



## A REVIEW ON ANTIMICROBIAL ACTIVITIES OF TRIPHALA AND ITS CONSTITUENTS

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Article Received on  
22 Jan 2016,

Revised on 13 Feb 2016,  
Accepted on 04 Mar 2016

DOI: 10.20959/wjpps20164-6341

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### ABSTRACT

An infection is the detrimental colonization of a host organism by a foreign replicator. Infectious diseases remain the first cause of premature death throughout the world. Search of new natural compounds with pharmacological properties is a field of interest widely growing, especially for the management of infectious diseases such as meningitis, tuberculosis, malaria, hepatitis, AIDS etc. Dietary intake of foods or plant based extracts with antioxidant properties were shown to have beneficial effects on human health and improve immune functions against these diseases. Triphala is an Ayurvedic herbal formulation which contains equal proportion of fruits of *Terminalia bellerica* Roxb. (Bibhitaki), *Terminalia chebula* Retz. (Haritaki) and *Embllica officinalis* Gaertn. (Amalaki). It is a well known phytomedicine with known antioxidant, antibacterial, antifungal,

antiviral, antiparasitic and anti-inflammatory activities. It has been used in the traditional medicines either alone or in combination with other plants for treatment of various health ailments. The most traditional usage of Triphala is to improve digestion, correct constipation, tone gastrointestinal tract and reduce oxidative stress. This review summarizes the current knowledge about the antimicrobial effects of Triphala and its constituents in counteracting oxidative stress as well as inflammatory mechanisms, using in vitro and in vivo models of acute and chronic infectious diseases.

**KEYWORDS:** Triphala, *T. bellerica*, *T. chebula*, *E. officinalis*, Infectious disease, Antimicrobial activity.

## 1. INTRODUCTION

Infectious diseases are the leading cause of death worldwide, and the numbers of deaths from infectious diseases are increasing day by day. Of all infectious disorders pneumonia, diarrhea, tuberculosis and malaria have been the leading causes of death.<sup>[1]</sup> According to recent literature 50,000 men, women and children are dying every day due to these diseases.<sup>[2]</sup> This translates into approximately 50,000 preventable deaths per day.<sup>[1,3]</sup> If present trends continue, 4.4 million people will still die in 2030.<sup>[1]</sup> Microbes that cause illness are also known as pathogens. The most common pathogens are bacteria and viruses, though a number of other microorganisms, including some kinds of fungi and protozoa, also cause disease (Table 1). An infectious disease is termed communicable if it is easily transmitted from one person to another. There is also strong evidence that microbes may contribute to many non-infectious chronic diseases such as some forms of cancer and coronary heart disease.<sup>[4]</sup> In the human host, a microorganism causes disease by either disrupting a vital body process or stimulating the immune system to mount a defensive reaction. An immune response against a pathogen, which can include high fever, inflammation and other damaging symptoms, may be more destructive than the direct damage caused by the microorganism.<sup>[5]</sup>

Antibiotics are a type of antimicrobials used to treat various infections and are used specially against bacteria. Several such agents are also effective against a number of fungi, protozoa, some are toxic to human and animals even when given in therapeutic dosage. Antibiotics are not effective against viruses and may be harmful when taken inappropriately. Promoting appropriate use of natural or synthetic antibiotics and preventing the spread of drug resistant bacteria are key issues in tackling the public health problem of antimicrobial resistance. Resistant microorganisms are able to survive attack by antimicrobial drugs, so that standard treatments become ineffective and infections persist, increasing the risk of spread of these microorganism to others hosts. Multidrug resistance (MDR) creates serious challenges to the medicinal field and infections caused by MDR bacteria.<sup>[6]</sup> Target based drug resistance to the antibiotics have also been reported. Recent studies also indicate that synthetic drugs are emerging as drugs of abuse for college students and young military personnel.<sup>[7]</sup> Nowadays, people suffering from the side effects of antimicrobial resistance are trying to find alternative solution in natural products. Medicinal plants provide a wealth of antimicrobial agents. Herbs are used to treat various infectious diseases worldwide. Interestingly, some herbs have antimicrobial activity against bacterial pathogens in addition to their flavoring effects.<sup>[8]</sup> From the earliest times, herbal spices are added for improving taste which naturally and safely

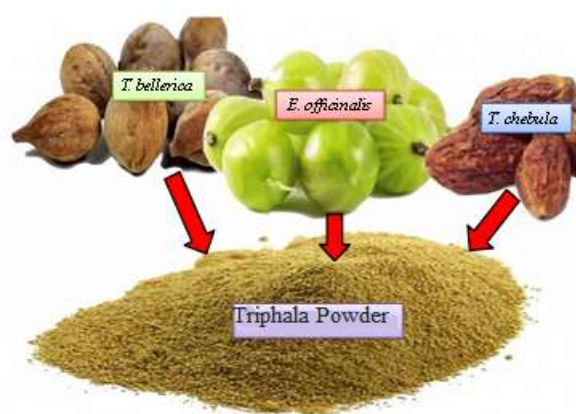
renew shelf life of food products. Bacterial pathogens are sensitive to extracts from many traditional plants.<sup>[9]</sup> Therefore, the natural plants have been used as antimicrobial agents which provide a promising safe solution. Natural products have certain advantages besides being cheap to produce; they are biodegradable and readily available. Herbal medicines are also used as dietary supplements and are produced without added artificial ingredients.<sup>[10]</sup>

**Table:1. Microbial infectious diseases and their annual case numbers**

Type of micro organism	Name of Microorganism	Disease name	Effect of disease	Annually reported cases
<b>Bacteria</b>	Haemophilus influenza	meningitis	brain damage, hearing loss or learning disabilities	4,100
	Shigella dysenteriae	Shigellosis	dysentery (diarrhea with blood), fever, abdominal pain, rectal tenesmus	300,000
	Clostridium perfringens	gas gangrene	skin injury, sudden pain, feeling of heaviness, a low-grade fever, apathetic mental status, necrosis	3000
	Bacillus anthracis	anthrax	fever, chills, nonproductive cough, chest pain, headache, myalgias, and malaise	20,000 to 100,000
	Mycobacterium leprae	Leprosy (a rare disease)	anesthesia, weakness, paralysis, muscular atrophy, skin lesions, affects mucous membrane & peripheral nerves	108
	Borrelia burgdorferi	syphilis	arthritis, brain damage, and blindness.	100,000
	Borrelia burgdorferi	lyme	irregular heartbeat, eye & liver inflammation (hepatitis), apoptosis, severe fatigue.	30,000
	Vibrio cholera	cholera	watery diarrhoea, vomiting and leg cramps, can quickly result in dehydration and death	> 500 000
	Clostridium tetani	tetanus	impairs in motor neurons, muscle stiffness and spasms	49,000
	Mycobacterium tuberculosis	tuberculosis	cough with sputum and blood at times, chest pains, weakness, weight loss, fever and night sweats	9.6 million
	Bordetella pertussis	pertussis	runny nose, sneezing, mild cough, low-grade fever.	32, 971
	Solmonella typhi	typhoid fever	poor appetite, abdominal pain, headaches, fever, intestinal bleeding.	724.6
	Yersinia pestis	plague	enlarged and painful lymph nodes, fever, chills, headaches, and weakness.	1,000 to 3,000
	Streptococcus pyogenes	scarlet fever	sore throat, fever, and a characteristic red rash	1000
Rickettsia parkeri	rickettsioses	fever, headache and skin rashes	579	
<b>Fungi</b>	Aspergillus niger	aspergillosis	fever, bloody cough, chest or joint pain, headaches or eye symptoms, nose bleed,	3.5 million

			facial swelling, skin lesions	
	Blastomyces dermatitidis	blastomycosis	fever, chills, cough, and discomfort or pain in the muscles, joints and chest pain	1.5 million
	Candida albicans	candidiasis	veginal itching, swelling and irritation.	7,000 to 28,000
	Histoplasma capsulatum	Histoplasmosis	Ch chills, cough, chest and joint pain, muscles s stiffness, shortness in breathing	144,462
	Pneumocystis firovecii	pneumocystis pneumonia	fever, mild and dry cough, rapid breathing, fatigue, major weight loss, chest pain during breathing	4.9 milliom
	Sporothrix schenckii	sporotrichosis	nodule formation, shortness of breath, cough, and fever.	1000
	Trichophyton rubrum	Ringworm	itchy, red, raised, scaly patches around the outside normal skin	>10,000
<b>Protozoa</b>	Entamoeba histolytica	amoebiasis	loose stools, stomach pain and stomach cramping	100,000
	Giardia lamblia	giardiasis	diarrhea, gas, greasy stool, stomach or abdominal cramps, nausea	19,888
	Trypanosoma brucei	African sleeping sickness	fever, severe headaches, irritability, fatigue, swollen lymph nodes, aching muscles and joints.	300 000
	Leishmania donovani	leishmaniasis	ulcers of skin, mouth, fever, low red blood cells, enlarged spleen and liver	20 000 to 30 000
	Toxoplasma gondii	Toxoplasmosis	fever, muscle aches, fatigue, swollen lymph glands.	20,000
	Plasmodium falciparum	malaria	high fever, profuse sweating, headache, vomiting, diarrhea, anemia	438 000
	Babesia microti	babesiosis	fever, chills, sweats, headache, body aches, loss of appetite, nausea, fatigue.	99, 226
	Trichomonas vaginalis	Trichomonias (STD)	vaginal itching and discharge, painful urination, swelling in the groin	100,000
<b>Virus</b>	Human papilloma virus (HPV)	HPV infections	formation of warts including genital, common, plantar, flat, dark color of skin, chances of cervical cancer	12,900
	Varicella Zoster virus.	Chicken pox	fever, rash of itchy inflamed pimples which turn to blisters and then loose scabs	100,000
	Human Immunodeficiency virus	AIDS	fever, chills, joint pain, muscle ache, sore throat, sweats enlarged glands, a red rash, tiredness, weakness, weight loss.	1.4 – 1.7 million
	Herpes	Hepatitis C	fatigue, abdominal pain, nausea, loss of appetite, or yellow jaundice. Jaundice	>350 000
	Chlamydia trachomatis	Chlamydia	lack of hunger, vomiting, belly aches, itchiness, feverish, muscle and joint pain.	> 20 million
	Influenza	Flu	fever, cough. sore throat. runny nose, muscle or body aches, headaches, fatigue vomiting and diarrhea.	250,000 to 500,000
	Herpes virus	Viral meningitis	headache, dislike of bright lights, neck stiffness, fever and nausea/vomiting.	36,000

Triphala is an Indian tridoshic herbal formulation consisting of fruits of *T. bellerica* (TB), *T. chebula* (TC) and *E. officinalis* (EO) in 1:1:1 ratio (Fig. 1). Recipe for this traditional herbal supplement is described in the traditional Indian texts, the Charaka and Susruta Samhita. According to Ayurveda the resultant formulation was shown to promote health, immunity and longevity when used in a recommended manner.<sup>[11]</sup> Triphala corrects constipation, act as restoratives for gastrointestinal tract and also detoxifies the whole body and improves digestion and assimilation.<sup>[12]</sup> Triphala and its constituent's acts as cardiogenic, control blood pressure, improve blood circulation and reduce cholesterol level and helps in improving body's defense system.<sup>[14]</sup> Triphala and its constituents have shown antioxidant, anti-inflammatory, hepatoprotective<sup>[15,16]</sup> etc. that make them one of the most commonly used traditional medicines. Triphala is known to give a long life and is invigorating to whosoever takes it, as it works slowly and gently and may be taken over long periods of time without any side effect. Triphala and its constituents also showed strong antimicrobial activity against different microorganism (Table 2,3,4). Many active antimicrobial compounds were isolated from it by FT-IR and GC-MS analysis.<sup>[14]</sup> Analysis of fruits of TB, TC and EO has shown significant microbial inhibition.<sup>[17]</sup> This review aims to give an overview of the recent scientifically tested antimicrobial activities of Triphala and its constituents which may be used as a good preventive/remedy against infectious diseases.



**Figure 1. Triphala and its constituents**

## **2. Ethanomediccal use of Triphala and its constituents**

The combined use of different plant extracts is useful in decreasing drug resistant problems.<sup>[18]</sup> Triphala contains various phenolic and nonphenolic compounds<sup>[19]</sup> effective against both pathogenic and non pathogenic bacterial strains.<sup>[20]</sup> Triphala was found to be effective against enteric bacterial pathogens and enterococci.<sup>[21]</sup> It has shown broad

spectrum antimicrobial activity against some resistant bacterial isolates. Triphala and its constituents are also reported effective against human pathogenic bacteria.<sup>[22]</sup> Aqueous and ethanolic extract of Triphala and its ingredients have shown promising effects against the growth of bacterial strains isolated from the HIV infected patients.<sup>[23]</sup> Use of Triphala as mouthwash effectively reduced the number of mutant streptococci in saliva.<sup>[24]</sup> The bacterial isolates isolated from the wounds also showed sensitivity towards the extracts of Triphala when tested *in vitro*.<sup>[25]</sup> Additionally, Triphala and its constituents are reported to possess antifungal and antiprotozoal activity (Table 3,4). Ingredients of Triphala showed significant inhibitory activity at lowest IC<sub>50</sub> values against human immunodeficiency virus-1 reverse transcriptase (Table 4)

### 2.1. *T. bellerica* (TB)

TB is a large deciduous tree which belongs to family Combretaceae. It is generally known as felleric mycobalane in English and locally as Baheda. It is laxative, astringent, analgesic, antipyretic and antiemetic in nature whereas seeds are rich in oil and have narcotic properties.<sup>[26,27]</sup> In Ayurveda fruits and its kernel are said to possess medicinal value. Fruits of TB contain different tannins, flavonoids and other phenolic compounds<sup>[27,28]</sup> that may be responsible for various biological activities. As per literature Ayurvedic plant parts are good in cold, cough, chronic diarrhea, dysentery and helps to increase appetite.<sup>[29]</sup> TB leaf showed good *in vitro* antioxidant activity against different reactive oxygen species.<sup>[15]</sup> The fruits are regarded as an excellent expectorant and strong rejuvenator against microbial infections.<sup>[30]</sup> Antimicrobial activity of TB fruit against a wide variety of pathogenic bacteria, yeast and fungi has been reported. TB fruit showed significant efficacy against virulence factors of respiratory and mammary gland infectious disease pathogens.<sup>[31]</sup> TB possesses antifungal compounds, that were shown promising challenge against fetal fungal diseases caused by cryptococcal pathogen.<sup>[32]</sup> Besides antibacterial and antifungal TB also has good activity against protozoa and virus (Table 4). TB showed potential to inhibit Hepatitis B surface antigen binding ability and HBV-DNA enzyme. It has also demonstrated trypanocidal activity against the related organism *Trypanosoma evansi* (Table 4)

### 2.2. *T. chebula* (TC)

It is top listed Ayurvedic medicinal plant belonging to family Comretaceae. TC is very rich source of tannins, phenolics, fatty acids and triterpenoid.<sup>[33,34]</sup> The leaves were found to contain polyphenols such as punicalin, punicalagin, terflavins B, C, and D. The plant is also

found to contain phloroglucimol and pyrogallol, along with phenolic acids such as ferulic, p-coumaric, caffeic and vanillic acids. The powder of the dried fruits of TC is used for the various therapeutic purposes to promote longevity. Due to strong antioxidant and in vivo wound healing capacity and significant medicinal properties,<sup>[15,16]</sup> TC is also known as the “King of Medicine” in Tibet.<sup>[35]</sup> TC exhibited antibacterial activity against a number of both Gram-positive and Gram-negative human pathogenic bacteria (Table 2). Ellagic acid isolated from TC showed good inhibiting potential against coliforms forming infectious pathogens.<sup>[36]</sup> The ethanolic extract of TC fruits demonstrated a strong antimicrobial activity against multidrug-resistant uropathogenic bacteria and phenolics were found to be responsible for this antibacterial activity. In vitro assessment of TC fruit, bark and leaves showed significant antifungal and antiviral activities (Table 3,4). Seed and fruit pericarp of TC have a potential against multi drug resistance parasite plasmodium falciparum to combat against malaria (Table 4).

### **2.3. E. officinalis (EO)**

It is a large tree belonging to Euphorbeaceae family, and is commonly known as Indian gooseberry in English and locally as Amla. It is well known that all parts of Amla plant are used to treat a range of diseases but most significant is the fruit. Fruit is used either alone or in combination with other plants to treat many common ailments such as cold, fever, peptic ulcer, dyspepsia and as digestive aid as well as serious diseases like cancer and cardiovascular disease.<sup>[37]</sup> The fruit of EO is also known for significant antimicrobial phenolic compounds.<sup>[37]</sup> It contains many active phytochemicals including flavonoids. It is one of the richest sources of vitamin C that were responsible for antioxidant, anti-inflammatory and antimicrobial activities. Gallic acid and tannic acid are the major phytoconstituents of EO and has strong antimicrobial potential.<sup>[38]</sup> In vitro assessment of EO fruit and leaf showed 100 percent antibacterial, antiprotozoal and antifungal activities. It has good antioxidant and anticancer activity and showed strong squabble against infectious diseases.<sup>[15,16]</sup> In more than 150 literatures cited in last 5 years little in vitro and in vivo studies of Triphala and its ingredients against infectious diseases caused by virus and protozoa were found. Thus Triphala and its constituents is emerging herbal warrior against infectious microorganisms.

### 3. Phytochemical of Triphala and constituents

Phytochemical analysis of Triphala and its constituent plants revealed the presence of a variety of antioxidant compounds such as phenolics, flavonoids, tannins, alkaloids, terpenoids, vitamins, glycosides, fatty acids and phytosterols.<sup>[27,39]</sup> Epidemiological studies have shown that many of these antioxidant compounds are responsible for antimicrobial activities to a greater or lesser extent. Antioxidant activity of phenolic compounds has been correlated to their chemical structures. Structure activity relationships of some phenolic compounds were thoroughly studied.<sup>[40]</sup>

#### 3.1. Flavonoids

Flavonoids are ubiquitous in photosynthesising cells and are commonly found in fruit, vegetables, nuts, seeds, stems, flowers etc. Plant samples containing high concentrations of flavonoids are frequently reported to show better antibacterial activity.<sup>[41]</sup> For centuries, preparations containing these compounds as the principal physiologically active constituents have been used to treat human diseases. Increasingly, this class of natural products is becoming the subject of anti-infective research and many research groups have isolated and identified the structures of flavonoids possessing antibacterial, antifungal and antiviral activities.<sup>[42]</sup> Owing to the widespread ability of flavonoids to inhibit spore germination of plant pathogens, they have been proposed for use against human fungal pathogens.<sup>[43]</sup> FT-IR analysis of extract of TB revealed the presence of alkaloids, phenol, tannins and flavonoids.

The flavonoid, 7-hydroxy-3,4-(methylenedioxy) flavan, isolated from TB fruit, has also been shown to possess activity against *C. albicans*. Inhibition of HIV-1 entry into cells expressing CD4<sup>+</sup> and chemokine co-receptors<sup>[44]</sup> and antagonism of HIV-1 reverse transcriptase by the flavone O-glycoside have been demonstrated by Li and colleagues<sup>[45]</sup>. Phytochemical analysis of TC showed promising activity of flavonoid against Gram positive bacteria and also showed good antifungal activity. Rutin and quercetin were isolated through HPTLC method from TC exhibit anti-inflammatory, antihepatotoxic, antiulcer and antimicrobial activities.<sup>[46]</sup>

#### 3.2. Tannins

Tannins were identified as another large class of phenolics present in Triphala and its constituents. These are generally subdivided into hydrolyzable and condensed tannins. Hydrolyzable tannins contain a central core of polyhydric alcohol such as glucose and hydroxyl groups, which are esterified either partially or wholly by gallotannins or



ellagitannins. In TC 33% of total phytoconstituents are hydrolysable tannins that are responsible for pharmacological activities.<sup>[47]</sup> These tannins contain phenolic carboxylic acids such as gallic acid, ellagic acid, chebulic acid and gallotannins. Ellagitannins such as punacalagin, casurarinin, corilagin, terchebulin, chebulanin, neochebulinic acid, chebulagic acid and chebulinic acid have been reported in literature as antimicrobials.<sup>[35]</sup> Tannins isolated from TB, TC and EO<sup>[48,49]</sup> have shown many biological and pharmacological activities.

### 8.3. Terpenoids

Terpenoids are naturally occurring organic chemicals that are under investigation for antibacterial, antiviral and other pharmaceutical functions. Terpenoids were found to exhibit antimicrobial activity<sup>[50]</sup>. GC-MS analysis of bark of EO showed very promising terpenoid compound such as lupeol and betulin.<sup>[51]</sup>

### 8.4. Phenolics

Phenolics are either direct or indirect antioxidants. They exhibit beneficial regulatory effects on signalling pathways. Antimicrobial action of phenolic acids against pathogens is hyperacidification of the plasma membrane.<sup>[52]</sup> This hyperacidification would alter cell membrane potential, making it more permeable, as well as affecting the sodium potassium ATPase pump implicated in ATP synthesis.<sup>[53]</sup> Antimicrobial phenolic compounds were isolated from TC, TB and EO through reverse phase chromatography, HPLC and confirmed by NMR and ESI-MS.<sup>[54]</sup>

### 8.5. Glycosides

Many bioactive glycosides have been reported such as alkyl, amino, cardiac or steroidal, cynogenic, terpenoidal etc. The alkyl glycoside agents have a particular utility in teeth cleaning preparations due to their improved activity against gram-positive bacteria.<sup>[55]</sup> Aminoglycoside antibiotics display concentration-dependent bactericidal activity against gram negative aerobes, some anaerobic bacilli and drug resistant staphylococci.<sup>[56]</sup> They require only short contact time and are most effective against susceptible bacterial populations that are rapidly multiplying. The inhibitory action of cardiac glycosides on active Na<sup>+</sup> and K<sup>+</sup> transporter has been reported.<sup>[57]</sup> Glycosides were isolated from TC, TB and EO revealed presence of antibacterial, antioxidant and other pharmacological activities.<sup>[58,59,60]</sup>

Table 2. Antibacterial activities of Triphala, *T. bellerica*, *T. chebula* and *E. officinalis*

Bacterial Name	Plant Name (strain name) <sup>[reference]</sup>			
	Triphala	<i>T. bellerica</i>	<i>T. chebula</i>	<i>E. officinalis</i>
<i>Bacillus subtilis</i>	NCIM 2718 <sup>[61]</sup>	ATCC6059 <sup>[62]</sup> ATCC 6633 <sup>[63]</sup>	MTCC 441 <sup>[35]</sup> MTCC 1790 <sup>[64]</sup> MTCC 121 <sup>[65]</sup>	MTCC 2274 <sup>[66]</sup>
<i>Escherichia coli</i>	ATCC 25922 <sup>[61]</sup> EC1211 <sup>[67]</sup> ATCC 8739 <sup>[67]</sup>	Enteropathogen <sup>[68]</sup> uropathogen <sup>[68]</sup> ATCC 25922 <sup>[62]</sup> NCIM 2931 <sup>[63]</sup>	ATCC 25922 <sup>[69]</sup> MTCC 1687 <sup>[70]</sup> ATCC 8739 <sup>[71]</sup> HM626200 <sup>[72]</sup> MTCC 2124 <sup>[73]</sup> MTCC 7410 <sup>[73]</sup> MTCC 46 <sup>[74]</sup> MTCC 452 <sup>[75]</sup> MTCC 448 <sup>[76]</sup> K-12 <sup>[77]</sup>	MTCC 730 <sup>[66]</sup> ATCC 25922 <sup>[62]</sup> ATCC 9637 <sup>[66]</sup> MTCC 723 <sup>[78]</sup> MTCC 443 <sup>[79]</sup> ATCC 632 <sup>[80]</sup>
<i>Klebsiella pneumonia</i>	ATCC 70063 <sup>[61]</sup> KP1221 <sup>[67]</sup> MTCC 4030 <sup>[14]</sup>	NCIM 2719 <sup>[63]</sup> MTCC 4030 <sup>[81]</sup>	ATCC 14380 <sup>[35]</sup> MTCC 6644 <sup>[70]</sup> ATCC 12657 <sup>[71]</sup> MTCC 3384 <sup>[73]</sup> MTCC 7407 <sup>[65]</sup> ATCC 70060 <sup>[69]</sup> MTCC 4030 <sup>[74]</sup>	ATCC 43816 <sup>[82]</sup> ATCC 15380 <sup>[62]</sup> MTCC 4030 <sup>[66]</sup> MTCC 2405 <sup>[78]</sup> MTCC 106 <sup>[79]</sup> ATCC 31488 <sup>[80]</sup>
<i>Pseudomonas aeruginosa</i>	ATCC 27853 <sup>[83]</sup> PA1231 <sup>[67]</sup> MTCC 1934 <sup>[14]</sup>	ATCC25619 <sup>[68]</sup> ATCC 27853 <sup>[63,68]</sup> MTCC 1934 <sup>[81]</sup>	ATCC 43495 <sup>[77]</sup> ATCC 27853 <sup>[35]</sup> ATCC 9027 <sup>[71]</sup> HM626201 <sup>[14]</sup> MTCC 1934 <sup>[74]</sup> MTCC 7093 <sup>[76]</sup> MTCC 2295 <sup>[73]</sup> MTCC 424 <sup>[75]</sup>	ATCC27853 <sup>[79]</sup> MTCC 1934 <sup>[80]</sup>
<i>Staphylococcus aureus</i>	ATCC 25923 <sup>[61]</sup> ATCC 6538P <sup>[67]</sup> ATCC 29213 <sup>[83,84]</sup> MTCC 3160 <sup>[14]</sup>	ATCC 9144 <sup>[68]</sup> ATCC 6538 <sup>[62]</sup> MTCC 3160 <sup>[81]</sup>	ATCC 25923 <sup>[69]</sup> MTCC 737 <sup>[70]</sup> ATCC 19615 <sup>[70]</sup> HM626197 <sup>[72]</sup> MTCC 7443 <sup>[72]</sup> MTCC 3160 <sup>[65]</sup> MTCC 740 <sup>[85]</sup> NCTC 6571 <sup>[69]</sup> MTCC447 <sup>[86]</sup> MTCC 87 <sup>[75]</sup>	ATCC 25923 <sup>[85]</sup> MTCC No.96 <sup>[78]</sup> ATCC 12600 <sup>[80]</sup> MTCC 3160 <sup>[81]</sup>
<i>Salmonella typhimurium</i>	NR	NCTC8393 <sup>[64]</sup> ATCC 13311 <sup>[68]</sup> ATCC 23564 <sup>[63,68]</sup>	SSFP 4S <sup>[35]</sup> MTCC 733 <sup>[65,66]</sup> MTCC3216 <sup>[77]</sup> MTCC 98 <sup>[73,75]</sup>	NR
<i>Streptococcus mutans</i>	MTCC 890 <sup>[88]</sup> DMST18777 <sup>[89]</sup>	MTCC 890 <sup>[90]</sup>	MTCC 497 <sup>[85]</sup>	MTCC 890 <sup>[91]</sup> ATCC 25175 <sup>[92]</sup>
<i>Proteus mirabilis</i>	NR <sup>[62]</sup>	NCIM 2241 <sup>[63]</sup>	MTCC 3310 <sup>[74]</sup>	NCIM2241 <sup>[62]</sup>

			MTCC 425 <sup>[73]</sup> HM626199 <sup>[72]</sup>	
<i>Proteus vulgaris</i>	NR	NR	MTCC 742 <sup>[65]</sup> MTCC 1771 <sup>[77]</sup> MTCC 744 <sup>[73]</sup> MTCC 426 <sup>[75]</sup>	ATCC 12454 <sup>[62]</sup> MTCC 0426 <sup>[66]</sup>
<i>Staphylococcus epidermidis</i>	NR	ATCC 12228 <sup>[63]</sup>	MTCC 3615 <sup>[35]</sup> ATCC 12228 <sup>[71]</sup> MTCC435 <sup>[75]</sup>	NR
<i>Bacillus cereus</i>	NR	ATCC 14579 <sup>[62]</sup> ATCC11778 <sup>[63]</sup>	NR	NR
<i>Lactobacillus Acidophilus</i>	MTCC 447 <sup>[93]</sup>	NR	ATCC 9361 <sup>[85]</sup>	MTCC 10307 <sup>[91]</sup>
<i>Streptococcus pyogenes</i>	ATCC 12204 <sup>[83]</sup>	NR	NR	MTCC1925 <sup>[94]</sup>
<i>Enterococcus faecalis</i>	EF1201 <sup>[67]</sup>	NR	NR	MTCC 2729 <sup>[94]</sup> MTCC 439 <sup>[66,91]</sup> ATCC 35550 <sup>[92]</sup>
<i>Corynebacterium rubrum</i>	NR	ATCC 14898 <sup>[63]</sup>	NR	NR
<i>Listeria monocytogens</i>	NR	ATCC 19112 <sup>[63]</sup>	NR	NR
<i>Shigella dysenteriae</i>	NR	ATCC 9361 <sup>[62]</sup>	NR	NR
<i>Salmonella paratyphi</i>	NR	ATCC 9150 <sup>[63]</sup>	NR	NR
<i>Streptococcus pneumoniae</i>	NR	UTI isolate <sup>[68]</sup>	NR	NR
<i>Yersinia enterocolitica</i>	NR	ATCC9610 <sup>[68]</sup>	NR	NR
<i>Acinetobacter sp.</i>	NR	NR	HM626198 <sup>[72]</sup>	NR
<i>AgrobacteriumT umefaciens</i>	NR	NR	MTCC 431 <sup>[74]</sup>	NR
<i>Bacillus amyloliquifaciens</i>	NR	NR	MTCC 1488 <sup>[95]</sup>	NR
<i>Brevundimonas diminuta</i>	NR	NR	ATCC 19146 <sup>[77]</sup>	NR
<i>Enterobacter aerogenes</i>	NR	NR	MTCC 7325 <sup>[65]</sup> MTCC 2822 <sup>[74]</sup> MTCC 111 <sup>[75]</sup> K-12 <sup>[77]</sup>	NR
<i>Helicobacter pylori</i>	NR	NR	NCTC RSB6 <sup>[77]</sup> NCTC 33098 <sup>[77]</sup>	NR
<i>Salmonella enteric</i>	NR	NR	NR	MTCC735 <sup>[93]</sup> ATCC23564 <sup>[87]</sup> ATCC 13311 <sup>[80]</sup>
<i>Staphylococcus saprophyticus</i>	NR	NR	NR	ATCC 35552 <sup>[61]</sup>
<i>Streptococcus</i>	NR	NR	NR	MTCC 0459 <sup>[66]</sup>

faecalis				ATCC 8043 <sup>[80]</sup>
Listeria seeligeri	NR	NR	NR	ATCC 35967 <sup>[80]</sup>
Micrococcus luteus	NR	NR	NR	MTCC 1538 <sup>[66]</sup>
Nocardia asteroides	NR	NR	NR	MTCC 274 <sup>[79]</sup>
Pasteurella multocida	NR	NR	NR	MTCC 1161 <sup>[78]</sup>
Erwinia carotovora	NR	NR	NR	MTCC 1428 <sup>[85]</sup>
Klebsiella aerogenes	NR	NR	NR	ATCC 9621 <sup>[80]</sup>
Alcaligenes faecalis	NR	NR	NR	ATCC8750 <sup>[87]</sup> MTCC 5521 <sup>[79]</sup>
Enterobacter cloacae	NR	NR	NR	ATCC 10699 <sup>[62]</sup>
Vibrio cholera	NR	NR	NR <sup>[77]</sup>	NR
Streptomyces aureofaciens	NR	NR	MTCC 325 <sup>[76]</sup>	NR

NR: Strain name not reported

**Table 3. Antifungal activities of Triphala, T. bellerica, T. chebula and E. officinalis**

Fungal Name	Plant Name (strain name) <sup>[reference]</sup>			
	Triphala	T. bellerica	T. chebula	E. officinalis
Aspergillus flavus	MTCC 277 <sup>[14]</sup>	MTCC 277 <sup>[82]</sup>	MTCC 277 <sup>[74]</sup>	NR
Aspergillus Fumigates	NR <sup>[94]</sup>	NR	MTCC3216 <sup>[97]</sup>	NR
Aspergillus niger	MTCC 282 <sup>[14]</sup>	NR	MTCC 282 <sup>[74]</sup>	MTCC 282 <sup>[82]</sup>
Aspergillus Terreus	NR	NR	NR	NR
Aspergillus Versicolor	NR	NR	NR	NR
Candida albicans	NR	NR	MTCC 183 <sup>[76]</sup> MTCC 227 <sup>[85]</sup> MTCC 3017 <sup>[73]</sup>	MTCC183 <sup>[94]</sup> MTCC 854 <sup>[91]</sup> ATCC 2091 <sup>[92]</sup> MTCC 183 <sup>[82]</sup>
Microsporum canis	NR	NR	NR	NR
Trichophyton rubrum	NR	NR	NR	NR
Candida glabrata	NR	MTCC 3019 <sup>[76]</sup>	MTCC 3019 <sup>[76]</sup>	NR
Cryptococcus neoformans	NR	MTCC 184 <sup>[76]</sup>	NR	NR
Candida	NR	NR	MTCC 184 <sup>[76]</sup>	MTCC 184 <sup>[91]</sup>

tropicalis				
Saccharomyces cerevisiae	NR	NR	MTCC170 <sup>[85]</sup>	NR

NR: Strain name not reported.

**Table 4. Antiprotozoal and antiviral activities of Triphala, T. bellerica, T. chebula and E. officinalis**

Activity Name	Plant Name <sup>[reference]</sup>			
	Triphala	T. bellerica	T. chebula	E. officinalis
<b>1. Antiprotozoal</b>				
Leishmanicidal	NR	<sup>[98]</sup>	NR	<sup>[99]</sup>
Plasmodial	NR	<sup>[100]</sup>	NR	<sup>[100]</sup>
Amoebic	NR	NR	<sup>[101,102]</sup>	NR
Helmintic	NR	NR	NR	<sup>[103,104]</sup>
<b>2. Antiviral</b>				
Influenza A virus	<sup>[105]</sup>	NR	NR	NR
Herpes simplex virus (HSV)	<sup>[106]</sup>	NR	NR	NR
Human cytomegalovirus	<sup>[107]</sup>	NR	NR	NR
HIV	NR	<sup>[108]</sup>	<sup>[109]</sup>	<sup>[110]</sup>
HBV	NR	<sup>[111]</sup>		NR
influenza A virus H3N8	NR	NR	<sup>[112]</sup>	NR
Encephalitis Virus	NR	NR	<sup>[113]</sup>	<sup>[113]</sup>
Human Enterovirus 71	NR	NR	<sup>[114]</sup>	NR
Hepatitis C virus	NR	NR	NR	<sup>[115]</sup>

NR: Antiprotozoal and antiviral activity not reported.

#### 4. Toxicology

While Triphala and its constituents have many health benefits, there are some of the potential concerns and side effects to be aware of while using it. The major precaution to be taken while using Triphala is that it shouldn't be taken by pregnant women as it is believed to favor miscarriage. Triphala can moderate blood sugar level, so patients of diabetes should take it under medical supervision. In case of an overdose, it can lead to dehydration which happens due to increased cleansing process of colon. Side effects are most common amongst first time users. EO is often recommended for pregnant women and lactating mothers in India. But, the other two ingredients of Triphala, TB and TC may be harmful during pregnancy. So, if someone still likes to keep the antioxidant intake high whilst pregnant, she may switch to powdered EO instead of Triphala. Interestingly, EO is also believed to enhance fertility in both men and women. Higher doses of TB, TC and EO may trigger healing crisis.

#### 5. Future consideration

Development of antimicrobial compounds from natural sources has great potential because they are easily available, economical and non-toxic in long term use. Since people consume

them in daily life, there is no need of clearance from regulatory authorities like the Food and Drug Administration (FDA) for their use in Ayurvedic formulations. Triphala and its constituents offers an inexpensive solution to more expensive modes of treatment for everyday issues like digestive and cold complaints, as well as for difficult conditions like microbial infection, inflammation including some forms of cancers. It is a natural, earth-friendly agricultural product which can be taken every day to improve health. In a world where general population reaches for digestive aids of all sorts, Triphala and its constituents still offers their benefits to those with less than healthy diets and lifestyles. Many *in vitro* antibacterial and antifungal studies of Triphala and its constituents have been reported. In comparison to antibacterial and antifungal activities, very few studies on antiviral and antiprotozoal activities are reported. As Triphala and its constituents are easily available and very cost effective, it may be a good source of phytomedicine against microbial infection, inflammation, cancer and other cardiovascular diseases. In future it can be explored further for other parameters such as inhibition of biofilm formation, microbial quorum sensing, and inhibition of microbial gene expression before recommending it for usage in routine clinical practice. Further *in vitro* as well as *in vivo* studies are required to explore the antiviral and antiprotozoal activities of TB, TC and EO. The methodology developed may be used for detection of new natural bioactive compounds.

## 6. CONCLUSION

Now a days, due to extensive use of antibiotics and vast majority of synthetic drugs, many multidrug resistant strains are developing specially in hospital environment. To overcome drug resistance and to avoid side effects associated with the commonly available antibiotics, there is need of an alternative treatment method to cure such infections by use of traditional medicinal herbs like TB, TC and EO which are potent antibacterial agents, clinically safer, economically cheaper and affordable. Present review builds a foundation for further *in vitro* and *in vivo* studies to understand the mechanism of antimicrobial action of Triphala and its constituents which may help in developing better therapeutic agents and healthy products.

### Conflict of interest

The authors declare that they have no conflict of interest.

**REFERENCES**

1. Liu Li, Oza S, Hogan D, et al. Global, regional, and national causes of child mortality in 2000–13, with projections to inform post-2015 priorities: an updated systematic analysis. *Lancet*, 2015; 385: 430-440.
2. Christopher Dye, John C. Reeder, Robert F. Terry, David B Evans, Justine Hsu, Ties Boerma., *Science Translational Medicine and Bulletin of the World Health Organization*, August, 2013.
3. Wayne W. LaMorte. *Transmission of infectious diseases*. Boston University School of Public Health, January, 2016.
4. National Institute of Allergy and Infectious diseases (NIAID). *Understanding microbes in Sickness and in Health*. U.S. department of health and human services. National Institutes of Health. NIH Publication No. 09-4914. September, 2009.
5. Janeway CA, Jr, Travers P, Walport M, et al. *Immunobiology: The Immune System in Health and Disease*. 5th edition. New York: Garland Science, 2001.
6. Mathur P and Singh S. Multidrug resistance in bacteria: A serious patient safety challenge for India. *J Lab Physicians*, 2013; 5: 5-10.
7. Bebarta VS, Ramirez S, Varney SM. Spice: A new “legal” herbal mixture abused by young active duty military personnel. *Substance Abuse*, 2012; 33: 191-194.
8. Benzie IFF and Wachtel-Galor S. *Herbal Medicine. Herbal Medicine: Biomolecular and Clinical Aspects*. 2nd edition. Boca Raton (FL): CRC Press/Taylor & Francis, 2011.
9. Awan UA, Andleeb S, Kiyani A, et al. Screening of traditional herbal plants and standard antibiotics against some human bacterial pathogens. *Pak J Pharm Sci.*, 2013; 26: 1109-1116.
10. Natural Products Foundation. Retrieved, 2013-12-07.
11. Baliga MS, Meera S, et al. Use of the Ayurvedic drug Triphala in medical conditions afflicting older adults. *foods and dietary supplements in the prevention and treatment of disease in older adults*. Academic Press, 2015; 398.
12. Duke JA, Bogenschutz-Godwin MJ, Duccellar J, Duke PAK. *Handbook of Medicinal Herbs*, second ed. CRC Press, Boca Raton, 2002; 70-71.
13. Thakur CP, Thakur B, Singh S, Sinha PK, Sinha SK. The Ayurvedic medicines, Haritaki, Amla and Bahira reduce cholesterol-induced atherosclerosis in rabbits. *Int. J Cardiology*, 1988; 21: 167–75.

14. Amala VE and Jeyaraj M. Determination of antibacterial, antifungal, bioactive constituents of Triphala by FT-IR and GC-MS analysis. *Int J Pharm Pharm Sci.*, 2014a; 6: 123-126.
15. Gupta R, Gupta A, Singh RL. Hepatoprotective Activities of Triphala and Its Constituents. *Int. J Pharma Res. & Rev.*, 2015; 4: 34-55.
16. Gupta R, Singh RL, Singh P. Quantification of Phytochemicals and Evaluation of Antioxidant Potential of Ethanolic Leaf Extract of Terminalia bellerica, Terminalia chebula and Emblica officinalis vis-a-vis Triphala. *Int. J. Pharm. Sci. Rev. Res.*, 2015; 32: 14-22.
17. Silva O, Serrano R. Terminalia genus as source of antimicrobial agents. The battle against microbial pathogens: basic science, technological advances and educational programs (A. Méndez-Vilas, Ed.), 2015: 236-245.
18. Mahboubi M and Ghazian Bidgoli F. In vitro synergistic efficacy of combination of amphotericin B with Myrtus communis essential oil against isolates of Candida albicans. *Phytomedicine*, 2010; 17: 771-774.
19. Sharma DK, Varshneya C, Mehta M. Total phenolic content and antioxidant activity of Triphala (an Ayurvedic formulation) and its constituents. *Am. j. PharmTech Res.*, 2012; 2: 458-465.
20. Ahmad I, Mehmood Z, Mohammad F. Screening of some Indian medicinal plants for their antimicrobial properties. *J of Ethnopharmacology*, 1998; 62: 183-193.
21. Mahalakshmi K, Prabhakar J, Sukumaran VG. Antibacterial activity of Triphala, GTP and curcumin on Enterococci faecalis. *Biomedicine*, 2006; 26: 43-46.
22. Sumathi P and Parvathi A. Antibacterial potential of the three medicinal fruits used in Triphala: An Ayurvedic formulation. *J. Med. Plants Res.*, 2010; 4: 1682-1685.
23. Amanullah S, Chandramoorthy HC, Kumar VK, Khatheeja S. Antimicrobial activity of Triphala against bacterial isolates from HIV infected patients. *Jundishapur J Microbiol.*, 2011; 4: 9-17.
24. Thomas B, Shetty SY, Shetty AV. Comparative Evaluation of Antimicrobial Activity of Triphala and Commercially Available Toothpaste: An in-vitro study. *International J Public Health Dent.*, 2011; 2: 8-12.
25. Kumar A, Kumar A, Kumar P, Patil S. Antibacterial activity of Triphala and Triphala Mashi extracts against bacteria isolates from wound infection. *Asian J Bioche & Pharma Res.*, 2011; 1: 142-148.



26. Pinmai K, Hiriotte W, Soonthornchareonnon N, Jongsakul K, Sireeratawong S, Tor-Udom S, 2010. In vitro and In vivo antiplasmodial activity and cytotoxicity of water extracts of *Phyllanthus emblica*, *Terminalia chebula*, and *Terminalia bellerica*. *J Med Assoc Thai.*, 2010; 93: 120-126.
27. Saraswathi MN, Karthikeyan M, Kannan M, Rajasekar S. *Terminalia bellerica* Roxb- A phytopharmacological review. *Int J Pharm Biomed Res.*, 2012; 3: 96-99.
28. The Ayurvedic Pharmacopeia of India. Government of India, Ministry of Health and Family Welfare, Department of Ayush. Part I, 5-8, 33-34: 62-63.
29. Sada TS. The Ayurveda Encyclopedia: Natural Secrets to Healing, Prevention and Longevity, 4<sup>th</sup> ed. Bayville, NY: Ayurveda Holistic Center, 2004: 78.
30. Vaidyaratnam PS. Varier's Indian medicinal plants. Oriental Longman Private Ltd. Chennai 2004; 5: 258-262.
31. Sabnis S. Antimicrobial efficacy of *Terminalia bellerica* against virulence factors of respiratory pathogens. *Int. J Curr. Microbiol & Appl Sci.*, 2014; 3: 215-221.
32. Valli S and Shankar SG. *Terminalia bellerica*-A promising challenge to cryptococcosis. *Int. J. Pharm. Biol. Sci.*, 2013; 2: 154-169.
33. Muhammad S, Khan BA, Akhtar N, Mahmood T, Rasul A, Hussain, et al. The morphology, extractions, chemical constituents and uses of *Terminalia chebula*: A review. *J. Med. Plants Res.*, 2012; 6: 4772-4775.
34. Mammen D, Bapat S, Sane R. An investigation to variation in constituents in the fruits of *Terminalia chebula* Retz. at different maturity stages. *Int. J. Pharm. Biol. Sci.*, 2012; 3: 416-419.
35. Kannan P, Ramadevi SA, Waheeta H. Antibacterial activity of *Terminalia chebula* fruit extract. *J Microbiol.*, 2009; 3: 180-184.
36. Kim HG, Cho JH, Jeong EY, Lim JH, Lee SH, Lee HS. Growth inhibitory activity of active component from *Terminalia chebula* fruits against intestinal bacteria. *J Food Prot.*, 2006; 69: 2205-2209.
37. Baliga MS and Dsouza JJ. Amla (*Emblica officinalis* Gaertn), a wonder berry in the treatment and prevention of cancer. *Eur J Cancer Prev*, 2011; 20: 225-239.
38. Patil SG, Deshmukh AA, Padol AR, Kale DB. In vitro antimicrobial activity of *Emblica officinalis* fruit by tube dilution method. *Toxicol Appl Pharmacol*, 2012; 2: 49-51.
39. Mishra P and Mahanta CL. Comparative analysis of functional and nutritive values of Amla fruit, seed and seed coat powder. *Am. J. Food Technol.*, 2014; 9: 151-161.

40. Son S and Lewis BA. Free radical scavenging and antioxidative activity of caffeic acid amide and ester analogues: structure-activity relationship. *J Agric Food Chem.* 2002; 50: 468-472.
41. Bosio K, Avanzini C, D'Avolio A, Ozino O, Savoia D. In vitro activity of propolis against *Streptococcus pyogenes*. *Lett Appl Microbiol.*, 2003; 31: 174-177.
42. Tim Cushnie TP and Lamb AJ. Antimicrobial activity of flavonoids. *Int J Antimicro Ag,* 2005; 26: 343–356.
43. Harborne JB and Williams CA. Advances in flavonoid research since 1992. *Phytochemistry*, 2000; 55: 481-504.
44. Li BQ, Fu T, Dongyan Y, Mikovits JA, Ruscetti FW, Wang JM. Flavonoid baicalin inhibits HIV-1 infection at the level of viral entry. *Biochem. Biophys. Res. Commun.*, 2000; 276: 534-538.
45. Li BQ, Fu T, Yan YD, Baylor NW, Ruscetti FW, Kung HF. Inhibition of HIV infection by baicalin-a flavonoid compound purified from Chinese herbal medicine. *Cell Mol Biol*, 1993; 39: 119-124.
46. Kumar A, Lakshman K, Jayaveera KN, Mani Tripathi SN, Satish KV. Estimation of Gallic Acid, Rutin and Quercetin in *Terminalia chebula* by HPTLC. *Jordan Journal of Pharmaceutical Sciences*, 2010; 3: 63-68.
47. Rathinamoorthy R and Thilagavathi G. *Terminalia Chebula*-Review on Pharmacological and Biochemical Studies. *Int J PharmTech Res.*, 2014; 6: 97-116.
48. Saxena V, Mishra G, Saxena A, Vishwakarma KK. A comparative study on quantitative estimation of tannins in *Terminalia Chebula*, *Terminalia Belerica*, *Terminalia Arjuna* and *Saraca indica* using spectrophotometer. *Asian J Pharm Clin Res.*, 2013; 6: 148-149.
49. Yang B and Liu P. Composition and biological activities of hydrolyzable tannins of fruits of *Phyllanthus emblica*. *J Agric Food Chem.*, 2014; 62: 529-541.
50. Gupta N, Saxena G, Kalra SS. Antimicrobial activity pattern of certain terpenoids. . *Int. J. Pharm. Biol. Sci.*, 2011; 2: 87-91.
51. Deepak P and Gopal GV. GC–MS Analysis of ethyl acetate extracts of *Phyllanthus emblica* L. bark. *British Biomedical Bulletin*, 2014; 2: 285-292.
52. Choi SH and Gu MB. Phenolic toxicity-detection and classification through the use of a recombinant bioluminescent *Escherichia coli*. *Environ Toxicol Chem*, 2001; 20: 248-255.
53. Vattem DA and Shetty K. Functional phytochemicals from cranberries: Their mechanism of action and strategies to improve functionality. In: *Food biotechnology*, 2<sup>nd</sup> Edition

- [Shetty, K., Paliyath, G., Pometto, A.L. III and Levin, R.E. (eds)]. Boca Raton, FA: CRC Press, 2005: 789-823.
54. Beate P, Samy K, Desouky EI, William E, Hull RH, Gerhard E, Robert, WO. Polyphenolic compounds in the fruits of Egyptian medicinal plants (*Terminalia bellerica*, *Terminalia chebula* and *Terminalia horrida*): characterization, quantitation and determination of antioxidant capacities. *Phytochemistry*, 2010; 71: 1132-1148.
  55. Hyams PJ, Simberkoff MS, Rahal JJ. In vitro bactericidal effectiveness of four aminoglycoside antibiotics. *Antimicrobial agents and chemotherapy*, 1973; 3: 87-94.
  56. Galimand M, Lambert T, Gerbaud G, Courvalin, P. Aminoglycoside resistance in the beta-hemolytic Group G *Streptococcus* isolate BM2721. *Antimicrobial agents and chemotherapy*, 1999; 47: 3008-3010.
  57. Schatzmann HJ. Herzglycoside als Hemmstoffe für den aktiven Kalium-und Natriumtransport durch die Erythrocytenmembran. *Helvetica Physiol Pharmacol Acta*, Basel., 1953; 11: 346-354.
  58. Chen X, Sun F, Ma L, Wang J, Qin H, Du G, In vitro evaluation on the antioxidant capacity of triethylchebulate, an aglycone from *Terminalia chebula* Retz fruit. *Indian Journal of Pharmacology*, 2011; 43: 320-323.
  59. Sharma S. Chemical investigations of *Terminalia Bellerica*. *Acta Chimica & Pharmaceutica Indica*, 2012; 2: 132-133.
  60. El-Desouky SK, Ryu S, Kim YK. A new cytotoxic acylated apigenin glucoside from *Phyllanthus emblica* L. *Natural Product Research*, 2008; 28: 91-95.
  61. Safiullah A, Harish CC, Vijay Anand, K, Saira K. Antimicrobial activity of *Triphala* against bacterial isolates from HIV infected patients. *Jundishapur J Microbiol.*, 2011; 4:9-17.
  62. Alam MB, Zahan R, Hasan M, Khan, et al. Thank You, a good research antioxidant, antimicrobial and toxicity studies of the different fractions of fruits of *Terminalia belerica* Roxb. *Global Journal of Pharmacology*, 2011; 5: 7-17.
  63. Chanda S, Menpara D, Desai D. Antimicrobial activity of *Terminalia bellerica* leaf and stem collected from two different sites. *American Journal of Phytomedicine and Clinical Therapeutics*, 2013; 1: 721-733.
  64. Koppula US and Koppula S. Antibacterial activity of some selected Indian medicinal plant barks. *Int. J Bio-Pharma Res.*, 2012; 1: 30-33.

65. Jayalakshmi B, Raveesha KA, Amruthesh KN. Phytochemical investigations and antibacterial activity of some medicinal plants against pathogenic bacteria. *J App Pharm Sci.*, 2011; 1: 124-128.
66. Jyothi SK and Rao BS. Screening of antibacterial activity of *Embllica officialis* fruits. *Pharmacologyonline*, 2011; 3: 848-852.
67. Bag A, Bhattacharyya SK, Pal NK, Chattopadhyay RR. Antibacterial potential of hydroalcoholic extracts of *Triphala* components against multidrug resistant uropathogenic bacteria-A preliminary report. *Indian J Exp Biol*, 2013b; 51: 709-714.
68. Elizabeth KM. Antimicrobial activity of *Terminalia Bellerica*. *Indian J Clin Biochem.* 2005; 20: 150-153
69. Dharmaratne MPJ, Manoraj A, Thevanesam V, Bandara BMR, Ekanayake EWMA, Kumar NS. Antibacterial activity of aqueous extracts of *Terminalia chebula* fruit against some multidrug resistant human pathogens. *Int J Pharm Biomed Res.*, 2013; 4: 1333-1337.
70. Rathinamoorthy R, Udayakumar S, Thilagavathi G. Antimicrobial efficacy of *Terminalia chebula* fruit extract treated cotton fabric for healthcare applications. *Int. J Pharma. Sci. and Nanotechnology*, 2012; 4: 1549-1555.
71. Shah CP, Vachhani UD, Trivedi MN, Joshi VJ, Santani DD. Evaluation of antibacterial potential of *Terminalia chebula* against pathogenic organisms of lacrimal system. *Pharmacologyonline*, 2011; 1: 506-509.
72. Sharma C, Aneja KR, Kasera R, Aneja A. Antimicrobial potential of *Terminalia chebula* Retz. Fruit extracts against ear pathogens. *World J Otorhinolaryngology*, 2012a; 2: 8-13.
73. Jinukuti MC and Giri A. Antimicrobial activity of aqueous extract of *Terminalia chebula* Retz. *Recent res. sci. technol.*, 2013; 189(5): 46-49.
74. Singh G and Kumar P. Evaluation of antimicrobial activity of alkaloids of *Terminalia chebula* Retz. Against some multidrug-resistant microorganisms. *Int J Green Phar*, 2012; 6: 57-62.
75. Tambekar DH, Khante BS, Dahikar SB, Zarey VM. Antibacterial properties of contents of *Triphala*: A traditional Indian herbal preparation. *Con. J Microb*, 2007; 1: 8-12.
76. Vats S, Kumar R, Negi S. Natural food that meets antibiotics resistance challenge: In vitro synergistic antimicrobial activity of *Azadirachta indica*, *Terminalia chebula*, *Piper nigrum* and photoactivated cow urine. *Asian j. pharm. biol Res.*, 2012; 2: 122-126.

77. Malekzadeh F, Ehsanifar H, Shahamat M, Levin M, Colwell RR. Antibacterial activity of black myrobalan (*Terminalia chebula* Retz) against *Helicobacter pylori*. *International Int. J. Antimicrob. Agents*, 2001; 18: 85–88.
78. Kumar A, Tantry BA, Rahiman S, Gupta U. Comparative study of antimicrobial activity and phytochemical analysis of methanolic and aqueous extracts of the fruit of *Emblica Officinalis* against pathogenic bacteria. *J Tradit Chin Med.*, 2011; 31: 246-250.
79. Patil SG, Deshmukh AA, Padol AR, Kadam RG, Kale BD. Antibacterial efficacy study of *Emblica officinalis* against experimentally induced *Escherichia coli* infection in broiler chicks. *International Journal of Pharmacology & Toxicology Science*, 2013; 3: 39-49.
80. Kapoor S. In vitro Antimicrobial and antioxidant potential of aqueous extract of common medicinal plants. *Biotechnology*, 2014; 4: 10-11.
81. Sharma S, Bharose R, Agarawal SK, Pal K. Antibacterial activities against different selective MIC extraction of Amla. *International Journal of Pharma Professional's Res.*, 2012b; 3: 680-681.
82. Amala VE and Jeyaraj M. Determination of antibacterial and antifungal activity of *Terminalia Bellirica* Roxb and *Phyllanthus Emblica* L. against Some pathogenic microorganism. *Biometric, Functional Group Analysis*, 2014b; 3: 531-533.
83. Kumar MS, Kirubanandan S, Sripriya R, Sehgal PK. Triphala promotes healing of infected full-thickness dermal wound. *Journal of Surgical Research*, 2008; 144: 94-101.
84. Kirubanandan S, Swethkamal K, Renganathan S. Activities of Triphala towards promoting collagen synthesis at wound site and inhibiting methicillin-resistant *Staphylococcus aureus* and its enzymes. *Int J Pharm Pharm Sci.*, 2013; 5: 54-62.
85. Aneja KR and Joshi R. Evaluation of antimicrobial properties of fruit extracts of *Terminalia chebula* against dental caries pathogens. *Jundishapur J Microb.*, 2009b; 2: 105-111
86. Gowd P, Kumar M, Shankar S, Sreedevi S. Evaluation of three medicinal plants for antimicrobial activity. *Int J Res Ayurveda Pharm*, 2014; 33: 423-427.
87. Rathish NAIR and Chanda SV. Antibacterial activities of some medicinal plants of the western region of India. *Turkish Journal of Biology*, 2007; 31: 231-236.
88. Shah S, Trivedi B, Patel J, Dave JH, Sathvara N, Shah V. Evaluation and comparison of antimicrobial activity of Tulsi (*Ocimum Sanctum*), Neem (*Azadirachta Indica*) and Triphala extract against *Streptococcus Mutans* & *Lactobacillus Acidophilus*: An In vitro Study. *Natl J Integr Res Med*, 2014; 5: 17-21.

89. Prajapati RA and Rao BV. The study on the efficacy of some herbal extracts for the control of dental caries pathogen-Streptococcus mutans. *International Journal of Pharmaceutical Science and Health Care*, 2014; 1: 49-58.
90. Chaiya A, Saraya S, Chuakul W, Tamsiririrkkul R. Screening for dental caries: preventive activities of medicinal plants against Streptococcus mutans. *Mahidol University Journal of Pharmaceutical Sciences*, 2013; 40: 9-17.
91. Yadav S, Singh S, Sharma P, Thapliyal A, Gupta V. Antibiofilm formation activity of Terminalia bellericaplant extract against clinical isolates of Streptococcus mutans and Streptococcus sobrinus: Implication in Oral Hygiene. *Int. j. pharm. biol. sci. arch.*, 2012; 3: 816-821.
92. Gauniyal P and Teotia UVS. Phytochemical screening and antimicrobial activity of some medicinal plants against oral flora. *Asian Pacific Journal of Health Sciences*, 2014; 1: 255-263.
93. Potdar S, Lakshminarayan N, Goud RS. Antimicrobial Efficacy of Emblica Officinalis Fruit Extracts on S. Mutans, E. Faecalis and C. Albicans. *Adv Hum Biol*, 2014; 4: 26–30.
94. Pujar M, Patil C, Kadam A. Comparison of antimicrobial efficacy of Triphala, (GTP) Green tea polyphenols and 3% of sodium hypochlorite on Enterococcus faecalis biofilms formed on tooth substrate: in vitro. *J Int Oral Health*, 2011; 3: 24-30.
95. Kavitha HU and Satish S. Eco-friendly management of plant pathogens by some medicinal plant extracts. *J. Agr. Sci. Tech.*, 2011; 7: 449-461.
96. Srikumar R, Devi RS, Ayyappan, SR, Thangaraj R, Jegadeesh R, Hariprasath L. Antifungal activity of aqueous and ethanolic extract of Triphala and its individual fruit components. *Int J Periodontics Restorative Dent*, 2010; 2: 64-66.
97. Takahashi M, Fuchino H, Satake M, Agatsuma Y, Sekita S. In vitro Screening of leishmanicidal activity in myanmar timber extracts. *Biol Pharm Bull*, 2004; 27: 921-925.
98. Kaur S, Kaur G, Sachdeva H, Kaur J. In vivo evaluation of the antileishmanial activity of two immunomodulatory plants, Emblica officinalis and Azadirachta Indica in BALB/C mice. *International journal of Ayurvedic and herbal medicine*, 2013; 3: 1066-1079.
99. Bagavan A, Rahuman AA, Kamraj C, Kaushik NK, Mohanakrishnana D, Sahal D. Antiplasmodial activity of botanical extract against Plasmodium falciparum. *Parasitology Research*, 2011; 108: 1099-1109.
100. Pinmai K, Hiriotte W, Soonthornchareonnon N, Jongsakul K, Sireeratawong S, Tor-Udom S, 2010. In vitro and In vivo antiplasmodial activity and cytotoxicity of water extracts of

- Phyllanthus emblica, Terminalia chebula, and Terminalia bellerica. J Med Assoc Thai., 2010; 93: 120-126.
101. Sohni YR, Kalmal P, Bhatt RM. The antiamebic effect of a crude drug formulation of herbal extracts against Entamoeba histolytica in vitro and in vivo. Journal of Ethanopharmacology, 1995; 45: 43-52.
  102. Rani D. Plant Extracts with Antiamoebic Properties: A Theoretical Study with reference to Entamoeba histolytica. Int J PharmTech Res, 2011; 3: 1113-1117.
  103. Dwivedi S, Dwivedi A, Kapadia R, Kaul S. Anthelmintic activity of alcoholic and aqueous extract of fruits of Terminalia chebula Retz. Ethnobotanical Leaflets, 2008; 12: 741-743.
  104. Kamaraj C and Rahuman AA. Efficacy of anthelmintic properties of medicinal plant extracts against Haemonchus contortus. Research in Veterinary Science, 2011; 91: 400-404.
  105. Badmaev V and Nowakowski M. Protection of epithelial cells against influenza A virus by a plant derived biological response modifier Ledretan-96. Phytotherapy Research, 2000; 14: 245-249.
  106. Yukawa TA, Kurokawa M, Sato H, et al. Prophylactic treatment of cytomegalovirus infection with traditional herbs. Antiviral Research, 1996; 32: 63-70.
  107. Shiraki K, Yukawa T, Kurokawa M, Kageyama S. Cytomegalovirus infection and its possible treatment with herbal medicines. Nippon rinsho, 1998; 56: 156-160.
  108. Valsaraj R, Pushpangadan P, Smitt UW, et al. New anti-HIV-1, antimalarial, and antifungal compounds from Terminalia bellerica. J. Nat. Pro, 1997; 60: 739-742.
  109. Theepireddy SKR, Chinthala R. Rao LV, Vishwakarma S. Reddy KR, Duvvada MR. Anti-HIV activity of Terminalia chebula crude extracts and gallic acid. International Journal of Pharmacy and Technology, 2014; 6: 6346-6354.
  110. Estari M, Venkanna L, Sripriya D, Lalitha R. Human Immunodeficiency Virus (HIV-1) reverse transcriptase inhibitory activity of Phyllanthus emblica plant extract. Biology and Medicine, 2012; 4: 178-182.
  111. Anbalagan S., Sankareswaran M., Rajendran P, Karthikeyan M, Priyadharshini K, Hamza HB. In vitro screening of anti-HBV properties of selected Indian medicinal plants from Kolli hills, Namakkal district of Tamilnadu, India. World journal of pharmacy and pharmaceutical sciences, 2015; 4: 909-915.

112. Oyuntsetseg N, Khasnatinov MA, Molor-Erdene P, Oyunbileg J, et al. Evaluation of direct antiviral activity of the Deva-5 herb formulation and extracts of five Asian plants against influenza A virus H3N8. *BMC Complement Altern Med*, 2014; 14: 226-235.
113. Singh VK, George CX, Gupta KP, Gupta BM. Antiviral activity of plant extract Liv.52 in mice experimentally infected with semliki forest encephalitis virus. *Science and Culture*, 1983; 49: 351-354.
114. Yang Y, Xiu J, Liu J, Zhang Li, Li X, Xu Y, Qin C, Zhan L. Chebulagic Acid, a hydrolyzable tannin, exhibited antiviral activity in vitro and in vivo against human Enterovirus 71. *Int. J. Mol. Sci.*, 2013; 14: 9618-9627.
115. Ravikumar YS, Ray U, Nandhitha, et al. Inhibition of Hepatitis C Virus replication by herbal extract: *Phyllanthus amarus* as potent natural source. *Virus Research*, 2011; 158:89-97.