig. 3) total protein concentrations in the D-galactose treated group further emphasized the fact that total protein concentration decreases in oxidative stress condition. These results (Fig. 1, Fig. 2 and Fig. 3) are in agreement with the results of others which reported evidences of generalized decline of whole body protein turnover and mixed muscle proteins respectively.^[49,51-53] Therefore, generalized decline in the remodelling process of tissues with oxidative stress may be responsible for the significant decrease in total protein reported in this study. In support of this, a previous study explained that the significant decrease in total protein with oxidative stress may indicate an impaired liver synthesis and excretory functions⁵⁴. Interestingly, combined administration of D-galactose and FRESML brought about increased serum (Fig. 1), hepatic (Fig. 2) and renal (Fig. 3) total protein concentrations. This may be due to the ability of the flavonoid contents of the extract to improve the synthesis and excretory functions of the tissues.^[54]

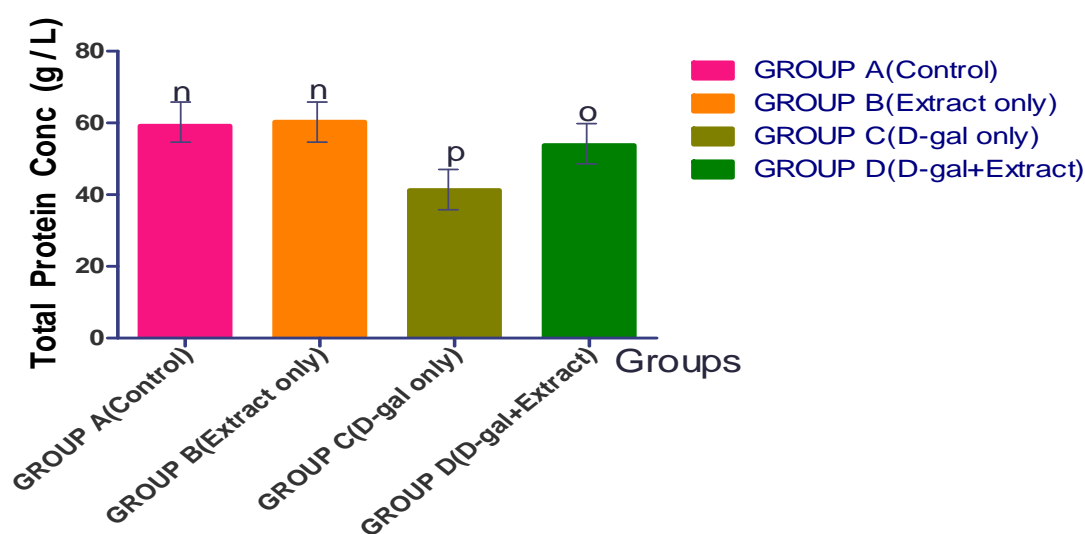


Fig. 1: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on serum total protein concentration in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

n = significantly different from **C&D** ($p < 0.05$)

p = significantly different from **A, B&D** ($p < 0.05$)

o = significantly different from **A, B&C** ($p < 0.05$)

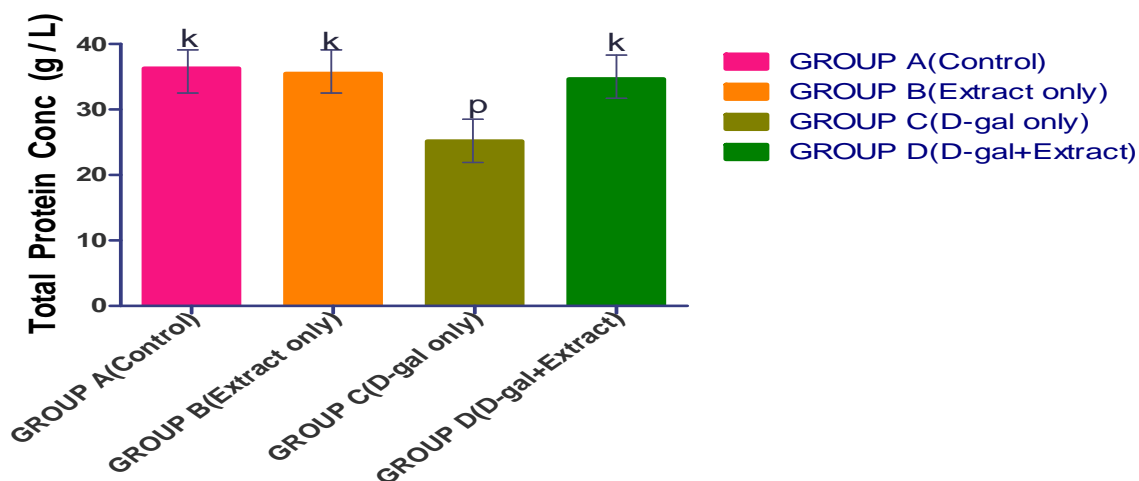


Fig. 2: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on liver total protein concentration in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

k = significantly different from **C** ($p < 0.05$)

p = significantly different from **A, B&D** ($p < 0.05$)

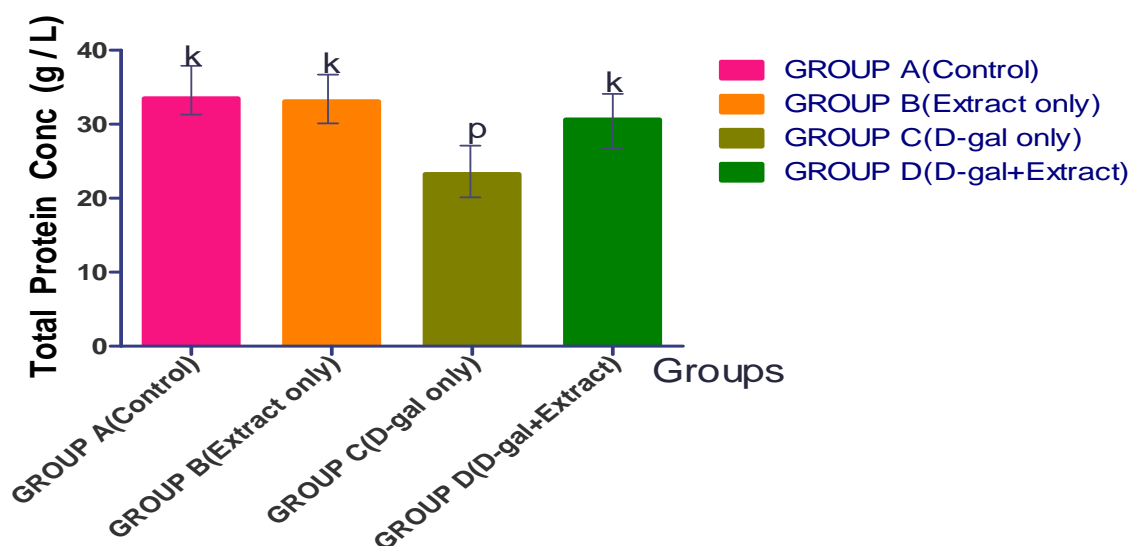


Fig. 3: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on kidney total protein concentration in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

k = significantly different from **C** ($p < 0.05$)

p = significantly different from **A, B&D** ($p < 0.05$)

The study of different enzyme activities such as ALT, AST and GGT have been found to be of great value in the assessment of clinical and experimental liver damage.^[55,56] In the present investigation, it was observed that the animals treated with D-galactose resulted in elevated levels of serum ALT (Fig. 4), AST (Fig. 5) and GGT (Fig. 6). These changes in the marker levels may affect the hepatic structural integrity. The rise in the AST (Fig. 5) is usually accompanied by an elevation in the level of ALT (Fig. 4), which play a vital role in the conversion of amino acids to keto acids.^[57] The combined administration of D-galactose and FRESML significantly attenuated the elevated levels of these serum markers (Fig. 4, Fig. 5 and Fig. 6). The normalization of these serum markers by FRESML suggests that the extract was able to condition the hepatocytes so as to protect the membrane integrity against oxidative stress that facilitated leakage of these marker enzymes into the blood circulation from the liver.^[58,59]

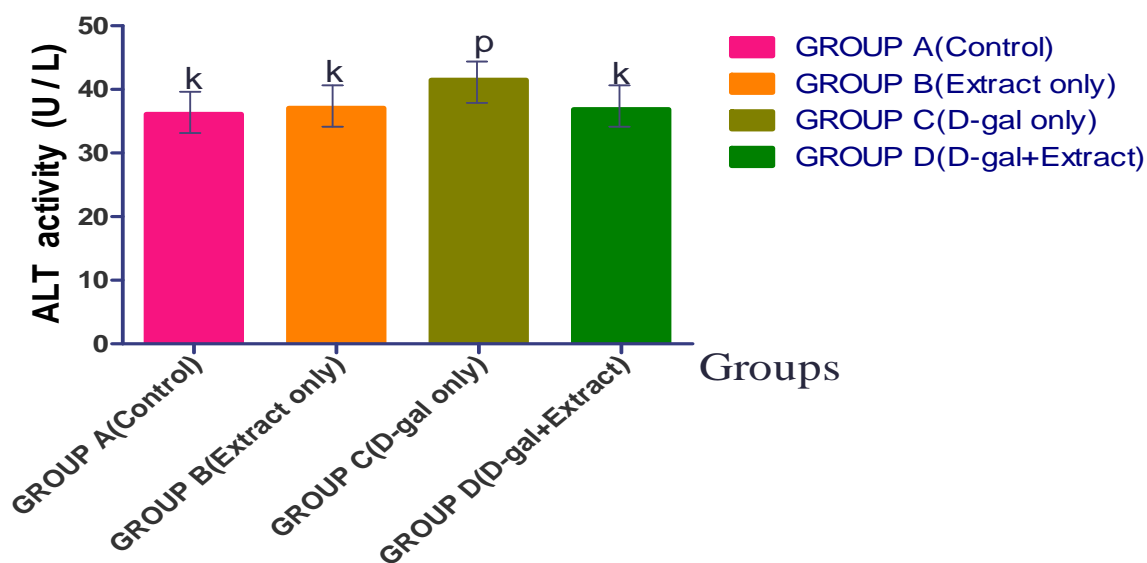


Fig. 4: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on serum ALT activities in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

k = significantly different from **C** ($p < 0.05$)

p = significantly different from **A, B&D** ($p < 0.05$)

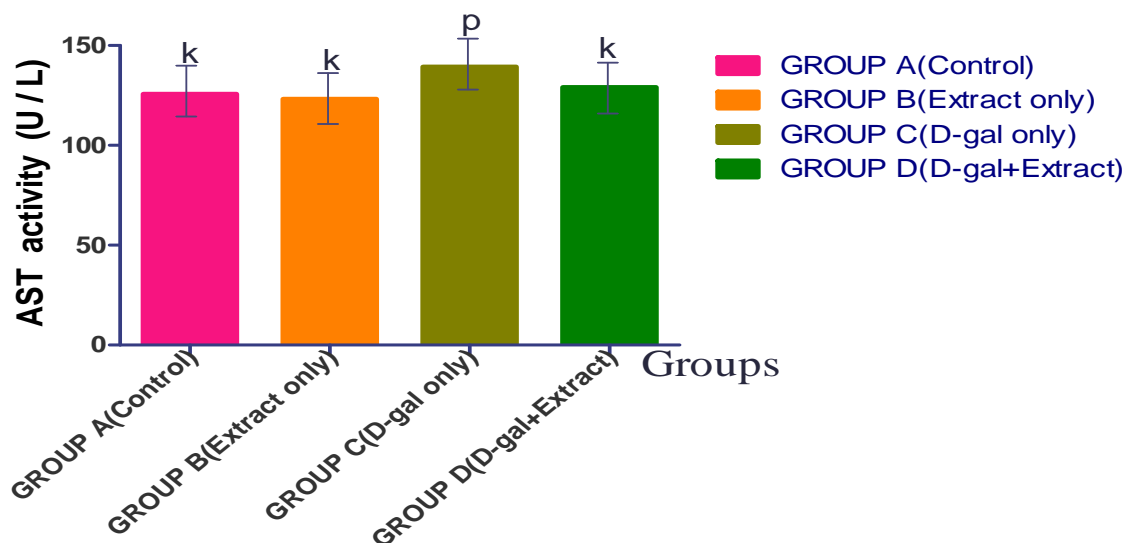


Fig. 5: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on serum AST activities in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

k = significantly different from **C** ($p < 0.05$)

p = significantly different from **A, B&D** ($p < 0.05$)

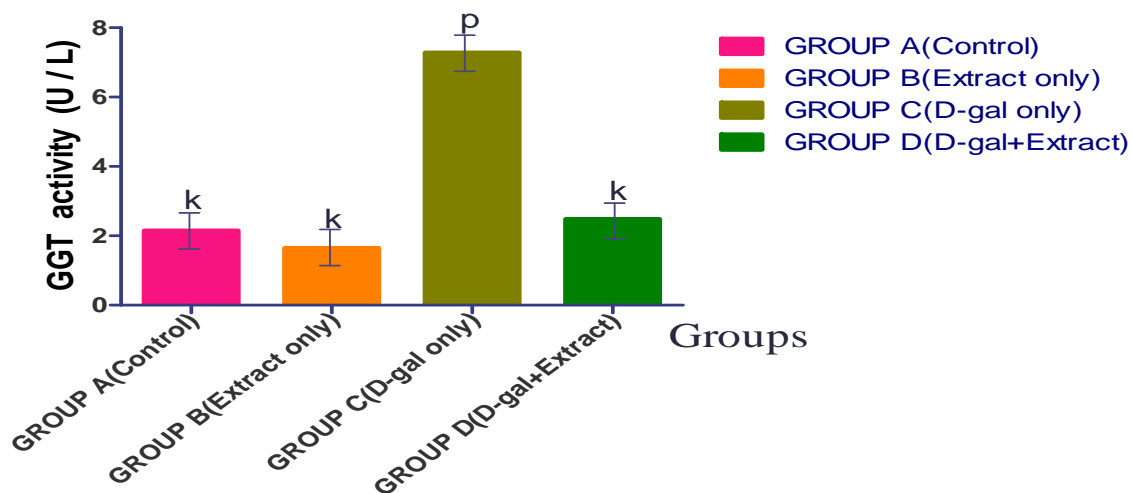


Fig. 6: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on serum GGT activities in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

k = significantly different from **C** ($p < 0.05$)

p = significantly different from **A, B&D** ($p < 0.05$)

The elevated levels of serum alkaline phosphatase (Fig. 7), urea (Fig. 8) and creatinine (Fig. 9) obtained in the D-galactose treated group in this study are suggestive of abnormal renal function caused by oxidative stress induced by D-galactose administration, especially as it relate to glomerular functions. Bishop *et al.*^[60] observed an increase in blood urea, creatinine and ALP levels which was closely correlated with histopathological degenerative changes in the kidney and these changes caused disturbance in the transport system of biochemical constituents. Urea is the principal end product of protein catabolism.^[60] Therefore its elevation in the serum (Fig. 8) may be due to enhanced protein catabolism and accelerated amino acid deamination for gluconeogenesis as most conditions that increases serum urea brings about decreases in serum total protein.^[61] Creatinine is the last variable of non-protein nitrogenous blood constituents.^[62] Low clearance rates of urea and creatinine (Fig. 8 and Fig 9) may indicate diminished ability of the kidneys to filter these waste products from the blood and excrete them in the urine.^[63] However, the ability of FRESML to restore the levels of these parameters to nearly control level (Fig. 7, Fig. 8 and Fig. 9) in the combined treatment group suggests the ability of the extract to prevent amino acid deamination (nephroprotective) and could be credited to its antioxidant activities.^[64]

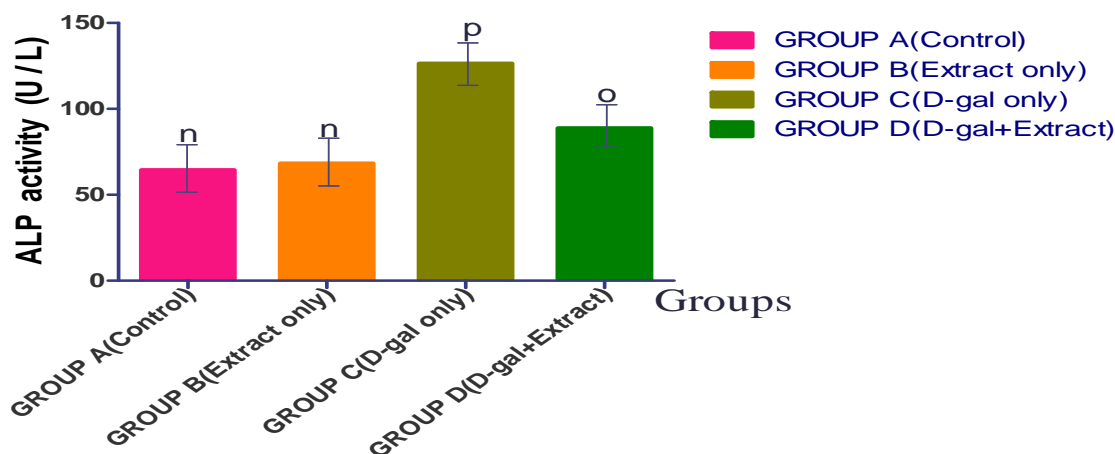


Fig 7: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on serum ALP activities in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

n = significantly different from **C&D** ($p < 0.05$)

p = significantly different from **A, B&D** ($p < 0.05$)

o = significantly different from **A, B&C** ($p < 0.05$)

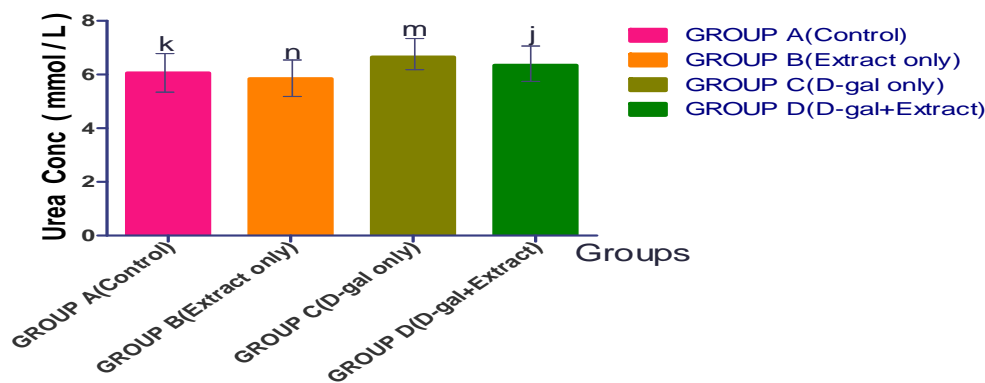


Fig. 8: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on serum urea concentration in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

k = significantly different from C ($p < 0.05$)

n = significantly different from C&D ($p < 0.05$)

m = significantly different from A&B ($p < 0.05$)

j = significantly different from B ($p < 0.05$)

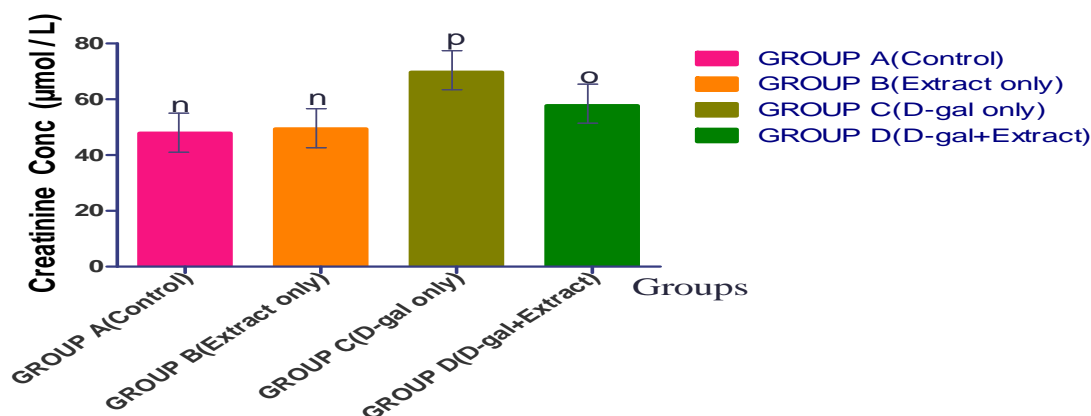


Fig. 9: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on serum creatinine concentration in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

n = significantly different from C&D ($p < 0.05$)

p = significantly different from A, B&D ($p < 0.05$)

o = significantly different from A, B&C ($p < 0.05$)

It has been reported that animal body had an effective mechanism to prevent the free radical induced tissue damage; this is accomplished by a set of endogenous antioxidant enzymes and proteins such as SOD, Catalase and GSH.^[65] In the present study, SOD and catalase activities

as well as GSH concentration were measured in liver (Fig. 10, Fig 12 and Fig. 14) and kidney (Fig. 11, Fig. 13 and Fig. 15). The depletion of SOD activity to nearly half of the control level in liver and kidney as seen in D-galactose treated group is another indication of oxidative stress induced by D-galactose. In the same manner, the depletion of Catalase activity to nearly quarter of the control level and half of the control level in liver and kidney respectively in D-galactose treated group also confirmed hepatotoxicity and nephrotoxicity of D-galactose. The reduction in GSH levels in the liver and kidney of the animals administered D-galactose alone by three-quarter of its concentration compared to the control group, further established the hepatotoxicity and nephrotoxicity of D-galactose. The decreases in these antioxidant parameters may be due to rapid consumption and exhaustion of storage of the enzymes and protein in fighting free radicals generated during development of oxidative stress.^[65]

Significant increases ($p < 0.05$) in the hepatic and renal levels of SOD, Catalase and GSH observed in the combined treatment group and extract only group showed that FRESML may reduce reactive free radicals and boost the antioxidant status. Similar claims were made by Iweala and Ogidigo,^[66] where they attributed the increase of these antioxidant indices by *Solanum macrocarpon* leaves extract to the abundant antioxidant polyphenolic compounds in the extract. However, they could not ascertain the specific class of polyphenolic compound responsible for this action. Having used Flavonoids-Rich Extract of *Solanum macrocarpon* Leaves in this study, it may be suggested that the flavonoids content of the extract was responsible for the ability of the extract to increase the levels of SOD, Catalase and GSH *in vivo*.

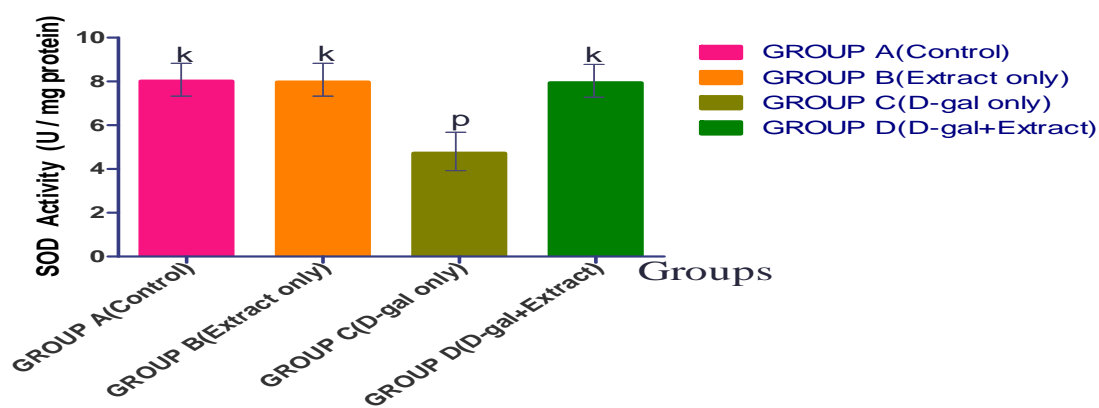


Fig. 10: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on liver SOD activities in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

k = significantly different from **C** ($p < 0.05$)

p = significantly different from **A, B&D** ($p < 0.05$)

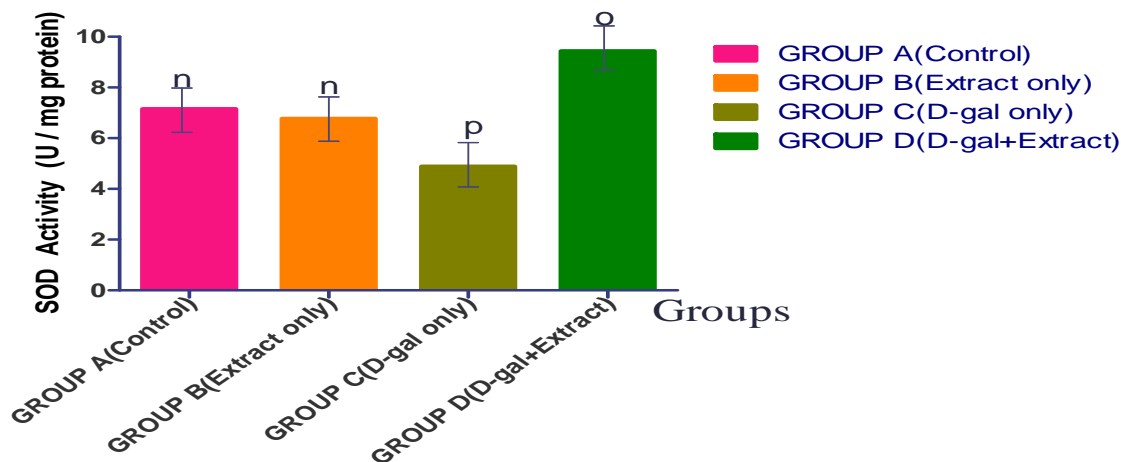


Fig. 11: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on kidney SOD activities in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

n = significantly different from **C&D** ($p < 0.05$)

p = significantly different from **A, B&D** ($p < 0.05$)

o = significantly different from **A, B&C** ($p < 0.05$).

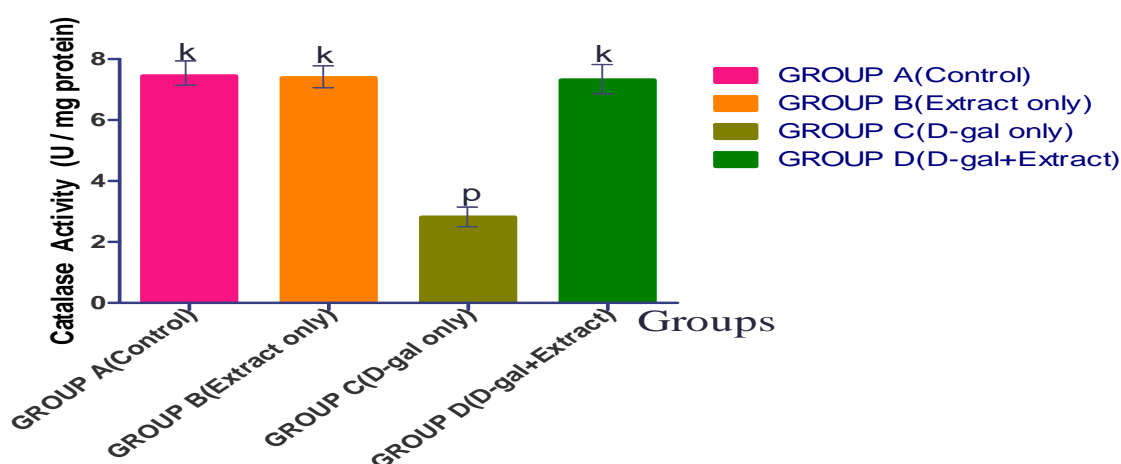


Fig 12: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on liver catalase activities in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

k = significantly different from **C** ($p < 0.05$)

p = significantly different from **A, B&D** ($p < 0.05$)

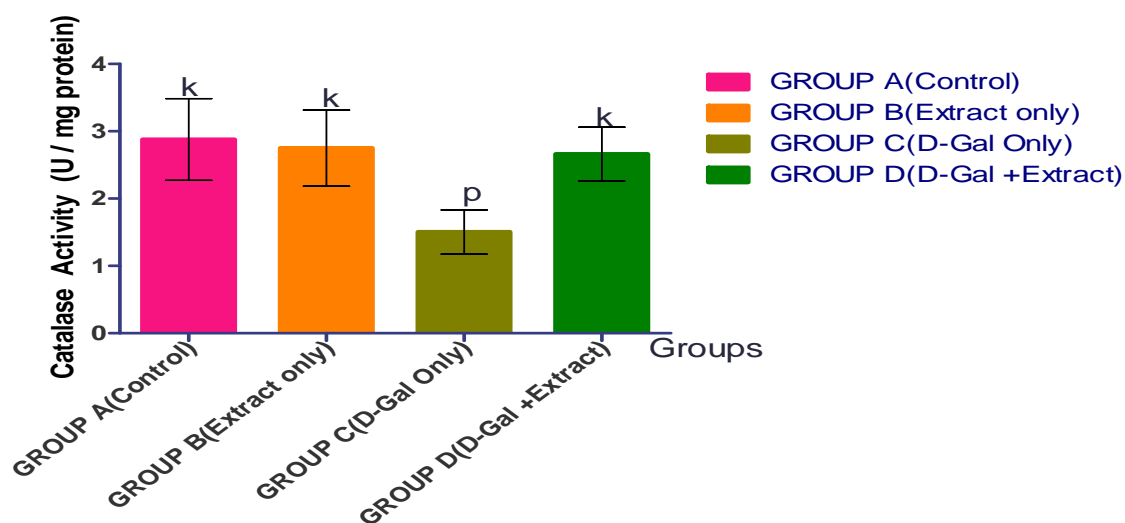


Fig. 13: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on kidney catalase activities in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

k = significantly different from **C** ($p < 0.05$)

p = significantly different from **A, B&D** ($p < 0.05$)

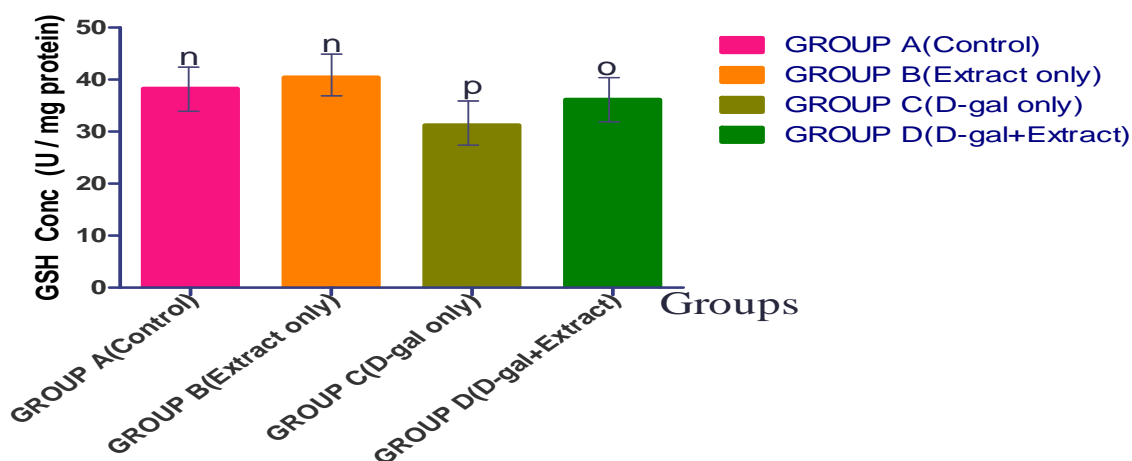


Fig. 14: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on liver GSH concentration in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

n = significantly different from **C&D** ($p < 0.05$)

p = significantly different from **A, B&D** ($p < 0.05$)

o = significantly different from **A, B&C** ($p < 0.05$)

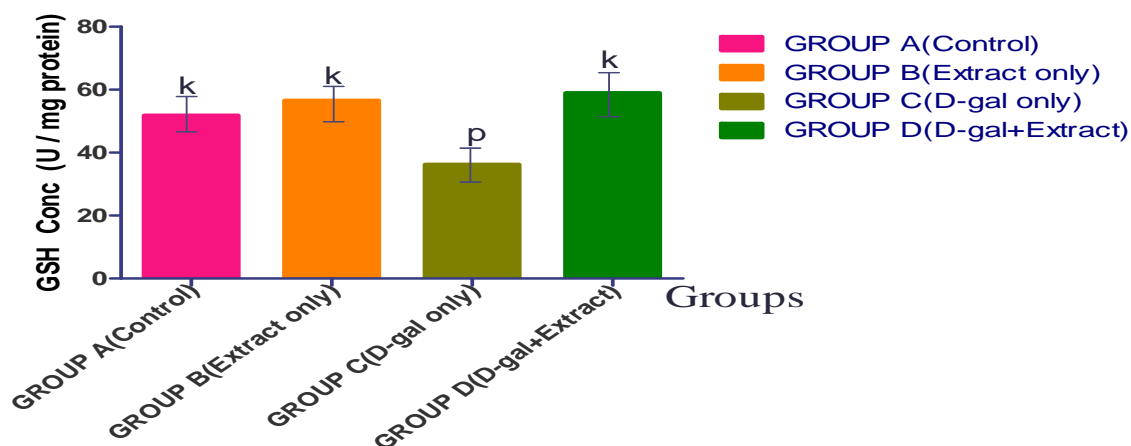


Fig. 15: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on kidney GSH concentration in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

k = significantly different from C ($p < 0.05$)

p = significantly different from A, B&D ($p < 0.05$)

Lipid peroxidation has been implicated in processes such as carcinogenesis, inflammation and aging.^[67] In the present study elevation in the levels of end products of lipid peroxidation, Malondialdehyde (MDA) in the liver (Fig. 16) and kidney (Fig. 17) of rats treated with D-galactose was observed. The increase in Malondialdehyde (MDA) levels in the liver and kidney suggests enhanced lipid peroxidation leading to tissue damage and failure of antioxidant defence system to prevent formation of excessive free radicals. Combined treatment of D-galactose and FRESML significantly reversed these changes. Hence it may be possible that the mechanism of liver protection and renal protection by FRESML may be due to its antioxidant effects.^[26,68-70]

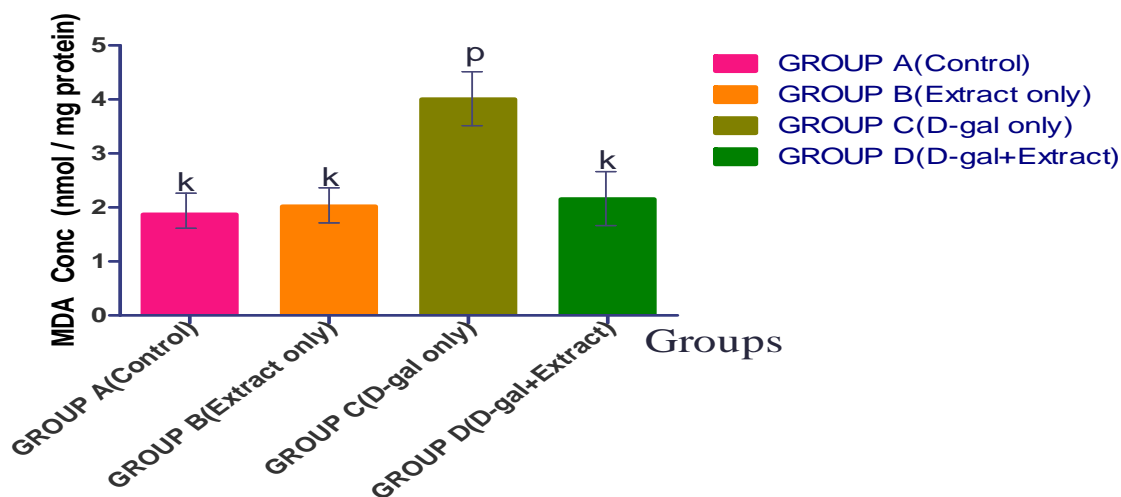


Fig. 16: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on liver MDA concentration in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

k = significantly different from C ($p < 0.05$)

p = significantly different from A, B&D ($p < 0.05$)

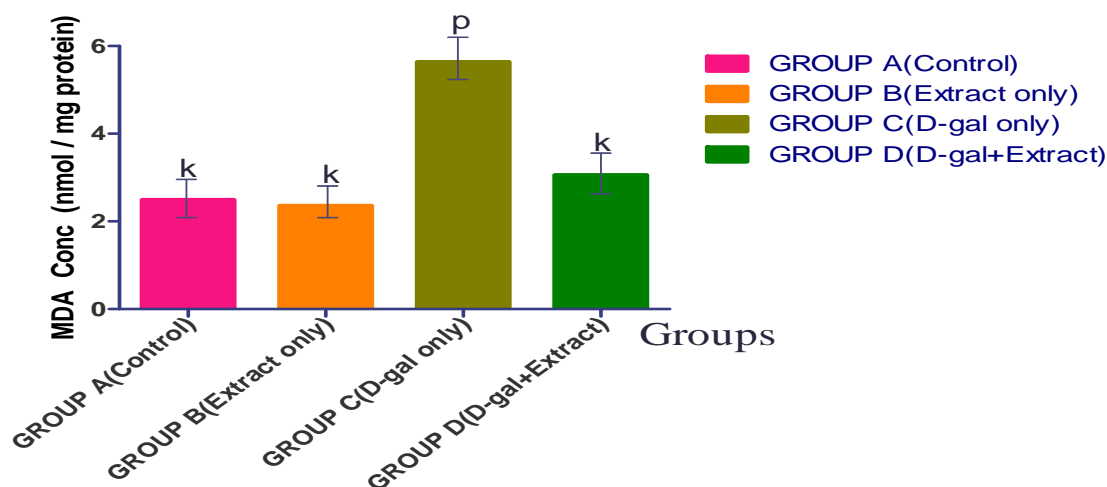


Fig. 17: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on kidney MDA concentration in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

k = significantly different from C ($p < 0.05$)

p = significantly different from A, B&D ($p < 0.05$)

Both haematological and inflammatory indices are needed to characterize tissues' response to injurious stimuli.^[71] The relative increase in percentage PCV of the animals treated with D-

galactose compared to the control group in this study (Fig. 18) signified acute systemic inflammation typical of oxidative stress in animal model.^[72] Evidence of increased PCV in oxidative stressed animals has been documented in literature.^[72] However, increased PCV has been strongly correlated with increased fibrinogen in the plasma.^[73,74] Therefore the increased percentage Packed Cell Volume witnessed in the D-galactose treated group in this study may be attributed to the increase in fibrinogen concentration in the rats. Interestingly, the increased PCV was adequately attenuated in the combined treatment group as proved by significant decrease in the percentage PCV of the animals in this group to nearly control level (Fig. 18). This correction may be as a result of FRESML ability to regulate haematopoiesis in bone marrow with a subsequent modulation of PCV as shown in this study.^[75]

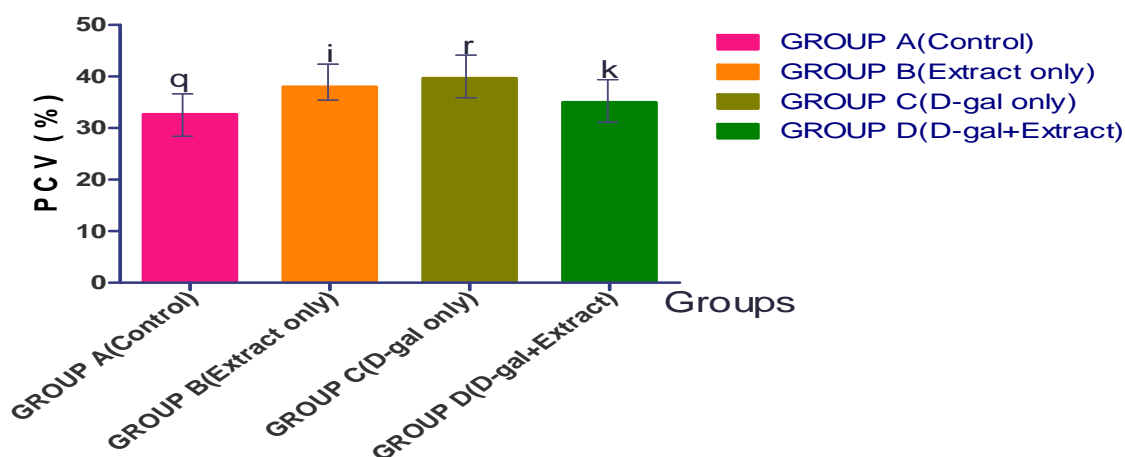


Fig. 18: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on blood PCV levels in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

q = significantly different from **B&C** ($p < 0.05$)

i = significantly different from **A** ($p < 0.05$)

r = significantly different from **A&D** ($p < 0.05$)

k = significantly different from **C** ($p < 0.05$)

In the present study, there was a significant increase in white blood cell count in D-galactose treated group compared to the control group and the extract only group (Fig 19). Mansour *et al.*^[76] revealed that increased WBC counts can occur as a result of immune response to an infection, cancer, or toxic chemical. Significant reduction of WBC counts in groups B (extract only group) and D (D-galactose + extract group) nearly to the control level (Fig 19) showed that the immune system may not be challenged in animals fed with FRESML. This

may be due to its content of immune-boosting flavonoids as flavonoids have been reported to have immune-boosting ability as a result of its antioxidant activities.^[77,78] This result (Fig 19) agreed with previous studies that *Solanum macrocarpon* may help preserve the body's adaptive response to stress.^[79]

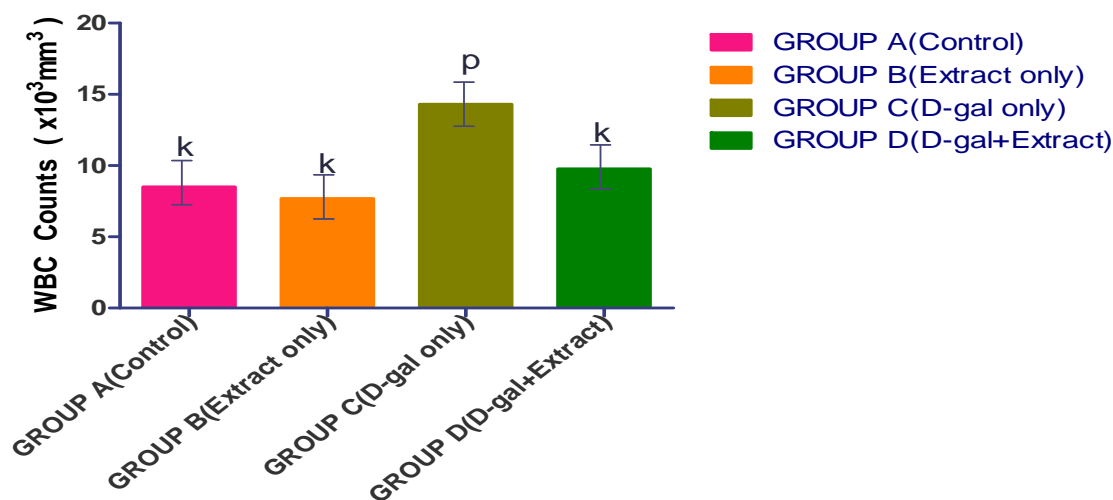


Fig. 19: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on plasma WBC counts in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

k = significantly different from C ($p < 0.05$)

p = significantly different from A, B&D ($p < 0.05$)

Inflammation is a protective attempt by the organism to remove the injurious stimuli and initiate the healing process.^[71] Fibrinogen concentration has previously been found to be particularly useful in detecting inflammatory diseases.^[80] Fibrinogen concentration rises with systemic inflammation.^[81,82] The significant increase in the plasma concentration of fibrinogen observed in D-galactose treated group in this study (Fig. 20) may be because its concentration increases gradually in response to tissue injury.^[83,84] However, the feature of inflammation disappeared in the combined treatment group of D-galactose and FRESML as the plasma fibrinogen concentration was restored nearly to the control levels (Fig. 20). This improvement noticed after FRESML administration might be attributed to the anti-inflammatory properties of the extract and strong antioxidant effect of flavonoids in *Solanum macrocarpon* extract on hematopoietic cells.^[75]

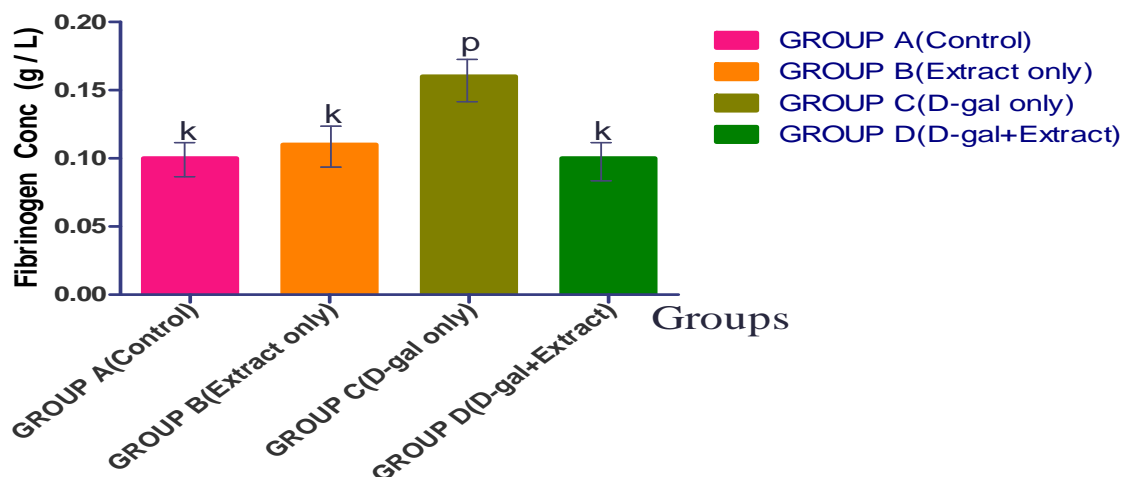


Fig. 20: Effects of flavonoid-rich extract of *Solanum macrocarpon* leaves on plasma fibrinogen concentration in D-galactose-treated rats.

Each value is Mean \pm S.E.M of 7 rats in each group.

k = significantly different from C ($p < 0.05$)

p = significantly different from A, B&D ($p < 0.05$).

CONCLUSION

The results obtained in this study demonstrated that combined treatment of D-galactose and Flavonoid-Rich Extract of *Solanum macrocarpon* Leaves showed significant improvement in protein synthesis, hepatic integrity, renal integrity and antioxidant status compared with D-galactose exposed rats. Therefore, Flavonoid-Rich Extract of *Solanum macrocarpon* Leaves could be seen as a good source of useful chemoprotective agents or food supplements and a possible template for discovery of drugs for ameliorating oxidative stress and other diseases associated with it.

ACKNOWLEDGEMENTS

The authors are grateful to the Postgraduate School and the Department of Biochemistry of Ladoké Akintola University of Technology, Ogbomoso, Nigeria for the supports received for the work.

CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

REFERENCES

1. Salganik RI. Apoptosis and other protective mechanisms in cancer patients and the human population. *J Am Coll Nutr*, 2001; 20: 464S–472S.
2. Prasad C, Imrhan V, Marotta F, Juma S, Vijayagopal P. Lifestyle and Advanced Glycation End Products (AGEs) Burden: Its Relevance to Healthy Aging. *Aging and Disease*, 2014; 5(3): 212-217.
3. Proctor PH. Free Radicals and Human Disease. *CRC Handbook of Free Radicals and Antioxidants*, 1989; 1: 209-221.
4. Dillin A, Goltschling DE, Nystrom T. The Good and the Bad of being Connected: the Integrons of Aging. *Curr Opin Cell Biol*, 2014; 26: 107-112.
5. Zima T, Fialova L, Mestek O, Janebova M, Crkovska J, Malbohan I, Stipek S, Mikulikova L, Popov P. Oxidative stress, metabolism of ethanol and alcohol related diseases. *J Biomed Sci*, 2001; 8: 59-70.
6. Astley SB. Dietary antioxidants past, present and future. *Trend Food Sci Technol*, 2003; 14: 93-98.
7. Munch G, Gerlach M, Sian J, Wong A, Riederer P. Advanced glycation end products in neurodegeneration: more than early markers of oxidative stress? *Ann Neurol*, 1998; 44: S85–S88.
8. Dammann P, Sell DR, Begall S, Strauch C, Monnier VM. Advanced glycation end-products as markers of aging and longevity in the long-lived Ansell's mole-rat (*Fukomys anelli*). *J Gerontol A Biol Sci Med Sci*, 2012; 67: 573–583.
9. Rolewska P, Al-Robaiy S, Santos AN, Simm A, Silber RE, Bartling B. Age-related expression, enzymatic solubility and modification with advanced glycation end-products of fibrillar collagens in mouse lung. *Exp Gerontol*, 2013; 48(1): 29-37.
10. Lee AT, Cerami A. Nonenzymatic glycosylation of DNA by reducing sugars. *Prog Clin Biol Res*, 1989; 304: 291–299.
11. Brownlee M. Negative consequences of glycation. *Metabolism*, 2000; 49: 9-13.
12. Bei C, Yi Z, Wei P, Yu S, Yu JH, Yang Y, Wei JK. Increase mitochondrial DNA damage and decrease base excision repair in the auditory cortex of D-galactose-induced aging rats. *Mol Biol Rep*, 2011; 38(6): 3635-3642.
13. Salawu SO, Akindahunsi AA, Comuzzo P. Chemical Composition and In-vitro Antioxidant Activities of some Nigerian Vegetables. *J Pharmacol Toxicol*, 2006; 1(5): 429-437.

14. Badmus JA, Adedosu OT, Fatoki JO, Adegbite VA, Adaramoye OA, Odunola OA. Lipid Peroxidation Inhibition and Antiradical Activities of some Leaf Fractions of *Mangifera indica*. Acta Poloniae Pharmaceutica – Drug Research, 2011; 68(1): 23-29.
15. Adedosu OT, Adekunle AS, Adedeji AL, Afolabi OK, Oyedeji TA. Antioxidant and Anti-Lipidperoxidation Potentials of the Ethylacetate and Chloroform Extracts of *Basella Alba* Leaves. Asian Journal of Natural & Applied Sciences, 2013; 2(2): 81-88.
16. Adedosu OT, Badmus JA, Adeleke GE, Afolabi OK, Adekunle AS, Fatoki JO, Fakunle PB. Oxidative Damage and Dietary Antioxidants: The Roles of Extract and Fractions of *Solanum aethiopicum* Leaves. Canadian Journal of Pure and Applied Sciences, 2015; 9(1): 3185-3192.
17. Liu RH. Potential synergy of phytochemicals in cancer prevention: mechanism of action. The American society for nutritional sciences, Journal of Nutrition, 2004; 134: 3479-3485.
18. Hung SH, Yu CW, Lin CH. Hydrogen peroxide functions as a stress signal in plants. Botanical Bulletin of Academia Sinica, 2005; 46: 1-10.
19. Materska M. Quercetin and its Derivatives: Chemical Structure and Bioactivity - A Review. Pol J Food Nutr Sci, 2008; 58(4): 407-413.
20. Vasu K, Goud JV, Suryam A, Singara CMA. Biomolecular and phytochemical analyses of three aquatic angiosperms. Afr J Microbiol Res, 2009; 3(8): 418-421.
21. Adewale OB, Onasanya A, Fadaka AO, Iwere H, Anadozie SO, Osukoya OA, Olayide II. In-vitro antioxidant effect of aqueous extract of *Solanun macrocarpon* leaves in rat liver and brain. Oxid Antioxid Med Sci, 2014; 3(3): 225-229.
22. Othman A, Ismail A, Ghani NA, Adenan I. Antioxidant capacity and phenolic content of cocoa beans. Food Chem, 2007; 100: 1523-1530.
23. Ghasemzadeh A, Ghasemzadeh N. Flavonoids and Phenolic acids: Role and Biochemical Activity in Plants and Humans. J Med Plants Res, 2011; 5(31): 6697-6703.
24. Bigoniya P, Singh K. Ulcer protective potential of standardized hesperidin, a citrus flavonoid isolated from *Citrus sinensis*. Rev Bras Farmacogn, 2014; 24: 330-340.
25. Schmitt-Schillig S, Schaffer S, Weber CC, Eckert GP, Muller WE. Flavonoids and the Aging Brain. J Physiol Pharmacol, 2005; 56(1): 23-36.
26. Olajire AA, Azeez L. Total antioxidant activity, phenolic, flavonoid and ascorbic acid contents of Nigerian vegetables. Afr J Food Sci Technol, 2011; 2(2): 22-29.
27. Bukenya-Ziraba R. Studies in the Taxonomy of *Solanum* L. in Southern Ghana. Msc Thesis, University of Ghana, Ghana, 1980; Pp 194.

28. Bukenya-Ziraba R. Studies in the Taxonomy of Genus *Solanum* in Uganda. PhD Thesis, Makerere University, Kampala, Uganda, 1993; Pp 456.
29. Bonsu KO, Fontem DA, Nkansah GO, Iroume RN, Owusu EO, Schippers RR, Diversity within the Gboma eggplant (*Solanum macrocarpon*), an indigenous vegetable from West Africa. *Ghana J Horticulture*, 2002; 1: 50-58.
30. Dalziel JM. The Useful Plants of West Tropical Africa. New York: Longman 1st edn., 1937; 433–435.
31. Bello SO, Muhammad BY, Gammaniel KS, Abdu-Aguye I, Ahmed H, Njoku CH, Pindiga UH, Salka AM. Preliminary Evaluation of the Toxicity and Some Pharmacological Properties of the Aqueous Crude Extract of *Solanum melongena*. *Res J Agric Biol Sci.*, 2005; 1(1): 1-9.
32. Wang N, Li PB, Wang YG, Peng W. Hepatoprotective effect of *Hypericum japonicum* extract and its fractions. *J Ethnopharmacol.*, 2008; 116: 1-6.
33. Yuan LP, Chen FH, Ling L, Dou PF, Bo H, Zhong MM, Xia LJ. Protective effects of total flavonoids of *Bidens pilosa* L. (TFB) on animal liver injury and liver fibrosis. *J Ethnopharmacol.*, 2008; 116: 539-546.
34. Ogundipe OO, Moody JO, Houghton PJ, Odelola HA. Bioactive chemical constituents from *Alchornea Laxiflora* (benth) pax and Hoffman. *J of Ethnopharmacology*, 2000; 74: 275-280.
35. Trease GE, Evans WC. *Pharmacognosy*, 15th Ed. Samders Publishers, London, 2002: Pps 42-44, 221-229, 246-249, 304-306, 331-332, 391-393.
36. Gornall AG, Bardwill CJ, David MM. Determination of serum protein by means of Biuret reaction. *J Biol Chem.*, 1949; 177: 751-766.
37. Reitman S, Frankel S. A colorimetric method for the determination of serum glutamic oxaloacetic and glutamic pyruvic transaminase. *Am J Clin Pathol.*, 1957; 28: 56-63.
38. Szasz G. A kinetic photometric method for serum γ -glutamyl transpeptidase. *Clin Chem.*, 1969; 15: 124-136.
39. Deutsche Gesellschaft fur Klinische Chemie., *J Clin Chem Clin Biochem.*, 1972; 10: 182.
40. Jaffe M. Ueber den Niederschlag welchen Pikrinsaure in normalen Harn ergeugt und uber eine neue reaction des kreatinins. *Z Physiol Chem.*, 1886; 10: 391-400.
41. Bartels H, Bohmer M, Heierli C. Serum creatinine determination without protein precipitation. *Clin Chim Acta.*, 1972; 37: 193-197.
42. Misra HP, Fridovich I. The role of superoxide anion in the autoxidation of epinephrine and a simple assay for superoxide dismutase. *J Biol Chem.*, 1972; 247: 3170-3175.

43. Sinha AK. Colorimetric Assay of Catalase. *Analytical Biochemistry*, 1972; 47(2): 389-394.
44. Ellman GL. Tissue sulphhydryl groups. *Arch Biochem Biophys*, 1959; 82: 70-77.
45. Jollow DJ, Michell JR, Zampaglione N, Gillete JR. Bromobenzene-induced Liver necrosis: Protective role of glutathione and evidence for 3,4-Bromobenzene oxide as hepatotoxic metabolite. *Pharmacology*, 1974; 11: 151-169.
46. Varshney R, Kale RK. Effects of Calmodulin Antagonist on Radiation Induced Lipid Peroxidation in Microsomes. *Int J Radiat Biol.*, 1990; 58: 733-743.
47. Mc Inory RA. A micro hematocrit for determining the packed cell and haemoglobin concentration on capillary blood. *J Clin Pathol.*, 1954; 7: 32-33.
48. Kabiri N, Setorki M. Regression of hypercholesterolemic atherosclerosis in rabbits by hydroalcoholic extracts of *Hypericum perforatum*. *J Med Plants Res.*, 2012; 6(13): 2540-2549.
49. Balagopal P, Rooyackers OE, Adey DB, Ades PA, Nair KS. Effects of aging on in vivo synthesis of skeletal muscle myosin heavy-chain and sarcoplasmic protein in humans. *Am J Physiol.*, 1997; 273 (36): 790-800.
50. Rooyackers OE, Adey DB, Ades PA, Nair KS. Effect of age on in vivo rates of mitochondrial protein synthesis in human skeletal muscle. *Proc Natl Acad Sci USA*, 1996; 93: 15364-15369.
51. Kataria N, Sareen M, Kataria AK, Bhatia JS, Ghosal AK. Some observations on total serum proteins in camels. *Indian Vet Med J.*, 1991; 15: 38-43.
52. Welle S, Thornton C, Statt M, McHenry B. Postprandial myofibrillar and whole body protein synthesis in young and old human subjects. *Am J Physiol.*, 1994; 267(30): 599-604.
53. Ogundu UE, Okoro VMO, Okeke GU, Durugo N, Mbaebie GAC, Ezebuike CI. Effects of age, breed and sex on the serum biochemical values of Turkeys (*Meleagris gallopova*) in South-eastern Nigeria, *Afr J Agric Res.*, 2013; 8(23): 2825-2828.
54. Nematalla KH, Sahar M, Arafa A, Ghada MY, Zainb AS. Effect of Echinacea as Antioxidant on Markers of Aging, *Australian Journal of Basic and Applied Sciences*. 2011; 5(2): 18-26.
55. Vaishwanar I, Kowale CN. Effect of two ayurvedic drugs Shilajeet and Eclinol on changes in liver and serum lipids produced by carbontetrachloride. *Ind J Exp Biol.*, 1976; 14: 58-61.

56. Venkateswaran S, Pari L, Viswanathan P. Anti-peroxidation effect of Livex, a herbal formulation against erythromycin estolate induced lipid peroxidation in rats. *Phytother Res.*, 1998; 12: 465-471.
57. Sallie R, Tredger JM, Willaiaam R. Drugs and the liver. *Biopharm Drug Dispos.*, 1999; 12: 251-259.
58. Balouchzadeh A, Rahimi HR, Ebadollahi-Natanzi AR, Minaei-Zangi B, Sabzevari O. Aqueous extract of Iranian green tea prevents lipid peroxidation and chronic ethanol liver toxicity in rat. *J Phamarcol Toxicol.*, 2011; 6: 691-700.
59. Adedosu OT, Badmus JA, Afolabi OK, Yakubu FO. Effect of methanolic leaf extract of *Ocimum gratissimum* (Linn) Leaves on Sodium Arsenite-Induced Toxicity in rats, *J Pharm and Toxicol.*, 2012; 7(5): 259-266.
60. Bishop LM, Fody PE, Schoe HL. *Clinical Chemistry: Principles, Procedures, Correlations*. 5th Edn., Lippincott Williams & Wilkins, Philadelphia, 2005; pp: 730, ISBN: 0781746116.
61. Varely H, Gowenlock AH, Bell M. *Practical Clinical Biochemistry. Hormones, Vitamins, Drugs and Poision*, 6th Edn., Heinemann Medical Books, London, 1987; pp: 477-549.
62. Usunobun U, Adegbeji JA, Okugbo T, Uduenevwo FE, Osibemhe M, Okolie NP. N-nitrosodimethylamine (NDMA), Liver function enzymes, Renal function parameters and Oxidative Stress Parameters: A Review. *Br J Pharmacol Toxicol.*, 2012; 3(4): 165-176.
63. Cameron JS. *Kidney Failure: The Facts*. Oxford University Press, New York. 1996.
64. Shereen BG, Doaa MZ. Beneficial Effects of Green Tea Extract on Liver and Kidney Functions, Ultrastructure, Lipid Profile and Hematological Parameters in Aged Male Rats. *Global Veterinaria*, 2013; 11(2): 191-205.
65. Noeman SA, Hamooda HE, Baalash AA. Biochemical Study of Oxidative Stress Markers in the Liver, Kidney and Heart of High Fat Diet Induced Obesity in Rats. *Diabetology & Metabolic Syndrome*, 2011; 3(17): 1-8.
66. Iweala EEJ, Ogidigo JO. Prostate Specific Antigen, Antioxidant and Hematological Parameters in Prostatic Rats Fed *Solanum macrocarpon* L. Leaves. *Asian Journal of Biological Sciences*, 2015; 8(1): 30-41.
67. Ames BN, Shigenaga MK, Hagen TM. Oxidants, antioxidants and the degenerative diseases of aging. *Proc Natl Acad Sci.*, 1993; 90: 7915-7922.
68. Shahidi F, Wanasundara PKJPD. Phenolic antioxidants. *Critical Reviews in Food Science and Nutrition*, 1992; 32: 67-103.

69. Cook NC, Samman S. Flavonoids- chemistry, metabolism, cardioprotective effects, and dietary sources. *Nutritional Biochemistry*, 1996; 7: 66-76.
70. Kessler M, Ubeaud G, Jung L. Anti- and pro-oxidant activity of rutin and quercetin derivatives. *J Pharm and Pharmacol.*, 2003; 55: 131-142.
71. Ferrero-Miliani L, Nielson OH, Andersen PS, Girardin SE. Chronic inflammation: importance of NOD2 and NALP3 in interleukin-1 β generation. *Clin Exp Immunol.*, 2007; 147(2): 227–235.
72. Adias TC, Ajugwo AO, Erhabor T, Nyenke CU. Effect of Pumpkin Extract (*Telfairia accidentalis*) on Routine Haematological Parameters in Acetone-Induced Oxidative Stress Albino Rats. *Am J of Food Sc. & Tech.*, 2013; 1(4): 67-69.
73. Ajmani RS, Rifkind JM. Hemorheological changes during human aging. *Gerontology*, 1998; 44: 111-210.
74. Fu A, Nair KS. Age effect on fibrinogen and albumin synthesis in humans. *Am J Physiol.*, 1998; 275: 1023-1030.
75. Monira AA, Nermin ME, Hamdy T. The Protective Role of Rosemary (*Rosmarinus officinalis*) in Lead Acetate Induced Toxicity in Rats. *J Appl Sci Res.*, 2012; 8(6): 3071-3082.
76. Mansour SA, Mossa AH, Heikal TM. Haematotoxicity of a new natural insecticide Spinosad on male Albino rats. *Int J Agric Biol.*, 2007; 9: 342-346.
77. Romieu I, Castro-Giner F, Kunzli N, Sunyer J. Air pollution, oxidative stress and dietary supplementation: A review. *Eur Respir J.*, 2008; 31: 179-197.
78. Bub A, Watzl B, Blockhaus M, Briviba K, Liegibel U. Fruit juice consumption modulates antioxidative status, immune status and DNA damage. *J Nutr Biochem.*, 2003; 14: 90-98.
79. Olajire AA, Azeez L. Effects of *Solanum macrocarpon* (African eggplant) on haematological parameters of wistar rats exposed to urban air pollution. *Adv Environ Res.*, 2012; 1: 109-123.
80. Wuijckhuise-Sjouke LA. Plasma fibrinogen as a parameter of the presence and severity of inflammation in horses and cattle. *Tijdschr Diergeneeskd*, 1984; 109: 869-872.
81. Allen BV, Kold SE. Fibrinogen response to surgical tissue trauma in horse. *Equine Vet J.*, 1988; 20: 441-443.
82. Pollock PJ, Prendergast M, Schumacher J. Effects of surgery on the acute phase response in clinically normal and diseased horses. *Vet Rec.*, 2005; 156: 538-542.

83. Jacobsen S, Jensen JC, Frei S. Use of serum amyloid A and other acute phase reactants to monitor the inflammatory response after castration in horses: A field study. *Equine Vet J.*, 2005; 37: 552-556.
84. Borges AS, Divers TJ, Stokol T, Mohammed OH. Serum Iron and Plasma Fibrinogen Concentrations as Indicators of Systemic Inflammatory Diseases in Horses. *J Vet Intern Med.*, 2007; 21: 489-494.