



PROTECTIVE ROLE OF COLOSTRUM AGAINST CCL₄ INDUCED LIVER DAMAGE IN RATS

Alaa M. Abd El –Fattah, Samia M. EL-Dieb*, Fouad M. F. Elshaghabee

Dairy Science Department, Faculty of Agriculture, Cairo University, Egypt.

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*Corresponding Author

Dr. Samia M. EL-Dieb

Dairy Science Department,
Faculty of Agriculture, Cairo
University, Egypt.

ABSTRACT

This study was carried out to evaluate the effect of cow and buffalo colostrum as a natural product on liver and kidney markers and antioxidant capacities after acute carbon tetrachloride (CCL₄) exposure in male Sprague-Dawley rats. Our results showed that the activities of liver and kidney markers were significantly ($P < 0.05$) increased while the activities of antioxidant enzymes were significantly decreased in treated rats with CCL₄. Oral administration of buffalo or cow colostrum for rats treated with CCL₄ resulted in a significant decrease in the activities of liver and kidney markers and a significant increase

in the activities of antioxidant enzymes were observed. No significant difference between rats administrated with cow or buffalo colostrum in their values of liver and kidney markers and antioxidant activities. Our results suggest that both types of colostrum could protect liver and kidney damage through enhancement the antioxidant capacities and improvement liver and kidney functions.

KEYWORDS: Buffalo colostrum, Natural products, Hepatoprotective, Biochemical markers, Antioxidant capacity.

1. INTRODUCTION

Hepatitis and kidney failure are the major public health problem worldwide, responsible for considerable morbidity and mortality from chronic liver and kidney diseases.^[1] Hepatitis C is one of several viruses in the hepatitis family causing liver inflammation and it cannot function properly and remove harmful material from the blood or convert food into energy. Worldwide, it is estimated that 130-170 million people are living with chronic hepatitis C infection, that it infects 3-4 million people per year, and that more than 350,000 people die

from hepatitis C related diseases each year. Countries with particularly high rates of infection include Egypt (22%), Pakistan (4.8%) and China (3.2%).^[2]

Carbon tetrachloride (CCL₄) has been used extensively one of chemical compounds in animal model system, with which it is studied liver hepatitis induced by free radicals in rats and mice. Liver damage caused by CCL₄ is regarded to inflammation in early stage as the analogue of hepatotoxins in humans. In the principle of liver damage, CCL₄ is reductively bioactivated by cytochrome P450 2E1 into a trichloromethyl radical ($\cdot\text{OCCl}_3$), which is subsequently converted into peroxy radical in the presence of oxygen. These reactive free radical metabolites can covalently bind to macromolecules and also initiate lipid peroxidation.^[3] Antioxidative action plays an important role in protection against CCL₄ induced liver injury. Protective effects of various natural products in CCL₄ hepatotoxicity have been reported.^[4]

Colostrum is an amazing material that like many other things in nature, reflects the evolutionary development of a unique composition that will serve the needs of the offspring for which it is intended. Colostrum is the first milk produced by mammals after birth as the first six postpartum (3-4 days) and is particularly rich in immunoglobulin, antimicrobial peptides (e.g., lactoferrin, and lactoperoxidase) and other bioactive molecules including growth factor, plus anti-inflammatory, antioxidant and immune enhancing components.^[5,6] Colostrum had higher concentrations of caseins and whey proteins than mature milk and there is evidence that caseins and whey proteins from colostrum exert antioxidant activities measured by reducing power, ferrous ion chelating abilities as well as inhibitory effects on lipid peroxidation.^[7] Different antioxidative enzymes in colostrum can prevent the formation of radicals or scavenge radicals or hydrogen peroxide and other peroxides. Among these enzymes, lactoperoxidase, catalase, superoxide dismutase and glutathione peroxidase. Non-enzymatic antioxidants in colostrum act as radical scavengers, such as lactoferrin, Vit. A and E, carotenoids and flavonoids.^[8]

There are not enough studies on the protective effect of buffalo colostrum against liver damage. Therefore, our study aimed to evaluate the hepatoprotective effect and antioxidative activities of cow and buffalo colostrum, the two main types of colostrum found in Egyptian markets, against CCL₄ induced liver damage in rat.

2. MATERIALS AND METHODS

2.1. Materials

Fresh cow and buffalo colostrum were collected after two hours of parturition from the herd of the Faculty of Agriculture, Cairo University, Giza, Egypt.

2.1.2. Chemicals

Gum acacia (5%), carbon tetra chloride (CCL₄), starch, cellulose, casein, vitamins and salt mixture were obtained from EL-Nasr pharmaceutical chemical Co., Cairo, Egypt. Corn oil was purchased from the local supermarket.

Kits for determination of aspartate and alanine aminotransferase (ALT and AST), alkaline phosphatase (ALP), serum gamma glutamyl transferase (GGT), superoxide dismutase (SOD), catalase (CAT), glutathion-S-transferase (GST), Glutathion peroxidase (GPx), urea, creatinine, total bilirubin and direct bilirubin were obtained from Biodiagonstic company, Giza, Egypt.

2.1.3. Animals

Male Sprague-Dawley (SD) rats weighting 180 ± 10 g were procured from animal house Lab., Agricultural Research Center, Giza, Egypt. This study was approved by the Committee of Scientific Ethics at Agricultural Research Center, Giza, Egypt and according to its guidelines.

2.2. Experimental protocol

2.2.1. Basel diet preparation

Basal diet was prepared according to Kawasaki et al. (2009).^[10] The chemical composition of the diet was as follows, casein (17%) starch (70%), soy bean oil (3%), cellulose (4.5%), salt mixture (4%) and vitamin mixture (1%).

2.2.2 Lyophilized colostrum preparation

Cow and buffalo colostrum samples were freeze – dried using freeze –drying (LY-5M-R133 inijders scientific, Amsterdam, Holland). Lyophilized cow and buffalo colostrum were suspended in 5 % gum acacia for rats oral administration.

2.2.3. Animal groups

All SD rats were fed basal diet and water for one week as adaption period and divided into four groups (G), each group contains eight rats.

G1: rats were fed basal diet.

G2: rats were injected with CCL₄ after 2 weeks of feeding basal diet.

G3: rats were injected with CCL₄ after 2 weeks of oral administration of cow colostrum and feeding basal diet.

G4: rats were injected with CCL₄ after 2 weeks of oral administration of buffalo colostrum and feeding basal diet.

The experiment period was 4 weeks. Daily colostrum dose for each rat was 0.1g/ 100g body weight as a recommended therapeutic dose of colostrum for human^[11] and^[12] in mice model.

2.2.4. Hepatitis induction

Rats were injected intraperitoneally by 50% CCL₄ (v/v) in liquid paraffin. Rats were treated with CCL₄ at dose of 0.5 mL/ kg body weight to induce hepatic injury.^[13]

2.2.5. Blood samples collection

After 4 weeks, animals were anesthetized with ether and blood samples were collected with or without anticoagulant (10 mM EDTA) from retro- orbital venus plexus. Blood samples of all treatments were centrifuged at 3000 rpm/20 min. separated plasma or serum was stored in Ependrop vials in freezer at -18 °C till the biochemical analysis.

2.3. Methods of Analysis

2.3.1. Chemical and microbiological analysis of colostrum

Cow and buffalo colostrum were examined chemically according to A.P.H.A (1992)^[14] and microbiologically according to Oxoid (1982).^[15] The obtained data are shown in Table (1).

2.3.2. Biochemical assay

The activities of AST, ALT, ALP, GGT, SOD, CAT, GST, GPx, urea, creatinine, total bilirubin and direct bilirubin were measured using the colorimetric methods described in the kits from Biodiagonstic, Giza, Egypt and according to the clinical guide to laboratory tests.^[16]

2.3.3 Statistical analysis

Results were analyzed statistically with SPSS 10.0 one – way ANOVA to assess the significance of differences between groups with $P < 0.05$ being considered significant. Data were expressed as mean \pm standard error (SE). The least significant difference (LSD) and one way analysis of variance were used for multiple comparisons between treatments. Each test was analyzed out in triplicate.^[17]

3. RESULTS AND DISCUSSION

3.1. Effect of oral administration of cow and buffalo colostrum on liver and kidney weight in CCL₄ treated rats.

The variations in the liver and kidney weights of rats subjected to difference treatments are shown in Figure (1). During the course of present investigations, it was observed that liver and kidney weights were significantly ($P < 0.05$) decreased in rats group injected with CCL₄ (G2) in comparison with rats in control group (G1). Also, the data obtained in the same Figure showed that oral administration of either cow (G3) or buffalo (G4) colostrum had significant protection of liver and kidney from weight loses induced by CCL₄ in comparison with G2. Furthermore, no significant differences in liver and kidney weight values could be observed between rats orally administrated with cow or buffalo colostrum. These results are in agreement with those reported by Ko et al (2006)^[13] who reported that CCL₄ resulted in a significant decrease in the body and liver weights of mice. On the other hand, the shrink effect of CCL₄ on liver and kidney weight was significantly decreased in rats of groups 3 and 4. Colostrum could enhance kidney function and regrowth kidney or liver tissues that ordinarily shrink with age or injury^[21] and protein synthesis and body weight in pigs.^[22]

3.2. Effect of oral administration of cow and buffalo colostrum on liver markers and plasma antioxidant enzymes activity.

The obtained data in Table (2) reveal that different liver markers including ALT, AST, ALP, GGT, total bilirubin and direct bilirubin are significantly increased in the CCL₄ treated rats (G2) in comparison with rats in G1, G3 and G4. Also, no significant difference could be found between rats administrated orally with cow (G3) or buffalo (G4) colostrum in their liver markers. Data in Table (3) show that oral administration of cow or buffalo colostrum can promote the plasma antioxidant enzymes in rats of G3 and G4 as compared with rats in G1 and G2.

Carbon tetrachloride (CCL₄) is used in production of toxic free radicals (CCL₃) which causes injury for different tissues including brain, liver, kidney and skin^[3] resulting in an increase in levels of liver markers^[19] and levels of direct bilirubin.^[20] Our results indicate degenerative changes and hypofunction of the liver and showed that the harmful and hepatotoxicity effect of CCL₄ that administrated in rats or mice.^[23,24] Also, the hepatoprotective effect of colostrum may be attributed to the colostrum contains high amounts of immunoglobulin A (IgA)^[9] and whey proteins^[35,36] that have protective effects against CCL₄ damage.

Antioxidant enzymes (SOD, CAT, GST, and GP_x) play an important role in liver protection against CCL₄ induced liver injury. SOD plays an important role in the elimination of reactive oxygen species derived from the peroxidative process in liver tissues.^[25] Moreover, SOD removes superoxide by converting it to hydrogen peroxide which can be rapidly converted to water by catalase.^[23] Treatment with CCL₄ (G2) in Table (3) shows a significant decrease ($P < 0.05$) in all plasma antioxidants parameters values that might be due to the principle causes of CCL₄ induced hepatic damage are lipid peroxidation, decrease of antioxidant enzymes activities and generation free radicals.^[26] On the other hand, treatments with administration cow or buffalo colostrum (G3 and G4, respectively) showed a significant ($P < 0.05$) increase in all plasma antioxidants parameters. The ability of colostrum to promote plasma antioxidant enzymes was due to that colostrum itself can be the source of reactive oxygen species (ROS) due to the presence of macromolecules susceptible to peroxidative damage as well as ROS generating systems.^[27] Also, human experiments showed that oxygen radical absorbing capacity was high in colostrum.^[28]

3.3. Effect of oral administration of cow and buffalo colostrum on serum kidney markers in CCL₄ treated rats.

Results in Figure (2) illustrate that CCL₄ injection caused a significant ($P < 0.05$) increase of serum creatinine and urea (renal failure markers) values in G2 in comparison with G1. On the other hand, oral administration of cow or buffalo colostrum (G3 and G4, respectively) could significantly reduce serum creatinine and urea values in rats treated with CCL₄ in comparison with rats in G2. Chronic kidney disease (CKD) is identified by a blood tests for creatinin and urea levels. High levels of creatinin and urea indicate a falling glomerular filtration rate and followed by a decrease in the capability of the kidneys to excrete waste products. Free radicals formed from CCL₄ metabolism induce lipid peroxidation and believed to be one of the major causes of cell membrane damage leading to a number of pathological situations by causing acute and chronic renal injuries in animal models^[29,30] and humans.^[31] Treatment with CCL₄ (G1) as can be observed in Figure (2) shows significant ($P < 0.05$) increase in urea and creatinin parameters as a result of kidney tissues damage.^[32] On the other hand, oral administration with either cow or buffalo colostrum could decrease levels of creatinin and urea in rats injected with CCL₄ (G3 and 4, respectively). Results reported by Davis et al (2007)^[33] illustrated that bovine colostrum didn't have any toxic effect on liver or kidney functions in rats.

Table 1. Chemical composition (%) and microbiological analysis (log cfu mL⁻¹) of colostrum samples.

Colostrum Samples	Chemical Composition %					Microbiological analysis Log cfu mL ⁻¹			
	Total Solids	Fat	Protein	Lactose	Ash	Total count	Mold & Yeast	Coliform	<i>Staph. aureus</i>
Cow	19.90± 0.50	5.15± 0.30	12.51±0.18	1.60±0.30	0.72±0.13	3.30±0.23	1.50±0.41	ND	ND
Buffalo	28.70± 0.25	10.00±0.42	14.00±0.15	2.62±0.26	1.88±0.16	3.40±0.35	1.30±0.52	ND	ND

±: Standard error

Table 2. Effect of oral administration of cow or buffalo colostrum on some liver functions in CCL₄ treated rats.

Treatments *	ALT**	AST**	ALP**	GGT**	Total Bilirubin	Direct Bilirubin
	UL ⁻¹				mg dL ⁻¹	
G1	186.00±0.70 ^b	355.77±0.60 ^b	128.73±0.60 ^b	5.06±0.50 ^b	16.12±0.60 ^b	12.13±0.70 ^b
G2	210.76±2.70 ^a	389.83±0.80 ^a	149.80±1.05 ^a	12.10±2.60 ^a	27.90±1.60 ^a	23.55±2.80 ^a
G3	186.50±1.50 ^b	362.50±1.30 ^b	139.56±1.70 ^b	6.20±1.05 ^b	20.32±2.02 ^b	17.53±0.90 ^b
G4	184.60±2.60 ^b	359.96±1.50 ^b	130.60±2.05 ^b	5.26±1.70 ^b	20.90±0.70 ^b	17.64±2.60 ^b

*: G1: rats were fed basal diet, G2: rats injected with CCL₄ after 2 weeks of feeding basal diet, G3: rats injected with CCL₄ after 2 weeks of oral administration of cow colostrum, G4: rats injected with CCL₄ after 2 weeks of oral administration of buffalo colostrum.

** : ALT: Aspartate aminotransferase, AST: Alanine aminotransferase, ALP: alkaline phosphatase, GGT: serum gamma glutamyl transferase.

±: Standard error.

Table 3: Effect of oral administration of cow or buffalo colostrum on antioxidant activity (UL⁻¹) in CCL₄ treated rats.

Treatments*	SOD**	CAT**	GST**	GP _x **	GSH**
G1	30.34±1.02 ^a	29.64±2.70 ^a	33.48±1.60 ^a	36.69±0.80 ^a	33.04±1.90 ^a
G2	17.32±2.70 ^b	13.76±1.80 ^b	15.22±2.80 ^b	12.19±2.90 ^b	19.46±2.70 ^b
G3	34.58±1.45 ^a	29.38±1.50 ^a	35.02±1.70 ^a	33.34±0.56 ^a	33.40±1.68 ^a
G4	39.94±1.90 ^a	36.46±2.60 ^a	37.78±0.90 ^a	34.63±1.00 ^a	43.30±1.80 ^a

*: G1: rats were fed basal diet, G2: rats injected with CCL₄ after 2 weeks of feeding basal diet, G3: rats injected with CCL₄ after 2 weeks of oral administration of cow colostrum, G4: rats injected with CCL₄ after 2 weeks of oral administration of buffalo colostrum.

** : SOD: Superoxide dismutase, CAT: Catalase, GST: glutathion-S-transferase, GP_x: Glutathion peroxidase and GSH: reduced glutathione.

±: Standard error

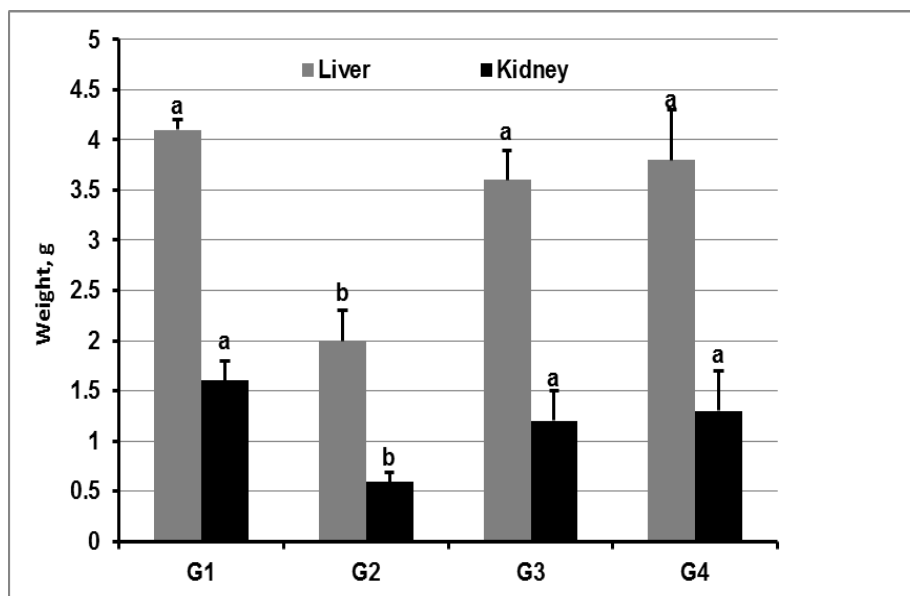


Figure 1: Effect of oral administration of cow or buffalo colostrum on liver and kidney weight (g) in treated CCL₄ rats.

G1: rats were fed basal diet, G2: rats injected with CCL₄ after 2 weeks of feeding basal diet, G3: rats injected with CCL₄ after 2 weeks of oral administration of cow colostrum, G4: rats injected with CCL₄ after 2 weeks of oral administration of buffalo colostrum.

Error bars represent means of three replicates.

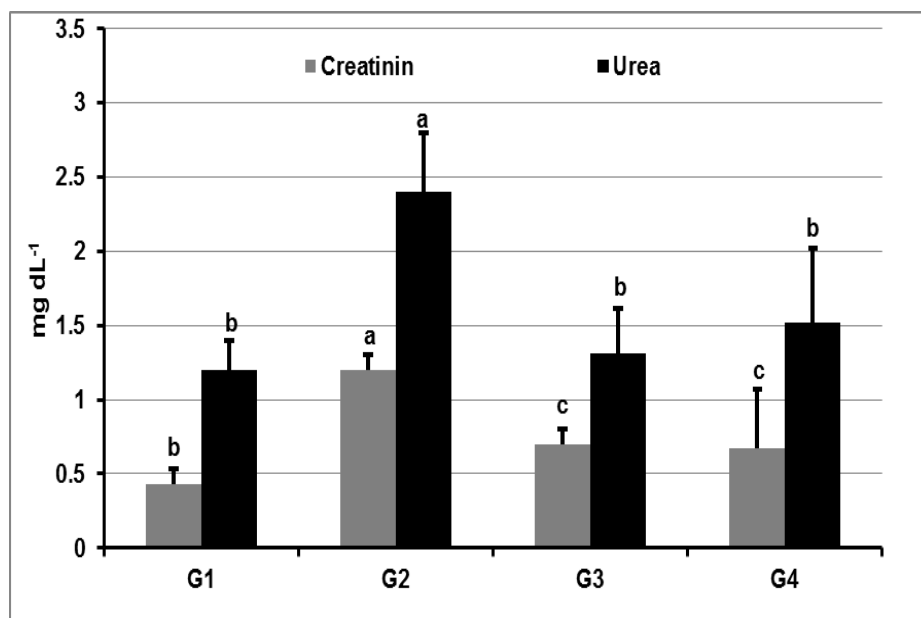


Figure 2: Effect of cow and buffalo colostrum administration on the concentration of serum creatinin and urea (mgdL⁻¹) in treated CCl₄ rats.

G1: rats were fed basal diet, G2: rats injected with CCl₄ after 2 weeks of feeding basal diet, G3: rats injected with CCl₄ after 2 weeks of oral administration of cow colostrum, G4: rats injected with CCl₄ after 2 weeks of oral administration of buffalo colostrum.

Error bars represent means of three replicates.

4. CONCLUSION

Colostrum is a natural product and it is rich source in bioactive components. Cow and buffalo colostrum are the major two types of colostrum in Egyptian markets. Our findings indicated that oral administration of either cow or buffalo colostrum could enhance the liver and kidney weights, liver and kidney functions and antioxidant activities after CCl₄ exposure. Therefore, both types of colostrum might be used as a dietary strategy for enhancing of liver functions and antioxidant capacities. Indeed, it would be interesting to investigate the molecular mechanisms by which both main types of colostrum in Egypt can protect liver and kidney from damage induced by CCl₄.

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6. REFERENCES

1. Lau DT and Membreno, FE. Antiviral therapy for treatment hepatitis B virus patients. *Gastroenterol. Clin North Am*, 2004; 33: 581-99.
2. World Health Organization (WHO), Hepatitis C factsheet, Retrieved, 2011; 7-13.
3. Goepter AR, Scheerens H and Vermeulen NP. Oxygen and xenobiotic reductase activities of cytochrome P450. *Crit Rev Toxicol*, 1995; 25: 25-65.
4. Hsiao G, Shen MY, Lin KH, Lan, MH, Wu, LY, Chou, DS, Lin, CH, Su, CH and Sheu, JR. Antioxidant and Hepatoprotective effects of antrodia camphorate extract. *J Agric Food Chem*, 2003; 51: 3302-3308.
5. Playford RJ, Macdonald CE and Johnson WS. Colostrum and milk derived peptide growth factors for the treatment of gastrointestinal disorders. *Am J Clin Nutr*, 2000; 72: 5-14.
6. Moller W, Reinhard L and Nitsche D. Use of bovine colostrum milk as a preparation for the protection of the liver. USA Patent; US 5710132: 1998.
7. Chiang SH and Chang CY. Antioxidant prosperities of caseins and whey proteins from colostrum. *J Food Drug Anal*, 2005; 13: 57-63.
8. Przybylska J, Albera E and Kankofer M. Antioxidants in bovine colostrum. *Reprod Dom Anim*, 2007; 42: 402-409.
9. Xiong H, Lib B, Wu J. Detection of hepatitis C virus markers in colostrum. *Zhonghua Fu Chan Ke Za Zhi*, 1997; 32: 138-140.
10. Kawasaki T, Igarashi K, Koeda T, Sugimoto K, Nakagawa K, Hayashi S, Yamaji R, Inui H, Fukusato T, Yamanouchi T. Rats fed fructose enriched diets have characteristics of nonalcoholic hepatic steatosis. *J Nutr*, 2009; 139: 2067-2071.
11. Antonio J, Sanders MS and Van Gammeren, D. The effects of bovine colostrum supplementation on body composition and exercise performance in active men and women. *Nutr*, 2001; 17: 243-247.
12. Yoshioka Y, Kudo S, Nishimura H, Yajima T, Kishihara K, Saito K, Suzuki T, Suzuki Y, Kuroiwa S and Yoshikai Y. Oral administration of bovine colostrum intestinal intraepithelial lymphocytes to polarize Th1-type in mice. *Int Immunopharmacol*, 2005; 5: 581-590.
13. Ko, JH, Lee, SJ and Lim, KT. Rhus verniciflua stockes glycoprotein (36 kDa) has protective activity on CCL₄ -induced liver injury in mice. *Environ Toxicol Pharmacol*, 2006; 22: 8-14.

14. A. P. H. A. Standard method of examination of dairy products. American public health association, 1992, 16th ed. Washington D.C. USA.
15. Oxoid Manual[®]. The oxide manual of culture media, ingredients and other laboratory services 1982, Oxide limited, Basingstoke, Hampshire, England.
16. Tietz NW (Ed): Clinical Guide to Laboratory Tests, 1995 3rd ed. W. B. Saunders, Philadelphia, PA.
17. SPSS for windows. Chicago: SPSS Inc, 2007. Release 16.0.0.
18. Suja SR, Latha PG, Pushpangadan P and Rajasekharan, P. Evaluation of hepatoprotective effects of *Helminthostachys Zeylanica* (L) Hook against carbon tetrachloride induce liver damage in wistar rats. *J Ethnopharmacol*, 2004; 92: 61-66.
19. Thomas L. Laboratory and diagnosis. Frankfurt/ Main, TH-Books, 2005.
20. Liu Y, Li P, Lu J, Xiong W, Oger J, Tetzlaff W and Cynader M. Bilirubin possesses powerful immunomodulatory activity and suppresses experimental autoimmune encephalomyelitis. *J Immunol*, 2008; 181: 1887-1897.
21. Batash, S, Weinschel, E, Falkenstein, D, Raicht, R, Ma, T, Katz, K and Hollander, D. Intestinal permeability in HIV infection: proper controls are necessary (letter). *Am J Gastroenterol*, 1992; 87: 680- 685.
22. Burrin DG, Davis TA, Ebner S, Schoknecht P, Fiorotto M and Reeds J. Colostrum enhances the nutritional stimulation of vital organ protein synthesis in neonatal pigs. *J Nutr*, 1997; 127: 1284-1289.
23. Abdel-Wahhab MA, Ahmed HH and Hagazi MM. Prevention of aflatoxin B₁-initiated hepatotoxicity in rat by marine algae extracts. *J Appl Toxicol*, 2006; 26: 229–238.
24. Bhattacharjee R and Sil PC. Protein isolate from the herb, *Phyllanthus niruri* L. (Euphorbiaceae), plays hepatoprotective role against carbon tetrachloride induced liver damage via its antioxidant properties. *Food Chem Toxicol*, 2007; 45: 817–826.
25. Packer JE, Slater TF and Willson RL. Reactions of the carbon tetrachloride-related peroxy free radical with amino acids: pulse radiolysis evidence. *Life Sci*, 1978; 23: 2617–20.
26. Poli G. Liver damage due to free radicals. *Br. Med. Bull*, 1993; 49: 604-20.
27. Shoji H, Oguchi S, Shimizu T, Yamashiro Y. Effect of human breast milk on urinary 8-hydroxyl-2 deoxyguanosine excretion in infants. *Pediatric Res*, 2003; 53, 850: 52.
28. Alberti-Fidanza A, Burini G and Perriello G. Total antioxidant capacity of colostrum and transitional and mature human milk. *J Matern Fetal Neonat Med*, 2002; 11: 275-79.

29. Ogeturk, M., Kus, I., Colakoglu, N., Zararsiz, I., Ilhan, N. and Sarsilmaz, M. Caffeic acid phenyl ester protects kidney against carbon tetrachloride toxicity in rats. *J of Ethnopharma*, 2005; 97: 273–80.
30. Adewole, SO, Salako, AA, Doherty, OW and Naicker, T. Effect of melatonin on carbon tetrachloride-induced kidney injury in Wistar rats. *Afr J of Biomed Res*, 2007; 10: 153–164.
31. Manna P, Mahua S and Parames CS. Aqueous extract of *Terminalia arjuna* prevents carbon tetrachloride induced hepatic and renal disorders. *BMC Complementary and Alternative Medicine*, 2006; 6: 33-35.
32. Waterfield CJ, Turton JA, Scales MD and Timbrell JA. Investigation into the effects of various hepatotoxic compounds on urinary and liver taurine levels in rats. *Arch Toxicol*, 1993; 67: 244-254.
33. Davis PF, Creenhill NS, Rowan AM and Schollum LM. The safety of new Zealand bovine colostrum: nutritional and physiological evaluation in rats. *Food and Chem. Toxicol*, 2007; 45: 229-236.
34. Sanmugapnya E, Venkataraman S. Studies on hepatoprotective and antioxidant actions of *strychnos potatorum* linn seeds on CCL₄ – induced acute hepatic injury in experimental rats. *J. Ethnopharmacol*, 2006; 105: 154-160.
35. Kume H, Okazaki K, Sasaki H. Hepatoprotective effects of whey proteins on D-galactosamine-induced hepatitis and liver fibrosis in rats. *Biosci. Biotechnol. Biochem*, 2006; 70: 1281-1285.
36. Gad AS, Khadrawy YA, El-Nekeety AA, Mohamed SR, Hassan NS, Abdel-Wahhab, MA. Antioxidant activity and hepatoprotective effects of whey protein and *Spirulina* in rats. *Nutr*, 2011; 5: 589-599.